

Withaferin A inhibits pro-inflammatory cytokine-induced damage to islets in culture and following transplantation

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Abstract

Aims/hypothesis Beta cell death triggered by pro-inflammatory cytokines plays a central role in the pathogenesis of type 1 diabetes and loss of transplanted islets. The nuclear factor κ B (NF- κ B) signalling pathway is a key regulator of beta cell stress response, survival and apoptosis. Withaferin A (WA), a steroidal lactone derived from *Withania somnifera*, has been demonstrated to be a potent, safe, anti-inflammatory molecule that can inhibit NF- κ B signalling. Therefore, we evaluated the ability of WA to protect mouse and human islets from the damaging effects of pro-inflammatory cytokines in vitro and following intraportal transplantation.

Methods Mouse and human islets were treated with a cytokine cocktail, and NF- κ B activation was measured by immunoblots, p65 nuclear translocation and chromatin immunoprecipitation of p65-bound DNA. Intraportal transplantation of a marginal

mass of syngeneic mouse islets was performed to evaluate the in vivo protective effect of WA.

Results Treatment with WA substantially improved islet engraftment of syngeneic islets (83% for infusion with 200 islets + WA; 0% for 200 islets + vehicle) in a mouse model of diabetes, compared with marginal graft controls with superior islet function in WA-treated mice confirmed by glucose tolerance test. Treatment of human and mouse islets with WA prevented cytokine-induced cell death, inhibited inflammatory cytokine secretion and protected islet potency.

Conclusions WA was shown to be a strong inhibitor of the inflammatory response in islets, protecting against cytokine-induced cell damage while improving survival of transplanted islets. These results suggest that WA could be incorporated as an adjunctive treatment to improve islet transplant outcome.

Keywords Beta cell · Cytokine · Islet transplantation · NF- κ B

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Abbreviations

G-CSF	Granulocyte-colony stimulating factor
IBMIR	Instant blood-mediated inflammatory response
I κ B	Inhibitor of κ B
IKK β	Inhibitor of κ B kinase beta
iNOS	Inducible nitric oxide synthase
IP-10	IFN γ -induced protein 10
IPGTT	Intraperitoneal glucose tolerance test
MCP-1	Monocytic chemotactic protein-1
MIG	Monokine induced by IFN- γ
MIP	Macrophage inflammatory protein
NEMO	NF- κ B essential modulator
NF κ B	Nuclear factor κ B
PI	Propidium iodide
RANTES	Regulated and normal T cell-expressed and secreted chemokine

STZ	Streptozotocin
TF	Tissue factor
WA	Withaferin A

Introduction

Type 1 diabetes mellitus is an autoimmune disorder that leads to destruction of the insulin-producing beta cells of the pancreas. Islet cell transplantation to replace depleted beta cells has emerged as an effective treatment in recent years. However, a substantial number of islets are destroyed by the instant blood-mediated inflammatory response (IBMIR), cytokines, hypoxia–reoxygenation injury, apoptosis, factors related to isolation and other factors [1, 2]. The blood-mediated nonspecific immune response is characterised by coagulation, complement activation, platelet aggregation and cytokine release [2]. Cytokines such as TNF- α , IFN- γ and IL-1 β have important pro-inflammatory and pro-apoptotic roles in type 1 diabetes and islet transplantation [3–5]. Although immunosuppressants are administered after transplantation of islets, effective control of cytokine-mediated damage to islets is still not achieved.

Inflammatory cytokines cause beta cell damage through a signal transduction pathway initiated at TNF and IL-1 receptors and mediated through a downstream signalling molecule, nuclear factor κ B (NF- κ B). It has been shown that a cytokine cocktail of TNF- α , IFN- γ , and IL-1 β led to NF- κ B activation in human islets [6]. A recent study examined the dual role of hypoxia and cytokines on NF- κ B activation and proposed that cytokine-mediated NF- κ B activation was modulated by different levels of hypoxia [7]. Pro-inflammatory cytokines have also been shown to upregulate mRNA of several chemokines in islets [8]. These chemokines act as chemoattractants for lymphocytes, directing them towards the site of transplantation. This has been observed with CCL2 (monocytic chemoattractant protein-1 [MCP-1]) secretion, demonstrating macrophage infiltration of islet grafts leading to a negative clinical outcome [9, 10]. In addition, pro-inflammatory cytokines promote beta cell damage by exacerbating cellular injury and death signalling pathways mediated by NF- κ B in islets [10]. Since NF- κ B is a cytokine-induced mediator of several inflammatory genes in beta cells, it provides an attractive therapeutic target for inhibition.

Withaferin A (WA), a naturally occurring plant-derived molecule, has been identified as a multifunctional, bioactive compound with strong anti-inflammatory, antioxidant and anti-angiogenic properties [11]. The origin of WA is from a plant species called *Withania somnifera*, which has been used for centuries in traditional eastern medicine with proven clinical safety [12, 13]. There are several extracts from the leaves of *W. somnifera*, but a study by Kaileh et al identified the steroidal lactone, WA, as the most potent

inhibitor of NF- κ B based on the hyperphosphorylation of inhibitor of κ B (I κ B) kinase beta (IKK β) preventing I κ B degradation [14]. Grover et al suggested that WA inhibits NF- κ B by preventing the formation of NF- κ B essential modulator (NEMO)/IKK β complex [15]. WA has been studied as an anti-inflammatory compound in cystic fibrosis [16] with further potential to treat chronic inflammatory diseases such as rheumatoid arthritis, asthma and inflammatory bowel disease [17, 18].

The objective of this study was to demonstrate the protective effects of WA in preventing cytokine-induced damage to islets under in vivo and in vitro conditions. We have previously demonstrated that inflammatory cytokines cause dramatically increased levels of several inflammatory cytokines and chemokines (inducible nitric oxide synthase [iNOS], IL-1 β , regulated and normal T cell-expressed and secreted chemokine [RANTES], monokine induced by IFN- γ [MIG] and IFN- γ -induced protein 10 [IP-10]) in human islets as determined by quantitative PCR analysis [8]. WA was shown to decrease the mRNA levels of several of these chemokines by tenfold or more. This led to further investigation of the role of WA as pharmacological modulator of the early inflammatory reaction in an in vivo mouse model. Inclusion of human pancreatic islets in this study allowed us to determine whether the results from the mice data can be translated to a clinical setting.

