Inflammatory Mechanisms in the Regulation of Insulin Resistance

Herbert Tilg and Alexander R Moschen

Christian Doppler Research Laboratory for Gut Inflammation and Department of Gastroenterology and Hepatology, Innsbruck Medical University, Innsbruck, Austria

Insulin resistance (IR) plays a key role in the pathophysiology of obesity-related diseases such as type 2 diabetes and nonalcoholic fatty liver disease. It has been demonstrated that IR is associated with a state of chronic low-grade inflammation, and several mediators released from various cell types, including immune cells and adipocytes, have been identified as being involved in the development of IR. Among those are several pro-inflammatory cytokines such as tumor necrosis factor- α (INF- α), interleukin (IL)-1, IL-6, and various adipocytokines. Furthermore, several transcription factors and kinases such as c-Jun N-terminal kinase (JNK) and inhibitor of kappa B kinase- β (IKK β), a kinase located proximal of nuclear factor- κ B (NF- κ B), participate in this process. Hepatocyte-specific overexpression of NF- κ B is associated with IR and can mimic all features of fatty liver disease. Whereas the evidence for an important role of many pro-inflammatory pathways in IR in in vitro and animal studies is overwhelming, data from interventional studies in humans to prove this concept are still minor. As a complex network of inflammatory cytokines, adipocytokines, transcription factors, receptor molecules, and acute-phase reactants are involved in the development of IR, new therapeutic approaches in IR-related diseases will be based on a better understanding of their complex interactions.

Online address: http://www.molmed.org doi: 10.2119/2007-00119.Tilg

INTRODUCTION

The number of obese and overweight individuals has risen dramatically over the last two decades. Obesity not only is associated with the development of type 2 diabetes (T2D) and hypertension but also has negative effects on liver function, leading to diseases such as nonalcoholic fatty liver disease (NAFLD). Insulin resistance (IR) is the key primary defect underlying the development of T2D and is a central component defining the metabolic syndrome, a constellation of abnormalities including obesity, hypertension, glucose intolerance, and dyslipidemia. IR has been defined in the last decade as being frequently associated with a state of low-grade inflammation, and therefore it is assumed that inflammation contributes in a major way to its development (1,2).

Besides the fact that IR is characterized by complex interactions between genetic determinants, nutritional factors, and lifestyle, it is increasingly recognized that mediators synthesized from cells of the immune system as well as by adipose tissue are critically involved in the regulation of insulin action (3). The details of insulin receptor signaling pathways are not presented here because of limited space (4). Briefly, insulin acts in all cells by binding to its specific receptor and thereby activating a cascade of intracellular signaling events. It stimulates tyrosine phosphorylation of insulin receptor substrate (IRS) proteins, which is a crucial event in mediating insulin action (4).

Address correspondence and reprint requests to Herbert Tilg, Christian Doppler Research Laboratory for Gut Inflammation and Department of Gastroenterology and Hepatology, Medical University Innsbruck, Anichstrasse 35, 6020 Innsbruck, Austria. Phone: +43-512-504-23374. Fax: +43-512-504-6723374; E-mail: herbert.tilg@i-med.ac.at. Submitted November 14, 2007; accepted for publication January 18, 2008; Epub (www.

submitted November 14, 2007; accepted for publication January 18, 2008; Epub (www. molmed.org) ahead of print January 23, 2008. This step in insulin-receptor signaling is one of the key molecular events in inflammation-associated IR (2). Serine phosphorylation of IRS-1 by various inflammatory signals thereby seems to be one of the key aspects that disrupt insulin-receptor signaling (2). This article summarizes current knowledge on involved inflammatory mediators in the process of IR (Table 1).

INSULIN RESISTANCE AND UNDERLYING INFLAMMATORY MECHANISMS

Cytokines and Chemokines

TNF-α. The first link between obesity increase in the expression of a proinflammatory cytokine, namely TNF-α and insulin action came from a study almost 15 years ago (5). These findings led to the concept of inflammation in obesity and demonstrated that adipocytes express TNF-α. In these studies, expression of this cytokine in obese animals (*fa/fa* rat and *ob/ob* mouse) was increased and shown to regulate insulin action (5). Further evidence supporting a

Table 1. Mediators of immune cells and adipocytes involved in the regulation of insulin resistance.

