

Common variants in or near *ZNRF1*, *COLEC12*, *SCYL1BP1* and *API5* are associated with diabetic retinopathy in Chinese patients with type 2 diabetes

Danfeng Peng¹ · Jie Wang¹ · Rong Zhang¹ · Feng Jiang¹ · Shanshan Tang¹ · Miao Chen¹ · Jing Yan¹ · Xue Sun¹ · Shiyun Wang¹ · Tao Wang¹ · Dandan Yan¹ · Yuqian Bao¹ · Cheng Hu^{1,2} · Weiping Jia¹

Received: 10 February 2015 / Accepted: 5 March 2015 / Published online: 28 March 2015
© Springer-Verlag Berlin Heidelberg 2015

Abstract

Aims/hypothesis Three recent genome-wide association studies (GWAS) identified several single-nucleotide polymorphisms (SNPs) with modest effects on diabetic retinopathy in Mexican-American and white patients with diabetes. This study aimed to evaluate the effects of these variants on diabetic retinopathy in Chinese patients with type 2 diabetes.

Methods A total of 1,972 patients with type 2 diabetes were recruited to this study, including 819 patients with diabetic retinopathy and 1,153 patients with diabetes of ≥ 5 years duration but without retinopathy. Forty SNPs associated with diabetic retinopathy in three GWAS were genotyped. Fundus photography was performed to diagnose and classify diabetic retinopathy.

Results rs17684886 in *ZNRF1* and rs599019 near *COLEC12* were associated with diabetic retinopathy (OR 0.812, $p=0.0039$ and OR 0.835, $p=0.0116$, respectively) and with the severity of diabetic retinopathy ($p=0.0365$ and $p=0.0252$, respectively, for trend analysis). Sub-analysis in patients with diabetic retinopathy revealed that rs6427247 near *SCYL1BP1*

(also known as *GORAB*) and rs899036 near *API5* were associated with severe diabetic retinopathy (OR 1.368, $p=0.0333$ and OR 0.340, $p=0.0005$, respectively). The associations between rs6427247 and rs899036 and severe diabetic retinopathy became more evident after a meta-analysis of published GWAS data (OR 1.577, $p=2.01 \times 10^{-4}$ for rs6427247; OR 0.330, $p=5.84 \times 10^{-7}$ for rs899036).

Conclusions/interpretation We determined that rs17684886 and rs599019 are associated with diabetic retinopathy and that rs6427247 and rs899036 are associated with severe diabetic retinopathy in Chinese patients with type 2 diabetes.

Keywords Diabetic retinopathy · Polymorphisms · Type 2 diabetes

Abbreviations

AER	Albumin excretion rate
EDIC	Epidemiology of Diabetes Intervention and Control Trial
GoKinD	Genetics of Kidney in Diabetes
GWAS	Genome-wide association study
NPDR	Non-proliferative diabetic retinopathy
PDR	Proliferative diabetic retinopathy
SNP	Single-nucleotide polymorphism
WESDR	Wisconsin Epidemiologic Study of Diabetic Retinopathy

Danfeng Peng and Jie Wang contributed equally to this work.

✉ Cheng Hu
alfredhc@sjtu.edu.cn

✉ Weiping Jia
wpjia@sjtu.edu.cn

¹ Shanghai Diabetes Institute, Shanghai Key Laboratory of Diabetes Mellitus, Shanghai Clinical Center for Diabetes, Shanghai Jiao Tong University Affiliated Sixth People's Hospital, 600 Yishan Road, Shanghai 200233, People's Republic of China

² Institute for Metabolic Diseases, Shanghai Jiao Tong University Affiliated Sixth People's Hospital South Campus, Shanghai, People's Republic of China

Introduction

Diabetes mellitus, particularly type 2 diabetes, has reached epidemic proportions in China and other countries. As a major chronic microvascular complication of diabetes, diabetic

retinopathy is the leading cause of vision loss among working-age adults around the world [1]. The increasing prevalence of diabetes has led to an increase in the number of patients suffering from diabetic retinopathy, which represents a heavy public health burden. A meta-analysis has indicated that the global prevalence of diabetic retinopathy among patients with diabetes is 34.6% [2]. There are approximately 93 million people with diabetic retinopathy, 17 million with proliferative diabetic retinopathy (PDR) and 28 million with vision-threatening diabetic retinopathy.

Diabetic retinopathy is a multifactorial disease. Epidemiological and prospective studies have demonstrated that the duration of diabetes, poor glycaemic control and blood pressure are major risk factors for diabetic retinopathy [3–5]. Some patients with intensive glycaemic control and a shorter duration of diabetes may still develop diabetic retinopathy, whereas some are spared despite poor glycaemic control and a longer duration of diabetes [6]. These phenomena may be explained by the genetic contributions to diabetic retinopathy. Studies have suggested a genetic influence on the development of diabetic retinopathy [7]. One study found the same degree of severity in high concordance in twins with diabetes [8]. A familial aggregation of diabetic retinopathy has been observed across different ethnicities. Siblings and relatives of patients with diabetic retinopathy have a significantly increased risk of diabetic retinopathy compared with siblings and relatives of patients with diabetes but without diabetic retinopathy [9–11]. This trend is even more pronounced in families exhibiting more severe diabetic retinopathy [12–14]. However, the study of the genetics of diabetic retinopathy is still in its infancy, and attempts to identify susceptible loci have been unsuccessful.