Methods

Human islet isolation Human-research-grade pancreases were used for isolating islets using methods described previously [19–21]. Human islets used in this study were handpicked to greater than 95% purity. Islets were cultured with Connaught Medical Research Laboratories (CMRL)-based human islet culture media containing 0.1% BSA with kanamycin (37°C, 5% CO₂ up to 96 h). These experiments were approved by the Institutional Review Board at Baylor Research Institute (BRI), Dallas, TX, USA.

Mouse islet isolation Islets were isolated from male inbred C57BL/6N mice (Harlan Labs, Houston, TX, USA) using a previously described procedure [22, 23]. Islets were cultured at a purity greater than 95% in DMEM supplemented with kanamycin and 0.1% BSA (37°C, 5% CO₂ up to 96 h).

Intraportal transplantation Diabetes was induced in the mouse models by injecting streptozotocin (STZ; 180 mg/kg) intravenously. Mice were considered diabetic when their blood glucose was greater than 19.5 mmol/l for two consecutive days. Two-hundred islets were injected into the portal vein by puncturing the hepatic portal vein [22]. WA (1.25 mg/kg body weight) or vehicle (Dulbecco's phosphate-buffered saline [DPBS]) was injected into the

peritoneal cavity. These experiments were approved by the Institutional Animal Care and Use Committee at BRI.

Intraperitoneal glucose tolerance test Naive, transplanted and STZ-induced diabetic mice were used to perform an intraperitoneal glucose tolerance test (IPGTT) [22]. The mouse's food was removed for 12–14 h, then an injection of glucose (1 g/kg) was administered intraperitoneally. Blood glucose was monitored with Accu-Chek (Roche Diagnostics, Indianapolis, IN, USA) at 0, 30 and 120 min after the injection.

Cytokine and WA treatment After isolation and overnight culture, islets were subjected to inflammatory conditions *in vitro* with a cytokine cocktail of the species-specific recombinant proteins (1,000 U/ml TNF- α , 1,000 U/ml IFN- γ and 50 U/ml IL-1 β) [24, 25]. Islets were treated with WA at concentrations in the range 0.25–2.0 μ g/ml. All islets were cultured in a humidified incubation chamber at 37°C with 5% CO₂ up to 96 h. Viability was determined by Hoechst 33342/propidium iodide (PI) staining. See the electronic supplementary material (ESM) Methods for detailed procedure.

Immunohistochemistry Islets and engrafted livers were fixed and stained as described previously and imaged via fluorescence microscopy (BX16 and DP-72; Olympus, Tokyo, Japan) [26]. Mean fluorescence intensity was calculated as the integrated density divided by the area in the insulin-positive region of interest.

TUNEL assay The TUNEL assay was performed with a ApopTag fluorescein *in situ* apoptosis detection kit (Millipore, Billerica, MA, USA) [22]. See ESM Methods for detailed procedure.

Measurement of secreted inflammatory cytokines and chemokines Fifty islets were cultured in quintuplet in islet culture media for 96 h with the cytokine and WA treatment. Pro-inflammatory cytokines in culture supernatant fractions were measured with a Luminex 200 system (Luminex, Austin, TX, USA) [9]. See ESM Methods for detailed procedure.

Western blotting After cytokine-treated cells were lysed, the proteins in the lysate were subjected to SDS-PAGE and transferred onto nitrocellulose membranes. The membranes were incubated with primary antibodies to I κ B- α and ERK1/2, followed by secondary antibodies conjugated with fluorescent dyes. See ESM Methods for further detail.

Chromatin immunoprecipitation assay Chromatin DNA–protein was crosslinked in human islets (5,000 islet equivalents [IEQ]) or MIN6 cells (5×10^7) with 1% formaldehyde. See ESM Methods for detailed procedure.

Real-time PCR assay After human islets had been lysed by lysis buffer, RNA was purified using an Absolutely RNA Miniprep kit (Stratagene, La Jolla, CA, USA) and then reverse transcribed into cDNA. See ESM Methods for further detail.

p65 Translocation assay The NF- κ B p65 nuclear localisation was determined by overlap of Alexa-594 labelled NF- κ B antibody and DAPI-stained nuclei with confocal laser scanning microscope (Leica Microsystems, Wetzlar, Germany). See ESM Methods for detailed procedure.

Neutrophil infiltration in purified human islets mixed with autologous blood A modified loop model was employed using human islets mixed with autologous human blood samples. See ESM Methods for detailed procedure.

Glucose-stimulated insulin secretion test Islets were incubated with low (1.67 mmol/l) and then high (16.7 mmol/l) concentrations of glucose solution. See ESM Methods for detailed procedure.

Reagents WA was purchased from Enzo Life Sciences (Plymouth Meeting, PA, USA). The recombinant human cytokines TNF- α (1.1×10^5 U/ μ g), IFN- γ (2.0×10^4 U/ μ g) and IL-1 β (2.0×10^5 U/ μ g) were purchased from R&D Systems (Minneapolis, MN, USA). The recombinant mouse cytokines TNF- α (2.7×10^5 U/ μ g), IFN- γ (8.43×10^3 U/ μ g) and IL-1 β (1.1×10^6 U/ μ g) were also purchased from R&D Systems.

Statistics The statistical significance between two groups was analysed by the Student's *t* test. For statistical analysis among three groups or more, one-way ANOVA with Tukey–Kramer post hoc tests was performed using StatView 5.0 (SAS Institute, Cary, NC, USA) [27]. A significant difference was achieved when $p < 0.05$.