Cytokines and chemokines	Adipocytokines	Transcription factors	Others
TNF-α (5,6)	Adiponectin (33,36,42)	NF-κΒ/ΙΚΚβ (74,77,78)	Osteopontin (104)
ΙΔ-1α/β (12,14)	Leptin (48-50)	JNK-1 (79,80)	SAA (127)
Gp130 family (IL-6, CNTF) (18,19,24)	Resistin (55,56,59)	PPARγ (84,86)	CRP (96,99,101)
IL-10 (128,129)	PBEF/nampt/visfatin (61-64)	SREBP-1c (92-95)	FABP-4 (102,103)
IL-18 (27-29)	RBP-4 (68,70)	LXR (87,89-91)	Oxidative stress (108-110)
MCP-1 (30,32)			ER stress (82,83)
ΜΙΡ-1α/β (130)			iNOS (107)
RANTES (131)			E-selectin (132,133)
			P-selectin (134)
			ICAM-1 (132,135)
			VCAM-1 (107,132)
			TLR-4 (136,137)
			PKC-theta (105)

key role for TNF- α in IR came from studies published by Uysal et al., where they showed that mice lacking TNF- α or TNF receptors had improved insulin sensitivity in both dietary and genetic (ob/ob) models of obesity (6). These observations were paralleled by similar findings in humans (7), with increased adipose tissue TNF- α expression in obesity and improvement of this increased TNF-α expression following weight loss (8). At a molecular level, exposure of cells to TNF- α or elevated levels of free fatty acids (FFAs) stimulated inhibitory phosphorylation of serine residues of IRS-1 (9,10). TNF- α has certainly been "the" pro-inflammatory mediator introducing the link between inflammation, obesity, and IR.

Interleukin-1. IL-1 α and IL-1 β are among the first identified cytokines and exert strong pro-inflammatory functions (11). IL-1 $\alpha^{-/-}$ mice have lower fasting glucose and insulin levels and improved insulin sensitivity, as determined by insulin tolerance testing, compared with wild-type controls (12). IL-1 β together with IL-6 concentrations predict risk for T2D in humans better than either cytokine alone (13). IL-1 β is able to reduce IRS-1 expression at a transcriptional level through a mechanism that is ERK dependent and at a posttranscriptional level independent of ERK activation (14). By targeting IRS-1, IL-1 β is capable of impairing insulin signaling and action and could thus participate, in concert with other cytokines, in the development of IR.

IL-6 and gp130 receptor ligands. The IL-6 cytokine family (or long type I or gp130 cytokines) consists of ciliary neurotrophic factor (CNTF) as well as IL-6, IL-11, leukemia inhibitory factor, oncostatin M, and cardiotrophin 1. IL-6 signals via induction of a gp130 homodimer after binding to the IL-6 receptor (15). Most research on obesity-related aspects has centered on IL-6 and CNTF.

IL-6 was among the first cytokines implicated as a predictor or pathogenetic marker of IR and cardiovascular disease. Concentrations of IL-6 decrease in parallel with weight loss and improvement of IR in patients undergoing bariatric surgery (16). Visceral fat has been demonstrated as an important site for IL-6 secretion in humans (17). IL-6 production in abdominal adipose tissue is at least three times higher compared with subcutaneous adipose tissue, thereby potentially contributing to hepatic IR. This cytokine may be indeed involved in the pathogenesis of hepatic IR, as insulin sensitivity increases in diet-induced obese mice treated with anti-IL-6 antibodies (18). IL- $6^{-/-}$ mice are insulin resistant and develop mature-onset obesity (19). However, these results were not reproducible in another IL-6^{-/-} mouse model, as those authors did not observe age-related obesity (20). This

discrepancy has been somewhat clarified by a later report, which initially described obesity in $IL-6^{-/-}$ mice (21) by demonstrating a decreased energy expenditure and thermogenesis compared with wild-type mice, which might help to explain development of obesity at an advanced age (22).

Gp130 cytokines signal through a receptor that has many similarities to leptin signaling, and leptin is known to activate signal transduction pathways that promote increased energy expenditure and insulin sensitivity (21). IL-6 has been shown (like leptin) to activate AMP-activated protein kinase (AMPK) in both skeletal muscle and adipose tissue. Consistent with activation of AMPK, IL-6 has also been shown to increase fat oxidation in vitro, ex vivo, and in humans in vivo (23). A clear answer to the role of IL-6 in IR will be possible when patients with T2D and/or IR receive treatment with an IL-6 neutralizing antibody.