Most of the genetic research into diabetic retinopathy has involved a candidate gene approach. A significant number of genes and genetic variants has been proposed through this approach (e.g. *AKR1B1*, *VEGFA*, *ACE* and *AGER*) [15]. However, individual studies have frequently yielded inconsistent and even conflicting findings. Since 2005, genome-wide association studies (GWAS) have been widely used for complex diseases, including diabetic retinopathy but, to date, no locus for diabetic retinopathy from GWAS reaches conventional significance criteria. In 2010, a GWAS among Mexican-American families identified several single-nucleotide polymorphisms (SNPs) and genes associated with severe diabetic retinopathy at a p value of less than 0.0001 [16]. Another genome-wide meta-analysis using Genetics of Kidney in Diabetes (GoKinD) and Epidemiology of Diabetes Intervention and Control Trial (EDIC) samples identified several SNPs at close to a genome-wide level for severe diabetic retinopathy (rs476141, $p=1.2\times 10^{-7}$; rs10521145, $p=3.4\times 10^{-6}$) [17]. A replication analysis for severe diabetic retinopathy was conducted in a cohort of diabetic individuals from the Wisconsin Epidemiologic Study of Diabetic Retinopathy (WESDR) [18].

The top two associations observed were for rs4865047 ($p=2.06\times 10^{-5}$) and rs1902491 ($p=2.81\times 10^{-5}$). Because the effects of these loci on diabetic retinopathy in the Chinese population are unknown, our study attempted to replicate the associations observed in these three studies in a cohort of Chinese patients with type 2 diabetes in Shanghai.

Methods

Participants This study involved 1,972 patients with type 2 diabetes recruited from the Shanghai Diabetic Complications Study [19] and Shanghai Diabetes Institute Inpatient Database of Shanghai Jiao Tong University Affiliated Sixth People's Hospital [20, 21]. All participants were unrelated patients with type 2 diabetes meeting the 1999 WHO criteria. Of these patients, 819 were diagnosed with diabetic retinopathy and 1,153 had diabetes for longer than 5 years but without diabetic retinopathy and were considered as cases and controls for diabetic retinopathy, respectively. This study was approved by the institutional review board of Shanghai Jiao Tong University Affiliated Sixth People's Hospital, and written informed consent was obtained from each participant.

Clinical measurements Fundus photography of all participants was performed according to a standardised protocol at the Department of Ophthalmology, Shanghai Jiao Tong University Affiliated Sixth People's Hospital. Both eyes of each patient were photographed with a 45° 6.3 megapixel digital nonmydriatic camera (Canon CR6-45NM; Lake Success, NY, USA). Retinopathy was graded according to the International Classification of Diabetic Retinopathy [22] as follows: mild non-proliferative diabetic retinopathy (NPDR), moderate NPDR, severe NPDR or PDR. The more severely affected eye of each patient was used to classify their retinopathy status. The 24 h albumin excretion rate (AER) was used to assess nephropathy. AER was measured on three consecutive days, and the mean value was recorded for each patient. Patients with $AER\geq 30$ mg/24 h were diagnosed with diabetic nephropathy. To evaluate glycaemic control the HbA_{1c} level was measured by high-performance liquid chromatography (Variant II; Bio-Rad, Hercules, CA, USA). Blood pressure and lipid profile data were also collected for each participant.

SNP selection, genotyping and quality control A total of 40 SNPs, including 21 SNPs reported in the Mexican-American GWAS (rs10501943, rs10519765, rs1197310, rs1445754, rs2300782, rs6427247, rs699549, rs763970, rs899036, rs599019, rs1106412, rs1033465, rs11583330, rs11635920, rs11812882, rs11927173, rs17083119, rs3014267, rs3098241, rs6726798 and rs6909083), 17 SNPs reported in the GoKinD and EDIC GWAS (rs476141, rs10521145, rs17670074, rs227455, rs238252, rs737141, rs11074904,

rs11647881, rs11736136, rs151227, rs151320, rs17063155, rs17684886, rs238250, rs4941432, rs9888035 and rs11871508) and 2 SNPs reported in the WESDR replication study (rs4865047 and rs1902491), were genotyped in all participants. Genotyping was performed by primer extension of multiplex products with detection by matrix-assisted laser desorption ionisation–time of flight mass spectroscopy using a MassARRAY Compact Analyzer (Sequenom, San Diego, CA, USA). rs11871508 and rs6909083 failed during the assay design and genotyping, respectively. The genotyping data underwent a series of quality control checks and cleaned data were used in further statistical analyses. The call rate for each SNP was more than 96%. The concordance rate based on 100 duplicates was greater than 99% for all SNPs. Eighty-three individuals were excluded due to sample call rate <80%. The Hardy–Weinberg equilibrium test was performed before the statistical analysis (a two-tailed p value <0.01 was considered statistically significant), and rs238250 was excluded ($p=0.004$). Another 12 SNPs (rs11583330, rs6726798, rs1106412, rs11812882, rs17063155, rs238252, rs151227, rs151320, rs10521145, rs11647881, rs11074904 and rs737141) were rare in our population (minor allele frequency <0.0015) and were excluded from statistical analyses.