Results

WA improves survival of intraportally transplanted mouse syngeneic islets To determine the anti-inflammatory effects of WA on *in vivo* islet transplantation, a syngeneic C57BL/6 mouse islet transplant model was used. Initial experiments determined that 400 islets were required to achieve normoglycaemia following intraportal infusion. Using only half that number of islets resulted in a 0% success rate, indicating that 200 islets alone were insufficient to reverse hyperglycaemia (Fig. 1a, b). Daily intraperitoneal injection of WA (1.25 mg/kg body weight) for 7 days post-transplant beginning immediately after islet infusion led to normoglycaemia in 5/6 mice (83%) with infusion of a suboptimal dose of 200 islets

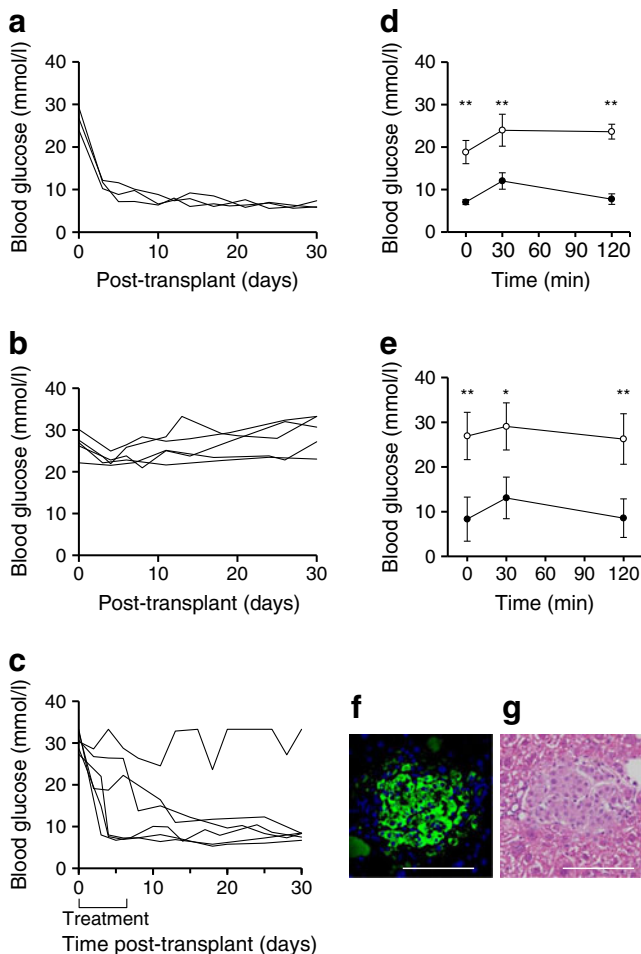


Fig. 1 Intraportal islet isograft survival is improved by WA administration as demonstrated by normoglycaemia, glucose tolerance test and histological evaluation. Intraportal transplantation of 400 islets into C57BL/6 mice had a success rate of 100% (3/3) (a), but transplantation of 200 syngeneic islets with injection of 0.5 ml PBS (vehicle) resulted in 0% (0/5) of recipients achieving normoglycaemia (b). (c) Daily administration of WA (1.25 mg/kg body weight) starting immediately after transplantation and lasting 7 days yielded a much higher success rate of 83% (5/6). (d) IPGTT was performed to assess glycaemic control of naive (black circles) and STZ-induced diabetic mice (white circle) for comparison against (e) WA-treated (black circles) and vehicle-treated mice (white circles) on day 30 post-transplant. There was a significant difference in blood glucose levels at each time. Naive and WA-treated mice demonstrated a similar pattern with normalisation of blood glucose levels below 11.1 mmol/l after 120 min. After 30 days, hepatectomy of recipient mice was performed and the liver was fixed in formalin, embedded in paraffin and stained for insulin (f) and with haematoxylin and eosin (g). Graphical representations are expressed as means \pm SD. * p <0.005, ** p <0.0005 as determined by Tukey–Kramer post hoc tests. Scale bar 50 μ m

(Fig. 1c). The IPGTT test performed to assess islet function showed a significant difference between normal and STZ-treated mice (Fig. 1d). A similar pattern was seen in the WA-treated mice, which showed a significant difference for each time point compared with the vehicle-treated mice (Fig. 1e, 8.4 ± 5.0 vs 26.9 ± 5.3 mmol/l glucose at 0 min,

p <0.0005; 14.9 ± 4.7 vs 29.1 ± 5.3 mmol/l glucose at 30 min, p <0.005; 8.7 ± 4.5 vs 26.3 ± 5.7 mmol/l glucose at 120 min, p <0.0005). This data demonstrated that mice receiving WA post-transplant had a sufficient islet mass to be functional and comparable with naive mice. Immunohistochemical staining of the engrafted islets in the presence of WA showed functionally intact islets demonstrated by positive insulin stain and retention of the islet architecture (Fig. 1f, g).

WA prevents NF- κ B translocation after intraportal transplantation WA has been reported to be a multifunctional molecule with an inhibitory action on NF- κ B [11, 14]. Hence we evaluated the ability of WA to prevent NF- κ B activation post-transplant by analysing p65 translocation in transplanted islets. Mice were transplanted with 200 syngeneic islets as described before and administered either vehicle or WA (1.25 mg/kg body weight) intraperitoneally. Livers from mice ($n=3$ per treatment) were removed 3 h post-transplant and stained for insulin and p65 (ESM Fig. 1). Islet sections were visualised with confocal microscopy. We observed that compared with vehicle-treated mice (ESM Fig. 1a–c), WA-treated mice (ESM Fig. 1d–f) had significantly decreased p65 nuclear localisation in islets (ESM Fig. 1g)