Unlike IL-6, CNTF lacks a signal peptide and is therefore not secreted by cells in a classic manner. The anti-obesogenic properties of CNTF treatment were uncovered in a study of patients with amyotrophic lateral sclerosis. Whereas this approach was clinically not successful, these patients experienced involuntary weight loss (24), a phenomenon which has also been observed after IL-6 administration to humans. CNTF is able to reverse obesity-induced IR by activating skeletal muscle AMPK activity (25). This was the first report providing evidence that CNTF could act in a manner independent of central mediation.

IL-18. IL-18 is another pro-inflammatory cytokine that plays a role in septic shock, joint inflammation, and inflammatory bowel diseases (26). A constitutive intracellular pool of IL-18 exists, and release is mainly regulated by activation of a caspase-1 that cleaves pro-IL-18. Its bioactivity on the other side is under tight control of its physiologic antagonist, the IL-18 binding protein. It has been postulated that increased IL-18 concentrations observed in patients with T2D might reflect a role in the regulation of IR (27,28). Indeed, as recently elegantly demonstrated, IL-18^{-/-} mice and IL-18R^{-/-} mice had increased body weight accompanied by IR, hyperglycemia, lipid abnormalities, and atherosclerosis compared with wild-type mice (29). Intracerebral administration of recombinant IL-18 inhibited food intake and reversed hyperglycemia in these mice by activation of STAT3 phosphorylation. Increased IL-18 levels in patients with T2D might reflect either a frustrated attempt of IL-18 to counteract hyperglycemia or could also reflect resistance to this cytokine, as observed for others such as insulin or leptin. Such an explanation could also be offered for the increased IL-6 levels observed in stages of IR.

MCP-1. Adipocytes secrete various chemoattractants that attract monocytes. It has been demonstrated that obese adipose tissue exhibits increased expression of CCL2 (or MCP-1), a key factor in the recruitment of macrophages (30). These authors demonstrated that CCL2^{-/-} mice exhibited reduced macrophage infiltration in the adipose tissue and reduced IR. Conversely, they observed an increase in macrophage infiltration when CCL2 was overexpressed. Another study also revealed reduced macrophage infiltration in the adipose tissue and decreased IR in CCL2^{-/-} mice (31). In contrast, Inouve et al. (32) recently demonstrated that the absence of CCL2

in mice does not limit obesity-associated infiltration of macrophages into adipose tissue. In that study, the authors used CCL2^{-/-} mice and adipose tissue was collected for analysis of macrophage infiltration. Surprisingly, CCL2^{-/-} mice on a high-fat diet showed no reductions in adipose tissue macrophages, although they were glucose intolerant and had mildly increased plasma glucose and decreased serum adiponectin levels compared with wild-type mice. These data could suggest that CCL2 might not be the only critical mediator for adipose tissue macrophage recruitment. There are, however, several other candidates that might play a role in the recruitment of monocytes/macrophages into the adipose tissue, such as migration inhibitory factor (MIF) or macrophage inflammatory protein-1 α (MIP-1 α).

Adipocytokines

Adiponectin. Adiponectin is mainly synthesized by adipocytes and to a small degree by other cells (33,34). It exists both as a full-length protein as well as a proteolytic cleavage fragment, also known as globular adiponectin. Adiponectin circulates at high concentrations in human serum (5 to 10 μ g/mL) and has a wide spectrum of biological activities (33). Serum levels of adiponectin are reduced in individuals with visceral obesity and states of IR (35). TNF- α suppresses the transcription of adiponectin in adipocytes, which might explain the lower adiponectin levels in serum in individuals who are obese (36). Weight loss induces adiponectin synthesis (37), as activation of peroxisome-proliferatoractivated receptor y (PPARy) by its ligands thiazolidinediones (TZDs), which are used in the treatment of T2D (38).

Initial studies suggested that adiponectin exerted anti-inflammatory effects on endothelial cells through the inhibition of TNF- α induced adhesionmolecule expression (39) and inhibited NF- κ B activation. In obese animals, treatment with adiponectin decreases hyperglycemia and levels of FFAs in the plasma and improves insulin sensitivity (33). Furthermore, adiponectin-deficient mice develop diet-induced IR on a highfat, high-sucrose diet (36). In other studies with adiponectin-deficient mice, however, these animals developed insulin resistance only if fed a high-fat diet (40) or failed to develop insulin resistance even when fed a high-fat diet (41).