Statistical analysis The allelic frequencies were compared between patients with or without diabetic retinopathy using a χ^2 test in PLINK (v1.07; <http://pngu.mgh.harvard.edu/~purcell/plink/>) [23], and ORs with 95% CIs are presented. Genotype distributions between patients with or without diabetic retinopathy were compared using logistic regression under an additive model with adjustment of confounding factors. The effects of SNPs on the levels of retinopathy severity were

analysed by trend analysis. Combined ORs from different studies were calculated by Comprehensive Meta Analysis (v2.2.057; Englewood, NJ, USA) with a fixed- or random-effect model after testing for heterogeneity. The test for homogeneity was assessed by the Cochran Q test. Correction for multiple testing was performed using PLINK through 10,000 permutation tests and empirical p values are presented. The statistical analyses were performed using SAS 9.3 (SAS institute, Cary, NC, USA) unless specified otherwise. A two-tailed p value <0.05 was considered statistically significant.

On the basis of an estimated effect size of genetic loci for diabetic retinopathy (~1.25), our samples had >85% power to detect an effect SNP with minor allele frequency of 0.3 and >75% power to detect an effect SNP with minor allele frequency of 0.2 at a level of significance of 0.05.

Results

The clinical characteristics of the samples that passed genotype quality control are shown in Table 1. Compared with patients without diabetic retinopathy, patients with diabetic retinopathy were younger, diagnosed with diabetes at earlier age, and had higher HbA_{1c} and blood pressure and higher prevalence of nephropathy.

We first analysed the association between SNPs and the risk of diabetic retinopathy. As shown in Table 2, rs1197310, rs4865047, rs17684886 and rs599019 were nominally associated with diabetic retinopathy (OR 1.155, 95% CI 1.013, 1.316, $p=0.0309$ for the rs1197310 T allele, OR 0.854, 95% CI 0.735, 0.991, $p=0.0379$ for the rs4865047 T allele, OR 0.829, 95% CI 0.729, 0.944, $p=0.0046$ for the

Table 1 Clinical characteristics of the samples

Characteristic	Patients with diabetic retinopathy	Patients without retinopathy	p value
Samples (n)	789	1,110	
Mild NPDR	490		
Moderate NPDR	136		
Severe NPDR	118		
PDR	45		
Sex, male/female (n)	370/419	494/616	
Age (years)	62.18±11.03	64.06±10.95	0.0001
BMI (kg/m ²)	24.58±7.46	24.40±3.50	0.51
Age at diagnosis of diabetes (years)	51.40±10.86	53.32±10.65	<0.0001
Duration of diabetes (years)	10 (5–15)	10 (7–13)	0.79
HbA _{1c} (%)	9.11±2.27	8.36±2.06	<0.0001
HbA _{1c} (mmol/mol)	76.11±24.90	67.91±22.42	<0.0001
Systolic blood pressure (mmHg)	137.74±19.14	134.53±17.66	0.0002
Diastolic blood pressure (mmHg)	81.34±9.71	80.30±9.30	0.0282
Percentage with nephropathy (%)	37.4	26.6	<0.0001

The data are n , mean±SD or median (interquartile range)