WA does not affect islet viability and function in vitro at low concentrations To determine its toxicity on islets, human ($n=3$) and mouse ($n=6$) islets were cultured in the presence of WA (0.5–2.0 μ g/ml) and viability and function were tested. After 24 h, human islets cultured with 0.5 and 1.0 μ g/ml of WA showed no significant change in viability whereas 2.0 μ g/ml WA caused a significant decrease (ESM Fig. 2a) compared with control islets ($70.5\pm 8.8\%$ vs $94.9\pm 4.5\%$, t test, p <0.0001). After 48 h, there was no noticeable difference between control ($91.8\pm 7.4\%$) and treatment with either 0.5 μ g/ml ($89.0\pm 4.6\%$, $p=0.36$) or 1.0 μ g/ml WA ($91.9\pm 3.5\%$, $p=0.87$); furthermore, at 96 h there was no change with 0.5 μ g/ml ($97.7\pm 0.8\%$, $p>0.05$) and 1.0 μ g/ml WA ($95.9\pm 2.6\%$, $p>0.05$) compared with control ($93.5\pm 4.6\%$). Concentrations of 0.25 and 0.5 μ g/ml WA caused no significant decrease in viability of mouse islets (ESM Fig. 3a, b) whereas 1.0 μ g/ml WA significantly decreased viability compared with control ($87.8\pm 11.1\%$ vs $97.3\pm 2.2\%$, p <0.05); this pattern was continued at 48 h and 96 h. Furthermore, there was no significant difference in islet function as indicated by stimulation index (ESM Fig. 2b) between control human islets (4.55 ± 1.53) or islets cultured with 0.5 (3.52 ± 0.41 , $p=0.10$) or 1.0 μ g/ml WA (4.21 ± 1.97 , $p=0.51$) after 24 h. Similarly, mouse islets showed no significant difference in stimulation index after 24 h between control (11.13 ± 9.28) and islets treated with 0.5 μ g/ml WA (7.05 ± 2.20 , $p=0.30$) (ESM Fig. 3c). These results showed that WA at low concentrations did not affect islet viability and function.

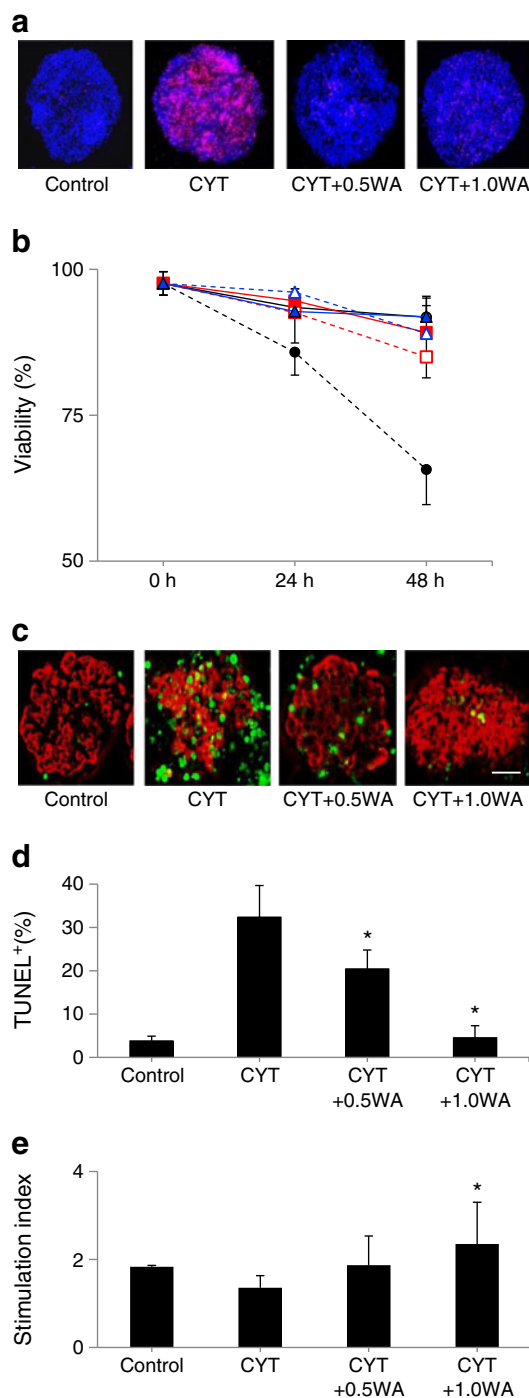
WA protects islets from cytokine-induced damage and preserves islet function We evaluated the effect of WA on islet cell viability and potency after exposure to cytokines. Cytokine cocktail significantly increased islet cell death at 24 h compared with control levels in human islets (viability: $85.8 \pm 3.9\%$ vs $93.5 \pm 3.2\%$, $p < 0.05$; Fig. 2a, b). WA prevented this damage at concentrations of 0.5 ($92.6 \pm 5.1\%$ viability) and 1.0 $\mu\text{g/ml}$ ($94.6 \pm 2.9\%$) ($p < 0.05$ [vs cytokine] for both concentrations of WA), while there was no significant difference in viability between control islets and islets treated with WA alone ($p > 0.05$). After 48 h there was a significant difference in viability of the human islets seen between each treatment and cytokines. Viability of mouse islets was also significantly decreased by cytokines compared with control (viability: $78.1 \pm 11.6\%$ vs $93.3 \pm 2.9\%$, $p < 0.05$; ESM Fig. 4a, b), and addition of WA to islets cultured with cytokines improved outcome. This pattern continued in mouse islets after 48 h.

In addition, apoptosis in human islets was increased by cytokine treatment as demonstrated by increased TUNEL positive staining in beta cells. At 24 h, there was a significant difference between each treatment group (Fig. 2c, d) except when comparing control vs treatment with cytokines plus 1.0 $\mu\text{g/ml}$ WA. The TUNEL assay demonstrated that WA can effectively negate the effect of cytokines on beta cell apoptosis.

Evaluation of the ability of human islets to secrete insulin after 24 h revealed that cytokines significantly decreased the stimulation index compared with control (1.36 ± 0.34 vs 1.83 ± 0.15 , $p < 0.05$; Fig. 2e). However, 1.0 $\mu\text{g/ml}$ WA significantly increased the stimulation index compared with cytokines alone (2.20 ± 0.61 , $p < 0.05$) and showed no significant difference compared with control islets ($p > 0.05$). Stimulation index values for mouse islets demonstrated a similar pattern (ESM Fig. 4c).