Table 2 Associations of SNPs with diabetic retinopathy

Chromosome	SNP	Position (Build 38)	Nearest gene	Minor/major allele	Risk allele	Minor allele frequency	Allelic OR (95% CI)	p value (empirical p value ^a)	Genotypic OR (95% CI) ^b	p value (empirical p value ^{a,b})	
											Cases
1	rs6427247	170411339	SCYL1BP1 (GORAB)	G/A	A	0.310	0.312	0.994 (0.864, 1.143)	0.93	0.974 (0.837, 1.134)	0.74
1	rs1033465	173018590	TNFSF18	T/A	T	0.022	0.018	1.170 (0.739, 1.853)	0.50	1.319 (0.813, 2.139)	0.26
1	rs3014267	227365216	CDC42BP4	G/A	A	0.238	0.249	0.939 (0.808, 1.092)	0.42	0.922 (0.784, 1.085)	0.33
1	rs476141	244013122	LOC339529	A/C	A	0.214	0.210	1.027 (0.877, 1.203)	0.74	1.000 (0.844, 1.185)	1.00
2	rs699549	4657673	LINC01249	T/C	C	0.246	0.257	0.945 (0.813, 1.097)	0.46	0.928 (0.790, 1.091)	0.37
2	rs763970	137878563	HNMT	A/C	A	0.260	0.254	1.033 (0.890, 1.198)	0.67	1.062 (0.905, 1.245)	0.46
3	rs11927173	23183703	UBE2E2	C/T	T	0.186	0.192	0.962 (0.815, 1.134)	0.64	0.985 (0.825, 1.176)	0.87
3	rs1197310	133409380	BFSF2	T/A	T	0.508	0.472	1.155 (1.013, 1.316)	0.0309 (0.53)	1.086 (0.945, 1.247)	0.24
4	rs4865047	55955640	CEP135	T/C	C	0.238	0.268	0.854 (0.735, 0.991)	0.0379 (0.61)	0.909 (0.772, 1.072)	0.26
4	rs11736136	82097121	LOC101928987	G/A	G	0.066	0.060	1.110 (0.851, 1.449)	0.44	1.145 (0.855, 1.534)	0.36
4	rs1902491	155134181	NPY2R	G/T	T	0.154	0.157	0.979 (0.819, 1.170)	0.81	1.020 (0.842, 1.237)	0.84
5	rs1445754	84279813	EDIL3	A/T	A	0.056	0.052	1.100 (0.828, 1.463)	0.51	1.119 (0.823, 1.522)	0.47
5	rs2300782	111453087	CAMK4	T/C	T	0.476	0.469	1.030 (0.905, 1.173)	0.66	1.018 (0.886, 1.169)	0.81
6	rs17083119	121080964	TBC1D32	G/A	A	0.096	0.098	0.978 (0.786, 1.216)	0.84	0.974 (0.772, 1.230)	0.83
6	rs227455	165064562	C6orf118	C/T	T	0.466	0.476	0.963 (0.846, 1.096)	0.57	0.971 (0.845, 1.117)	0.68
8	rs3098241	103413076	SLC25A32	G/A	G	0.366	0.343	1.108 (0.968, 1.268)	0.14	1.126 (0.970, 1.307)	0.12
10	rs17670074	19416454	MALRD1	C/A	A	0.367	0.372	0.982 (0.859, 1.123)	0.79	1.086 (0.937, 1.260)	0.27
10	rs9888035	19426089	MALRD1	C/T	C	0.010	0.007	1.500 (0.739, 3.042)	0.26	1.484 (0.705, 3.124)	0.30
11	rs899036	41661360	API5	C/A	A	0.102	0.105	0.968 (0.783, 1.196)	0.76	0.936 (0.743, 1.180)	0.58
11	rs10501943	100076267	CNTN5	C/T	C	0.041	0.039	1.045 (0.751, 1.454)	0.79	0.828 (0.571, 1.199)	0.32
13	rs4941432	42524211	TNFSF11	A/G	G	0.171	0.189	0.888 (0.750, 1.052)	0.17	0.868 (0.725, 1.040)	0.13
15	rs10519765	32913223	FMN1	A/G	G	0.080	0.083	0.968 (0.764, 1.225)	0.78	0.905 (0.698, 1.172)	0.45
15	rs11635920	32920456	FMN1	A/T	T	0.081	0.082	0.989 (0.781, 1.251)	0.92	0.928 (0.717, 1.201)	0.57
16	rs17684886	75052977	ZNRF1	A/T	T	0.460	0.507	0.829 (0.729, 0.944)	0.0046 (0.11)	0.812 (0.705, 0.935)	0.0039 (0.0783)
18	rs599019	294495	COLEC12	C/A	A	0.385	0.430	0.832 (0.729, 0.950)	0.0065 (0.15)	0.835 (0.725, 0.961)	0.0116 (0.24)

The OR with 95% CI shown is for the minor allele

^a Empirical *p* values are based on 10,000 permutations

^b Adjusted for duration of diabetes, HbA_{1c}, systolic blood pressure, diastolic blood pressure, BMI and nephropathy under an additive model

rs17684886 A allele and OR 0.832, 95% CI 0.729, 0.950, $p=0.0065$ for rs599019 C allele). After adjusting for confounding factors, including the duration of diabetes, HbA_{1c} level, systolic and diastolic blood pressure, BMI and nephropathy, rs17684886 and rs599019 still exhibited significant association with diabetic retinopathy (OR 0.812, 95% CI 0.705, 0.935, $p=0.0039$, empirical $p=0.0783$ for rs17684886 and OR 0.835, 95% CI 0.725, 0.961, $p=0.0116$, empirical $p=0.24$ for rs599019). We then analysed the effects of rs17684886 or rs599019 on the severity of diabetic retinopathy in all samples. As shown in Table 3, both SNPs were associated with the level of diabetic retinopathy, with the risk allele more frequent in patients with more severe diabetic retinopathy ($p=0.0365$ for rs17684886 and $p=0.0252$ for rs599019 for trend analysis).

Because the 40 SNPs were reported to be possibly associated with severe diabetic retinopathy, we examined this relationship in patients with diabetic retinopathy. For severe diabetic retinopathy analysis, patients with mild NPDR ($n=485$) were regarded as controls and patients with severe NPDR or PDR were regarded as cases ($n=163$). As shown in Table 4, rs2300782 and rs899036 were significantly associated with severe diabetic retinopathy (OR 1.380, 95% CI 1.073, 1.777, $p=0.0121$ for the rs2300782 T allele and OR 0.448, 95% CI 0.268, 0.749, $p=0.0017$ for the rs899036 C allele). rs6427247 and rs10501943 were marginally associated with severe diabetic retinopathy ($p=0.0525$ for rs6427247 and $p=0.0516$ for rs10501943). After adjusting for the duration of diabetes, HbA_{1c} level, systolic and diastolic blood pressure, BMI and nephropathy, rs6427247 and rs899036 were still associated with severe diabetic retinopathy (OR 1.368, 95% CI 1.025, 1.825, $p=0.0333$ for rs6427247 and OR 0.340, 95% CI 0.185, 0.624, $p=0.0005$ for rs899036). The association between rs899036 and severe diabetic retinopathy remained significant even after adjusting for multiple comparisons (empirical $p=0.0093$).

We also performed a meta-analysis with the fixed-effect model combining our data and the published GWAS data for rs6427247 and rs899036 [16]. As shown in Table 5, the associations of these two SNPs with

severe diabetic retinopathy became more evident. rs899036 showed an association with severe diabetic retinopathy that approached genome-wide significance (OR 0.330, 95% CI 0.214, 0.510, $p=5.84 \times 10^{-7}$).

Discussion

In this study, we analysed the effects of 40 SNPs reported by GWAS on diabetic retinopathy in Chinese patients with type 2 diabetes. We determined that rs17684886 and rs599019 were associated with diabetic retinopathy in Chinese patients with type 2 diabetes after adjusting for confounding factors, with the minor allele conferring a lower risk of diabetic retinopathy (OR 0.812, $p=0.0039$ for rs17684886 and OR 0.835, $p=0.0116$ for rs599019). In our sub-analysis, we detected associations between rs6427247 and rs899036 and severe diabetic retinopathy after adjustment for confounding factors (OR 1.368, $p=0.0333$ for rs6427247 and OR 0.340, $p=0.0005$ for rs899036). However, we cannot fully exclude the possibility that the association between rs17684886 or rs599019 and diabetic retinopathy was a false-positive finding because only a trend association or no association was observed after adjusting for multiple comparisons (empirical $p=0.0783$ for rs17684886 and empirical $p=0.24$ for rs599019). Nevertheless, we also determined that these two SNPs were significantly associated with the severity of diabetic retinopathy ($p=0.0365$ for rs17684886 and $p=0.0252$ for rs599019 for trend analysis), further supporting a role for them in diabetic retinopathy and limiting the possibility of a false positive. Besides, risk alleles of rs17684886 and rs59919 for diabetic retinopathy are the same as those reported for severe diabetic retinopathy in previous GWAS [16, 17] and the association of rs6427247 and rs899019 with severe diabetic retinopathy in our study is in agreement with the results of previous GWAS [16].

The four variants associated with diabetic retinopathy or severe diabetic retinopathy in our study are located in the noncoding regions—rs17684886 is an intron SNP of *ZNRF1* and the other three SNPs are located in intergenic regions:

Table 3 Distribution of rs17684886 and rs599019 among patients with different severities of diabetic retinopathy

SNP	Minor allele	Patients without retinopathy ($n=1,110$)		Mild NPDR ($n=490$)		Moderate NPDR ($n=136$)		Severe NPDR ($n=118$)		PDR ($n=45$)		p value for trend analysis
		MAF	Genotype count 11/12/22 ^a	MAF	Genotype count 11/12/22 ^a	MAF	Genotype count 11/12/22 ^a	MAF	Genotype count 11/12/22 ^a	MAF	Genotype count 11/12/22 ^a	
rs17684886	A	0.493	285/555/270	0.542	101/242/142	0.544	26/72/38	0.517	26/62/30	0.556	12/16/17	0.0365
rs599019	C	0.430	363/517/210	0.388	194/200/86	0.396	48/67/20	0.343	52/51/15	0.443	14/21/9	0.0252