Fig. 2 WA protects human islets from inflammatory cytokine-mediated cell death. (a, b) Human islets were incubated with the inflammatory cytokine cocktail (CYT) for 24 and 48 h with or without WA, and with WA alone. Islets were recovered and viability was assessed by Hoechst 33342/PI nuclear staining. (b) Black solid line with white circle, control; black dotted line with black circle, cytokine; red dotted line with white square, cytokine plus 0.5 $\mu\text{g/ml}$ WA (CYT+0.5WA); red solid line with red square, cytokine plus 0.5 $\mu\text{g/ml}$ WA (CYT+1.0WA); blue dotted line with white triangle, 0.5 $\mu\text{g/ml}$ WA (0.5WA); blue solid line with blue triangle 1.0 $\mu\text{g/ml}$ WA (1.0WA). (c) Cytokine-mediated beta cell apoptosis was confirmed by TUNEL staining. Human islets were stained for insulin (red) and apoptotic nuclei (green). (d) Manual counting of more than 250 nuclei revealed higher levels of apoptosis when islets were exposed to cytokines. This raised level was significantly reduced by the addition of WA, which returned apoptosis to control levels (1.0 $\mu\text{g/ml}$ WA). (e) Islet potency was assessed by static incubation after 24 h. Results shown are representative of six independent human islet preparations. Graphical representations are expressed as means \pm SD. $p < 0.05$ compared with CYT as determined by Tukey–Kramer post hoc tests. Scale bar = 50 μm

WA prevents degradation of I κ B and NF- κ B association with DNA when human islets and beta cells are exposed to cytokines Inflammation elicited by immune cells requires the activation of NF- κ B [28–30]. NF- κ B is activated as a direct result of phosphorylation and rapid degradation of inhibitory I κ B protein [31, 32]. To identify effects of cytokines on NF- κ B signalling in islets, we analysed the effects of cytokine cocktail on I κ B in human islets and beta cell lines (MIN6 and INS1). In response to cytokine cocktail,



I κ B was rapidly reduced (up to 90%) relative to levels of extracellular-signal-regulated kinase 1/2 (ERK1/2) (Fig. 3a, b). In each case, WA prevented the reduction of I κ B, indicating that it inhibits NF- κ B activation in human islets and beta cell lines. Furthermore, immunoblot analysis showed that WA increased ERK1/2 activation and had little effect on c-jun N terminal kinase and p38 (ESM Fig. 5). We evaluated DNA binding and *NOS2* mRNA levels, a cytotoxic gene product of the NF- κ B pathway encoding inducible nitric oxide synthase (iNOS). WA significantly inhibited both binding of NF- κ B to the DNA promoter region for iNOS (Fig. 3c) and mRNA levels (Fig. 3d). These results showed that WA had an impact on NF- κ B signalling.

NF- κ B translocation in human beta cells is inhibited by WA We evaluated NF- κ B nuclear localisation at 24 h by immunohistochemical staining of the p65 subunit. All insulin-positive cells in human islets were counted and

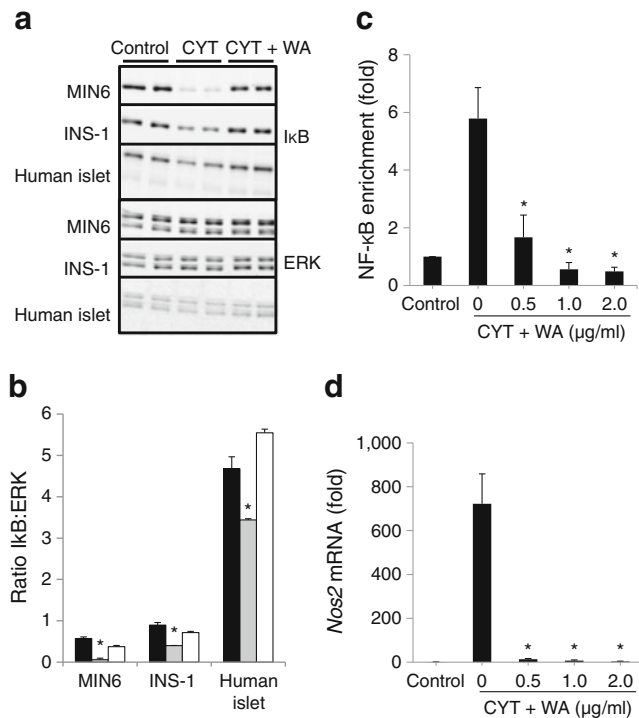


Fig. 3 WA prevents I κ B degradation and subsequent association of NF- κ B with iNOS promoter in human islets. (a) Immunoblot analysis of I κ B protein in human islets and beta cell lines treated with a cytokine cocktail (CYT) for 20 min in the presence or absence of WA. Experiments were repeated three times and performed in duplicate. (b) Graphical representations are expressed as means \pm SD. Black bar, control; grey bar, cytokine treated; white bar, cytokine plus 1 μ g/ml WA. * p <0.05 compared with control. (c) Chromatin immunoprecipitation analysis of fold increase vs control NF- κ B associated with the iNOS promoter in human islets treated with CYT for 30 min. Cells were pretreated with 0–2 μ g/ml WA. * p <0.05 compared with CYT. (d) mRNA levels (fold vs control) of *NOS2* in human islets treated under the same conditions were analysed with quantitative PCR standardised to control levels. * p <0.05 compared with CYT. Statistically significant differences in mean values was determined by Tukey–Kramer post hoc tests

when NF- κ B was stained in the nucleus it was classified as NF- κ B⁺ and nuclei without NF- κ B were classified as NF- κ B⁻ (Fig. 4a). Following addition of cytokines, NF- κ B⁺ nuclei doubled to 41.3 \pm 13.5% when compared with control (21.6% \pm 5.3%, Fig. 4b). Treatment with 0.5 μ g/ml WA (19.5 \pm 8.1% NF- κ B⁺) returned NF- κ B activation to control levels. Adding 1.0 μ g/ml WA to human islets exposed to the cytokine cocktail reduced NF- κ B translocation fourfold (10.0 \pm 4.1% NF- κ B⁺) compared with cytokine treatment alone. There was a significant difference (p <0.05) between the cytokine- and WA-treated groups.