^a 11, major allele homozygotes; 12, heterozygotes; 22, minor allele homozygotes

MAF, minor allele frequency

Table 4 Associations of SNPs with severe diabetic retinopathy

Chromosome	SNP	Position (Build 38)	Nearest gene	Minor/ major allele	Risk allele	Minor allele frequency		Allelic OR (95% CI)	p value (empirical p value ^a)	Genotypic OR (95% CI) ^b	p value (empirical p value ^b)
						Severe NPDR or PDR	Mild NPDR				
1	rs6427247	170411339	SCYL1BP1 (GORAB)	G/A	G	0.362	0.304	1.298 (0.997, 1.690)	0.0525	1.368 (1.025, 1.825)	0.0333 (0.55)
1	rs1033465	173018590	TNFSF18	T/A	T	0.025	0.021	1.195 (0.521, 2.740)	0.67	1.313 (0.558, 3.090)	0.53
1	rs3014267	227365216	CDC42BP4	G/A	G	0.255	0.237	1.103 (0.825, 1.474)	0.51	1.197 (0.873, 1.641)	0.27
1	rs476141	244013122	LOC339529	A/C	C	0.194	0.220	0.856 (0.625, 1.172)	0.33	0.912 (0.643, 1.293)	0.61
2	rs699549	4657673	LINC01249	T/C	C	0.219	0.252	0.833 (0.616, 1.125)	0.23	0.734 (0.526, 1.024)	0.07
2	rs763970	137878563	HNNMT	A/C	C	0.258	0.268	0.947 (0.711, 1.261)	0.71	1.027 (0.757, 1.393)	0.86
3	rs11927173	23183703	UBE2E2	C/T	T	0.169	0.194	0.844 (0.607, 1.175)	0.32	0.858 (0.607, 1.214)	0.39
3	rs1197310	133409380	BFSF2	A/T	A	0.509	0.487	1.092 (0.848, 1.405)	0.50	1.192 (0.899, 1.580)	0.22
4	rs4865047	55955640	CEP135	T/C	T	0.250	0.241	1.052 (0.786, 1.407)	0.74	0.956 (0.688, 1.329)	0.79
4	rs11736136	82097121	LOC101928987	G/A	A	0.047	0.072	0.635 (0.358, 1.127)	0.12	0.641 (0.326, 1.259)	0.20
4	rs1902491	155134181	NPY2R	G/T	T	0.150	0.152	0.990 (0.697, 1.407)	0.96	0.997 (0.678, 1.467)	0.99
5	rs1445754	84279813	EDIL3	A/T	T	0.043	0.060	0.706 (0.388, 1.283)	0.25	0.691 (0.360, 1.324)	0.26
5	rs2300782	111453087	CAMK4	T/C	T	0.537	0.457	1.380 (1.073, 1.777)	0.0121 (0.26)	1.081 (0.817, 1.432)	0.58
6	rs17083119	121080964	TBC1D32	G/A	A	0.096	0.101	0.939 (0.614, 1.437)	0.77	1.055 (0.658, 1.691)	0.83
6	rs227455	165064562	C6orf118	C/T	T	0.454	0.470	0.938 (0.729, 1.207)	0.62	0.996 (0.754, 1.315)	0.98
8	rs3098241	103413076	SLC25A32	G/A	A	0.371	0.378	0.970 (0.748, 1.257)	0.82	1.028 (0.752, 1.404)	0.86
10	rs17670074	19416454	MALRD1	C/A	C	0.387	0.374	1.055 (0.815, 1.365)	0.69	1.051 (0.786, 1.407)	0.74
10	rs9888035	19426089	MALRD1	C/T	T	0.003	0.010	0.292 (0.037, 2.292)	0.21	0.300 (0.036, 2.475)	0.26
11	rs899036	41661360	API5	C/A	A	0.055	0.116	0.448 (0.268, 0.749)	0.0017 (0.0454)	0.340 (0.185, 0.624)	0.0005 (0.0093)
11	rs10501943	100076267	CNTN5	C/T	C	0.059	0.034	1.765 (0.989, 3.149)	0.0516	1.803 (0.866, 3.751)	0.12
13	rs4941432	42524211	TNFSF11	A/G	A	0.203	0.168	1.257 (0.914, 1.728)	0.16	1.277 (0.904, 1.804)	0.17
15	rs10519765	32913223	FMN1	A/G	A	0.092	0.079	1.175 (0.756, 1.828)	0.47	1.249 (0.744, 2.096)	0.40
15	rs11635920	32920456	FMN1	A/T	A	0.089	0.079	1.132 (0.724, 1.770)	0.59	1.264 (0.752, 2.125)	0.38
16	rs17684886	75052977	ZNRF1	A/T	A	0.472	0.458	1.061 (0.825, 1.364)	0.65	0.958 (0.723, 1.268)	0.76
18	rs599019	294495	COLEC12	C/A	A	0.370	0.388	0.930 (0.717, 1.206)	0.58	0.942 (0.713, 1.244)	0.67

The OR with 95% CI shown is for the minor allele

^a Empirical *P* values are based on 10,000 permutations

^b Adjusted for duration of diabetes, HbA_{1c}, systolic blood pressure, diastolic blood pressure, BMI and nephropathy under an additive model

Table 5 Meta-analysis of associations between SNPs and severe diabetic retinopathy

SNP	Chinese population			Mexican-American population			Meta-analysis			
	Risk allele	OR (95% CI)	<i>p</i> value	Risk allele	OR (95% CI)	<i>p</i> value	Risk allele	OR (95% CI)	<i>p</i> value	<i>Q</i> value
rs6427247	G	1.577 (1.240, 2.004)	0.0333	G	2.17 (1.41, 3.35)	4.56×10^{-4}	G	1.577 (1.240, 2.004)	2.01×10^{-4}	0.08
rs899036	A	0.330 (0.214, 0.510)	0.0005	A	0.32 (0.17, 0.59)	2.52×10^{-4}	A	0.330 (0.214, 0.510)	5.84×10^{-7}	0.89

The OR with 95% CI shown is for the minor allele with adjustment for confounding factors

The *Q* value is calculated by the Cochran *Q* test to assess homogeneity

rs599019 is located downstream of *COLEC12*; rs6427247 is located upstream of *SCYL1BP1* (also known as *GORAB*) and rs899036 is located upstream of *API5*. To date, 43% of all trait-associated SNPs identified in GWAS studies are located in intergenic regions [24]. However, the intergenic SNPs may have an impact on neighbouring gene function. *ZNRF1* encodes an E3 ubiquitin-protein ligase that plays a role in neural-cell differentiation. It has been reported to interact with tubulin, promote Wallerian degeneration and regulate Na^+/K^+ ATPase [25–27]. *COLEC12* encodes a member of the C-lectin family. This protein is an endothelial cell surface receptor that displays several functions associated with host defence [28, 29]. *SCYL1BP1* encodes SCY1-like 1-binding protein 1, a regulator of the p53 pathway with tumour-suppressive function [30–32]. *API5* encodes the protein apoptosis inhibitor 5, which is reported to be a suppressor of apoptosis and to play a key role in tumour progression [33, 34]. However, the relationships of these SNPs and other genes are largely unknown. Thus, additional studies are needed to identify the causal loci and genes and to elucidate the underlying mechanism of diabetic retinopathy.

This study has a number of limitations. First, the small sample size was insufficient to identify SNPs with smaller effects on diabetic retinopathy. Second, we did not adjust for lifestyle factors, such as alcohol consumption and cigarette smoking, as confounding factors. Whether there is an interaction between lifestyle and these genetic variants on diabetic retinopathy remains unknown. Third, we could not entirely exclude population stratification in patients with and without diabetic retinopathy as a potential source of bias and incorrect inferences in genotype–disease association. The effect of population stratification may be minimal in the current study, however, as all the participants were recruited from the same geographic area with the same ancestry. Hence, studies with a larger sample size are needed to further replicate the associations identified in our study of a Chinese population.

In summary, we determined that rs17684886 in *ZNRF1* and rs599019 near *COLEC12* were associated with the risk of diabetic retinopathy and that rs6427247 near *SCYL1BP1* and rs899036 near *API5* were associated with the risk of severe

diabetic retinopathy in Chinese patients with type 2 diabetes. Additional studies are needed to replicate this finding.

Acknowledgements The authors are grateful to the patients who participated in this research and gratefully acknowledge the skilful technical support of the nursing and medical staff at the Shanghai Clinical Centre for Diabetes and Department of Endocrinology and Metabolism.

Funding This work was supported by grants from the national 973 programme (2011CB504001), the National Science Foundation of China (81200582, 81322010 and 81170735), the national 863 programme (2012AA02A509), the Shanghai Talent Development Grant (2012041) and the Excellent Young Medical Expert of Shanghai (XYQ2011041).

Duality of interest The authors declare that there is no duality of interest associated with this manuscript.

Contribution statement DP and JW performed the majority of the analyses and drafted the manuscript. RZ, FJ and ST participated in the design of the study. MC, JY, XS, SW, TW and DY participated in the data analysis. YB provided helpful comments on study design and data analysis. CH and WJ conceived the study, participated in its design and helped to draft the manuscript. All authors contributed to the drafting or critical revision of the manuscript for important intellectual content. All authors read and approved the final manuscript. CH and WJ are the guarantors of this work.

References

- Klein BE (2007) Overview of epidemiologic studies of diabetic retinopathy. *Ophthalmic Epidemiol* 14:179–183
- Yau JW, Rogers SL, Kawasaki R et al (2012) Global prevalence and major risk factors of diabetic retinopathy. *Diabetes Care* 35:556–564
- Stratton IM, Adler AI, Neil HA et al (2000) Association of glycaemia with macrovascular and microvascular complications of type 2 diabetes (UKPDS 35): prospective observational study. *BMJ* 321:405–412
- Tapp RJ, Shaw JE, Harper CA et al (2003) The prevalence of and factors associated with diabetic retinopathy in the Australian population. *Diabetes Care* 26:1731–1737
- Zhang X, Saaddine JB, Chou CF et al (2010) Prevalence of diabetic retinopathy in the United States, 2005–2008. *JAMA* 304:649–656
- Sun JK, Keenan HA, Cavallerano JD et al (2011) Protection from retinopathy and other complications in patients with type 1 diabetes of extreme duration: the Joslin 50-Year Medalist Study. *Diabetes Care* 34:968–974