Increased MCP-1, IP-10, iNOS, and TF production in human islets is reduced by WA We studied the inflammatory response in human islets by immunohistochemical staining for IP-10, MCP-1, iNOS, tissue factor (TF) and insulin to confirm inhibition by WA in beta cells. Staining and micrograph capture conditions (exposure time, etc.) for each treatment group

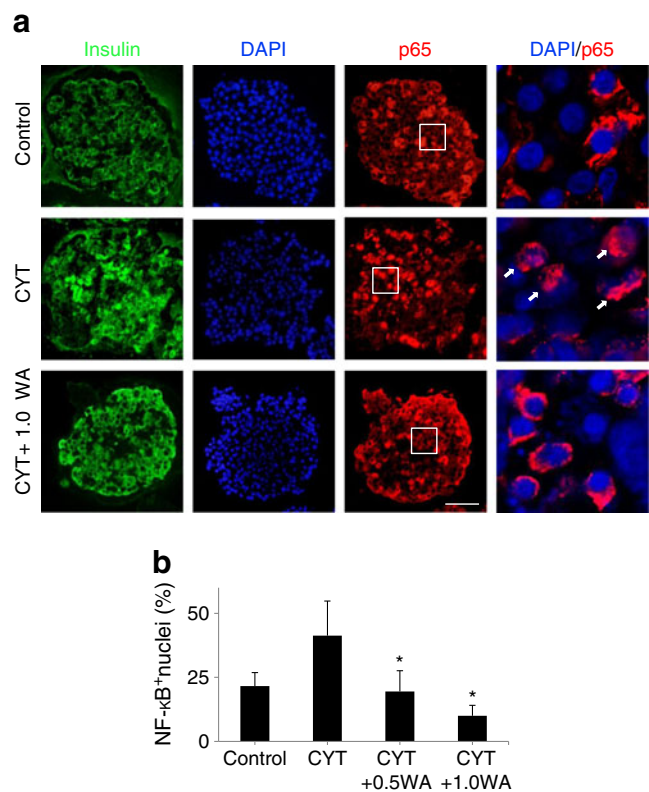


Fig. 4 NF- κ B nuclear translocation is inhibited by WA. (a) Human islets were exposed to the cytokine cocktail (CYT) with or without 1.0 μ g/ml WA for 24 h. Sections were stained for insulin (green), NF- κ B p65 (red) and for nuclei (blue). Merged nuclear and NF- κ B staining revealed translocation with overlap of nuclei and p65 (indicated by white arrows). (b) Manual counting of at least 25 beta cell nuclei in ten islets for NF- κ B translocation revealed that WA significantly inhibits NF- κ B activation down to control levels, and that 1.0 μ g/ml WA decreases NF- κ B translocation to below control levels. Results shown are representative of three independent human islet preparations. Graphical representations are expressed as means \pm SD. * p <0.05 compared with CYT as determined by Tukey–Kramer post hoc tests

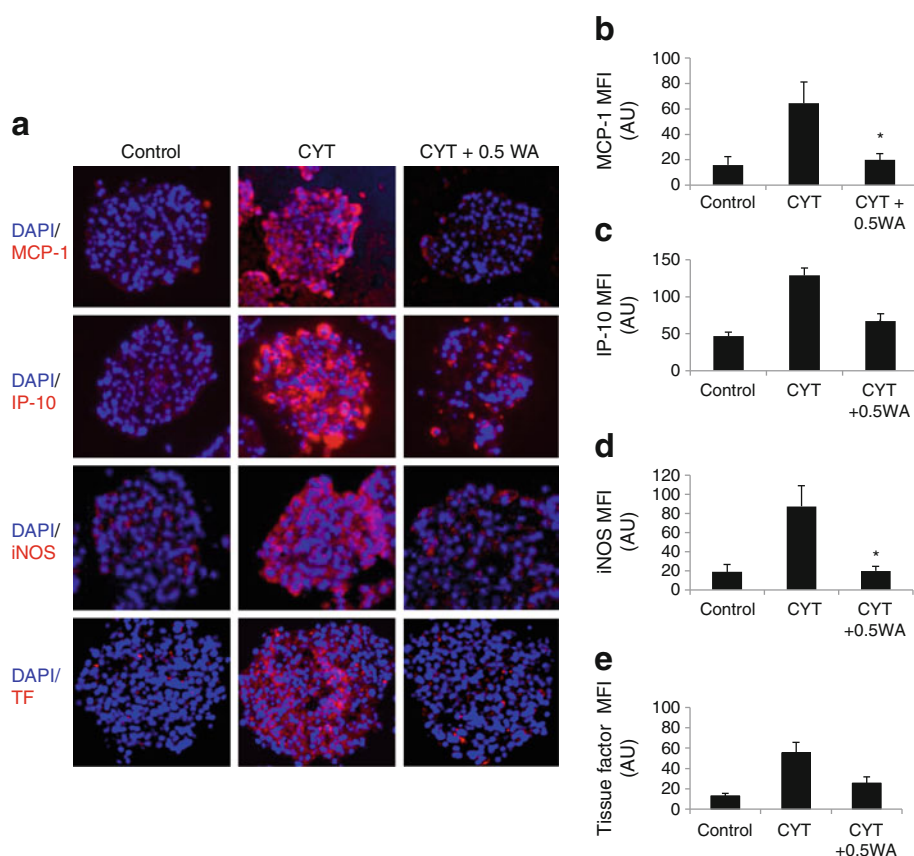
were held constant, as described previously [26]. Treatment with cytokines for 48 h increased levels of MCP-1, IP-10, iNOS and TF (Fig. 5) in insulin-positive cells when compared with control islets, but these raised levels were significantly decreased by 0.5 $\mu\text{g/ml}$ WA in each case ($p < 0.05$).

To further support a direct role of NF- κB in preventing an inflammatory response in human islets, we used the highly selective inhibitor of IKK β , BMS-345541, in cytokine induction of inflammation-related genes [7]. In human islets, inflammatory genes (including those encoding TNF- α , iNOS and IP-10) were all activated in response to cytokines (ESM Fig. 6). Induction of each of these genes was blocked in human islets when pretreated with BMS-345541. This data showed that although other signalling pathways are activated in islets by cytokines, NF- κB signalling is a requirement for activation of genes involved in inflammation.