7. Liew G, Klein R, Wong TY (2009) The role of genetics in susceptibility to diabetic retinopathy. *Int Ophthalmol Clin* 49:35–52
8. Leslie RD, Pyke DA (1982) Diabetic retinopathy in identical twins. *Diabetes* 31:19–21
9. The Diabetes Control and Complications Trial Research Group (1997) Clustering of long-term complications in families with diabetes in the diabetes control and complications trial. *Diabetes* 46:1829–1839
10. Zhang X, Gao Y, Zhou Z, Wang J, Zhou Q, Li Q (2010) Familial clustering of diabetic retinopathy in Chongqing, China, type 2 diabetic patients. *Eur J Ophthalmol* 20:911–918
11. Rema M, Saravanan G, Deepa R, Mohan V (2002) Familial clustering of diabetic retinopathy in South Indian type 2 diabetic patients. *Diabet Med* 19:910–916
12. Looker HC, Nelson RG, Chew E et al (2007) Genome-wide linkage analyses to identify Loci for diabetic retinopathy. *Diabetes* 56:1160–1166
13. Hietala K, Forsblom C, Summanen P, Groop PH (2008) Heritability of proliferative diabetic retinopathy. *Diabetes* 57:2176–2180
14. Arar NH, Freedman BI, Adler SG et al (2008) Heritability of the severity of diabetic retinopathy: the FIND-Eye study. *Invest Ophthalmol Vis Sci* 49:3839–3845
15. Ng DP (2010) Human genetics of diabetic retinopathy: current perspectives. *J Ophthalmol* 2010:172593
16. Fu YP, Hallman DM, Gonzalez VH et al (2010) Identification of diabetic retinopathy genes through a genome-wide association study among Mexican-Americans from Starr County, Texas. *J Ophthalmol* 2010:861291
17. Grassi MA, Tikhomirov A, Ramalingam S, Below JE, Cox NJ, Nicolae DL (2011) Genome-wide meta-analysis for severe diabetic retinopathy. *Hum Mol Genet* 20:2472–2481
18. Grassi MA, Tikhomirov A, Ramalingam S et al (2012) Replication analysis for severe diabetic retinopathy. *Invest Ophthalmol Vis Sci* 53:2377–2381
19. Jia W, Gao X, Pang C et al (2009) Prevalence and risk factors of albuminuria and chronic kidney disease in Chinese population with type 2 diabetes and impaired glucose regulation: Shanghai diabetic complications study (SHDCS). *Nephrol Dial Transplant* 24:3724–3731
20. Hu C, Zhang R, Wang C et al (2010) Effects of GCK, GCKR, G6PC2 and MTNR1B variants on glucose metabolism and insulin secretion. *PLoS One* 5:e11761
21. Hu C, Wang C, Zhang R et al (2010) Association of genetic variants of NOS1AP with type 2 diabetes in a Chinese population. *Diabetologia* 53:290–298
22. Wilkinson CP, Ferris FL 3rd, Klein RE et al (2003) Proposed international clinical diabetic retinopathy and diabetic macular edema disease severity scales. *Ophthalmology* 110:1677–1682
23. Purcell S, Neale B, Todd-Brown K et al (2007) PLINK: a toolset for whole-genome association and population-based linkage analysis. *Am J Hum Genet* 81:559–575
24. Hindorf LA, Sethupathy P, Junkins HA et al (2009) Potential etiologic and functional implications of genome-wide association loci for human diseases and traits. *Proc Natl Acad Sci U S A* 106:9362–9367
25. Yoshida K, Watanabe M, Hatakeyama S (2009) ZNRF1 interacts with tubulin and regulates cell morphogenesis. *Biochem Biophys Res Commun* 389:506–511
26. Wakatsuki S, Saitoh F, Araki T (2011) ZNRF1 promotes Wallerian degeneration by degrading AKT to induce GSK3B-dependent CRMP2 phosphorylation. *Nat Cell Biol* 13:1415–1423
27. Hoxhaj G, Najafov A, Toth R, Campbell DG, Prescott AR, MacKintosh C (2012) ZNRF2 is released from membranes by growth factors and, together with ZNRF1, regulates the Na⁺/K⁺ ATPase. *J Cell Sci* 125:4662–4675
28. Nakamura K, Funakoshi H, Miyamoto K, Tokunaga F, Nakamura T (2001) Molecular cloning and functional characterization of a human scavenger receptor with C-type lectin (SRCL), a novel member of a scavenger receptor family. *Biochem Biophys Res Commun* 280:1028–1035
29. Jang S, Ohtani K, Fukuo H et al (2009) Scavenger receptor collectin placenta 1 (CL-P1) predominantly mediates zymosan phagocytosis by human vascular endothelial cells. *J Biol Chem* 284:3956–3965
30. Yan J, Di Y, Shi H, Rao H, Huo K (2010) Overexpression of SCYL1-BP1 stabilizes functional p53 by suppressing MDM2-mediated ubiquitination. *FEBS Lett* 584:4319–4324
31. Hu L, Liu M, Chen L et al (2012) SCYL1 binding protein 1 promotes the ubiquitin-dependent degradation of Pirh2 and has tumor-suppressive function in the development of hepatocellular carcinoma. *Carcinogenesis* 33:1581–1588
32. Yang ZP, Xie YH, Ling DY et al (2014) SCYL1BP1 has tumor-suppressive functions in human lung squamous carcinoma cells by regulating degradation of MDM2. *Asian Pac J Cancer Prev* 15: 7467–7471
33. Morris EJ, Michaud WA, Ji JY, Moon NS, Rocco JW, Dyson NJ (2006) Functional identification of Api5 as a suppressor of E2F-dependent apoptosis in vivo. *PLoS Genet* 2:e196
34. Cho H, Chung JY, Song KH et al (2014) Apoptosis inhibitor-5 overexpression is associated with tumor progression and poor prognosis in patients with cervical cancer. *BMC Cancer* 14:545