Inflammatory cytokine and chemokine secretion by islets is inhibited by WA Previous studies have shown that NF- κB is implicated in the transcription of several inflammatory molecules [32–35]. These cytokines and chemokines also play a strong role in immune cell chemotaxis leading to poor engraftment. Cell culture supernatant fractions from human

islets exposed to cytokines and WA (0.5 or 1.0 $\mu\text{g/ml}$) were evaluated by a multiplex bead assay using xMAP technology ($n = 5$ independent experiments). IP-10 levels (Fig. 6a) were increased almost 6,000-fold by cytokine treatment ($6,516 \pm 1,497.3$ pg/islet) from control levels (1.12 ± 1.01 pg/islet). WA (1.0 $\mu\text{g/ml}$) decreased IP-10 to $2,915.3 \pm 356.1$ pg/islet in cytokine-treated islets. MCP-1, which is constitutively produced by islets [8], was marginally increased from $1,052.9 \pm 644.0$ pg/islet to $1,538.5 \pm 427.3$ pg/islet by cytokine treatment (Fig. 6b). WA treatment reduced MCP-1 levels to below control at 0.5 (951.0 ± 333.6 pg/islet) and 1.0 $\mu\text{g/ml}$ (805.6 ± 217.8 pg/islet) in cytokine-treated islets. Pro-inflammatory cytokines caused a six- to tenfold increase in islet basal levels of IL-6, IL-8, macrophage inflammatory protein (MIP)-1 α , granulocyte-colony stimulating factor (G-CSF) (Fig. 6c–f), IL-1 α and MIP-1 β (data not shown). Addition of WA led to a dose-dependent reduction in levels of these inflammatory molecules, with inhibition ranging from 71% to 88%; 1.0 $\mu\text{g/ml}$ WA significantly decreased the cytokine-stimulated levels ($p < 0.05$). This multiplex experiment was repeated with mouse islets, demonstrating dose-dependent inhibition of cytokine secretion (ESM Fig. 7).

Fig. 5 Inflammatory cytokine production and secretion from islets is decreased by WA treatment. (a) Human islets were exposed to the cytokine cocktail (CYT) with or without WA for 48 h. Sections were probed for insulin (not shown), MCP-1 (top, red), IP-10 (second, red), iNOS (third, red), TF (bottom, red) and DNA (DAPI, blue). Intracellular staining revealed higher levels of MCP-1 (b), IP-10 (c), iNOS (d), and TF (e) in islets exposed to cytokines, whereas these levels were significantly decreased by adding 0.5 $\mu\text{g/ml}$ WA. Graphical representations are expressed as averages of mean fluorescence intensity (MFI) as arbitrary units (AU) \pm SD. * $p < 0.05$ compared with CYT as determined by Tukey–Kramer post hoc tests. Scale bar, 50 μm



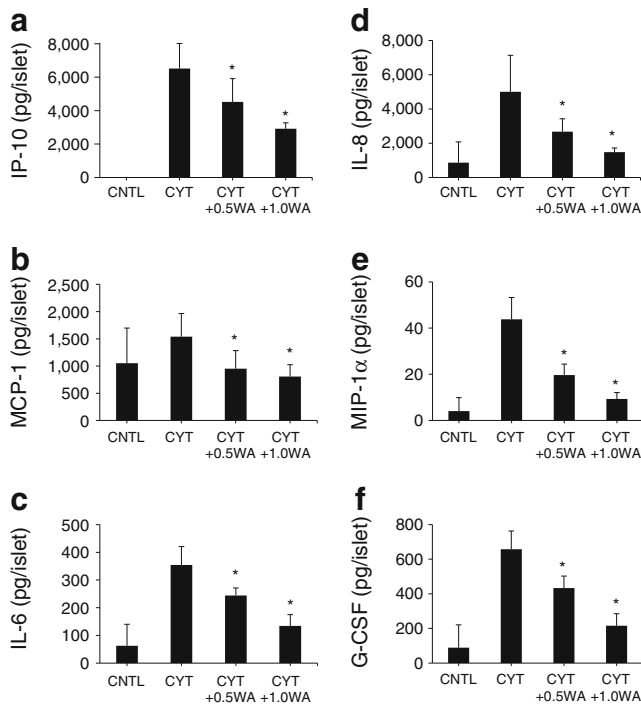


Fig. 6 WA inhibits a multitude of inflammatory cytokines and chemokines. Quintuplets of 50 handpicked human islets were cultured for 96 h with cytokines or cytokines supplemented with WA at 37°C. The supernatant fraction was collected and analysed in duplicate with Millipore's human cytokine detection kit in the Luminex 200. Results were standardised to the number of islets. Results shown are representative of three independent human islet preparations. Graphical representations are expressed as means \pm SD. * p <0.05 compared with CYT as determined by Tukey–Kramer post hoc tests

To test the inhibitory effect of WA on islets alone or other inflammatory cells, we tested human neutrophil infiltration into purified human islets using an in vitro model [31]. WA was added to either human blood alone or purified islets alone and subsequently islets and blood were mixed for analysis of neutrophil infiltration. Under both conditions, significant inhibition of neutrophil infiltration was observed (ESM Fig. 8) indicating that WA can inhibit IL-8 (neutrophil chemoattractant) in different cell types. Resveratrol, a known NF- κ B blocker, was also tested in this experiment and showed weaker inhibition when compared with WA.

Discussion

One of the major hurdles to improving the current outcomes in clinical islet transplantation is the poor engraftment of transplanted islets. During the early post-transplant period, a major inflammatory response, termed IBMIR, which includes several pro-inflammatory cytokines, exerts a damaging effect on islets resulting in the loss of up to 75% of transplanted cell mass [36]. We attempted to abrogate the

detrimental effects of pro-inflammatory cytokines using WA, which is known to influence several key signalling molecules including NF- κ B [11]. Intraportal transplantation of syngeneic islets in diabetic C57BL/6 mice, a model representing inflammatory response to transplanted islets, showed that the engraftment of islets is significantly improved by WA. While several reports have shown that WA inhibits the NF- κ B pathway through its binding to NEMO/IKK β complex [14, 15], no study has been undertaken on the use of WA to protect islets from inflammatory damage in islet transplants into diabetic mice. Despite significant work on islet reaction to inflammation, there has been little research into the contribution of islets to the immediate immune reaction. Hence we performed mechanistic studies to document inhibition of NF- κ B and pro-inflammatory cytokines in human and mouse islets by WA. Activation of NF- κ B in beta cells and islets by a cytokine cocktail was strongly inhibited by WA. This was demonstrated by the prevention of I κ B degradation, NF- κ B nuclear translocation and NF- κ B binding to DNA transcription sites as judged by p65 translocation, western blotting and DNA binding to a probe for the NF- κ B consensus region of the iNOS promoter region. WA treatment led to significant inhibition of the production and secretion of inflammatory and chemotactic cytokines in islets as determined by immunohistochemistry and multiplex analysis. We have demonstrated that treatment with WA leads to significant improvement in islet survival and function in both human and mice islets under in vitro conditions. The in vitro results correlated with the in vivo data using the C57BL/6 mouse model showing improved achievement of normoglycaemia using marginal mass syngeneic islet transplants and inhibition of p65 translocation post-transplant.

Furthermore, we showed that induction of inflammatory cytokines, such as TNF α , iNOS and IP-10, in human islets is significantly inhibited by the IKK β specific inhibitor BMS-345541. These results indicate a prominent role for NF- κ B in cytokine-mediated damage to islets, which is prevented by WA. All of the human islet results corroborated the data from mouse islets indicating that WA has clinically relevant applications in humans. In fact two clinical trials have been reported using WA to treat anxiety disorder and osteoarthritis [12, 13] documenting the safety of its use in clinical settings.

Other studies have shown improvement of intraportal graft survival by NF- κ B inhibition using a genetically enhanced, non-degradable I κ B protein [37, 38] but this strategy is not easily applied to the clinical setting. A recent report by McCall et al [39] showed that resveratrol, another NF- κ B inhibitor, failed to improve marginal mass engraftment of transplanted islets in mice. This report is in contrast to our current results and also data published by Chen et al [7], using BMS345541, which selectively blocks the I κ B

kinase subunit. This discrepancy could be due to specificity of the compounds in their inhibition of target proteins in the NF- κ B pathway.

Previous studies have shown that WA can be used without causing damage to treated cells at concentrations similar to those used in this investigation [40, 41]. NF- κ B translocation and inhibition data conformed to the results of cytokine studies of WA in pulmonary epithelial cells (A549) and cystic fibrotic KKLEB cells [16, 41]. Inactivation of protein kinase B (Akt) along with NF- κ B inhibition murine macrophage cells has also been reported [42]. Effects on mitogen-activated kinases by WA were consistent with previous studies showing that, through the MAPK/ERK (MEK/ERK) pathway, WA induces IKK β phosphorylation with I κ B phosphorylation and degradation in mouse fibrosarcoma cells [14]. Secretion and localisation of chemokines, such as MCP-1, has been previously demonstrated in MCP-1 knockout mice and human studies [8, 9]. However, MCP-1 is likely only one of many inflammatory molecules that contribute to immediate graft failure. Therefore, this study not only identifies additional cytokines and chemokines involved in inflammation (MCP-1, IP-10, IL-6, IL-8, G-CSF, MIP-1 α , MIP-1 β and IL-1 α), but also demonstrates specific, potent inhibition of each by WA.

A recent study of an IKK- β inhibitory agent found that the same set of cytokines did not affect cell death in islets under conditions favouring normal NF- κ B activation but only under conditions of reduced NF- κ B activation and hypoxia [7]. However, several other reports support our findings that cytokines cause cell death in islets and beta cells following activation of NF- κ B [43]. Other reports concur with our findings that NF- κ B inhibition prevents apoptosis and islet damage when islets are exposed to cytokines, using a degradation-resistant I κ B α transgenic mouse and islet model [37, 44]. Since NF- κ B activation is involved in a broad range of biological processes, including cell survival, stress responses and maturation of various cell types, the present data is inadequate to confirm that NF- κ B inhibition alone is sufficient to protect islets from cytokine-induced apoptosis [45]. Oxygen free radicals (such as those produced from cytokine-stimulated iNOS) play a role in cytokine-mediated damage [46]. WA has been shown to exhibit antioxidant properties, which could protect islets from oxidative damage in addition to iNOS specific inhibition [11]. As there are many factors that potentially contribute to early graft loss, WA could have other effects that lead to graft survival (e.g. effects on TF levels, immune cell activation and neutrophil infiltration). Our preliminary findings indicate that WA can inhibit key aspects of IBMIR such as neutrophil infiltration [35] and TF expression.

The advantage of WA is that it could easily be incorporated into an immunosuppressive regimen for effective inhibition of inflammatory response. Recent clinical studies in islet

transplantation have used etanercept, a soluble receptor antagonist to TNF- α , alone [47] or in combination with anakinra (a soluble receptor antagonist to IL-1 β [19]), to improve islet engraftment using marginal mass islet transplants. The improved outcome of clinical transplants with the combined use of etanercept and anakinra were further corroborated in marginal mass human islet transplants in immunodeficient mice [48]. Taken together, these reports emphasise the need to control inflammatory cytokines during the early transplant period to improve islet engraftment. The present study demonstrated that WA can profoundly inhibit the deleterious action of pro-inflammatory cytokines as well as the production of several chemotactic and inflammatory molecules by islets. Due to such protective effects, more islets could survive leading to euglycaemia in marginal islet infusions. The transplanted graft mass could be enhanced leading to prolonged insulin independence. This implication has significant ramifications for islet transplantation where it is often difficult to achieve sustained insulin independence with one donor pancreas.

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Contribution statement JAS wrote the manuscript and researched data. TI researched data, designed part of the study and contributed in equal amount to a majority of the research. MAK researched data and contributed to the discussion. KS researched data and provided aid with animal studies. HP researched data and contributed to the discussion. SM researched data and contributed to the discussion. MFL interpreted data. ML researched data and contributed to the discussion. BN contributed to the conception and design of the study and the discussion. All authors were involved in drafting or revision of the manuscript and gave final approval for submission.

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