Two receptors for adiponectin have been identified (adipoR1 and adipoR2). AdipoR1 is widely expressed in mice, whereas adipoR2 is mainly expressed in the liver (42). The importance of targeted disruption of adipoR1 and R2 has recently been demonstrated (43). Disruption of both receptors abolished adiponectin binding and actions, resulting in increased triglyceride content, inflammation, and oxidative stress, thus leading to IR and marked glucose intolerance. These studies together strongly support a major role for adiponectin in regulating insulin sensitivity.

Kim et al. (44) have recently presented an exciting work demonstrating that expansion of adipose tissue could also be associated with an improved metabolic profile. In their studies, they created a mouse lacking leptin and overexpressing adiponectin. Importantly, in these mice, despite being severely obese, the increase in circulating full-length isoform of adiponectin resulted in a reversal of the diabetic phenotype of ob/ob mice with normalization of glucose and insulin levels. In this model, a massive expansion of subcutaneous adipose tissue mass was associated with a modest two- to threefold elevation of steady-state adiponectin levels in the plasma. Interestingly, macrophage infiltration into expanded adipose tissue was quite minimal. The mechanism of action of TZDs relies on the ability of their ligands to reduce hepatic lipid content and induction of adiponectin. Previous studies and this report fully support the notion that the potent antisteatotic effect of adiponectin in the liver reduces liver fat content, increases subcutaneous fat mass, and improves IR (44,45).

Leptin. The discovery of leptin and the leptin receptor, the latter of which has

both a long, full-length form (OBRb) and a short, truncated form, led to the hope that researchers had identified a highly effective molecule and/or pathway that could be targeted in the treatment of obesity (46). However, it soon became evident that obesity (a situation associated with high circulating leptin levels) resulted in leptin resistance in the central nervous system (CNS) where endogenous leptin was no longer effective (leptin resistance). This phenomenon, although not completely understood, has been linked to a decreased uptake of leptin into the CNS. Another potential mechanism for this resistance has been elevated suppressor of cytokine signaling (SOCS) protein expression, which occurs in both obese humans and rodents. SOCS3 binds to the leptin receptor and to phosphorylated JAK protein. This inhibits STAT from binding to the leptin receptor and getting phosphorylated/ activated. SOCS3 competes with Src homology-containing tyrosine phosphatase 2 (SHP-2) of the same phosphor-site on the receptor (46,47).

In addition to its well-defined role in energy balance, leptin has important effects on glucose homeostasis. First, leptin is able to reverse hyperglycemia in *ob/ob* mice before body weight is corrected (48). It also improves glucose homeostasis in lipodystrophic mice and in humans with lipodystrophy or congenital leptin deficiency (49,50). Importantly, however, leptin failed to correct hyperglycemia in patients with obesity, further supporting the concept of "leptin resistance" in these patients (51). The glucose-lowering effects of leptin are mediated through different organs. Leptin improves insulin sensitivity in muscle by reducing intramyocellular lipid levels and activating AMPK (52). Leptin also improves insulin sensitivity in the liver. As in muscle, leptin decreases intracellular hepatic triacylglycerol levels (53). There might also be a direct interaction with insulin metabolism, as leptin inhibits insulin release (54). Altogether, data clearly support a role for leptin in the regulation of glucose homeostasis.

Resistin. Resistin has been implicated in the pathogenesis of obesity-associated insulin resistance and T2D in mouse models (55), whereas such a role in humans is under debate (56). Although a clear function for resistin in humans is still lacking, its pro-inflammatory properties indicate a role in inflammatory processes (57). Resistin and adiponectin have reciprocal effects on vascular endothelial cells: resistin induces the expression of VCAM1, ICAM1, and pentraxin 3, whereas adiponectin downregulates the expression of these molecules (58). Muse et al. (59) recently showed that intrahyophthalmic resistin results in increased hepatic IR, which was associated with increased expression of TNF- α , IL-6, and SOCS-3 in the liver. This observation not only provides a new link how resistin might affect IR but also again demonstrates the role of the CNS in these metabolic processes.

Pre-B cell colony-enhancing factor/nampt/visfatin. PBEF was originally cloned by Samal et al. (60) in search of novel cytokine-like molecules secreted from human peripheral blood lymphocytes. They described a 52-kDa secreted molecule termed pre-B cell-enhancing factor (PBEF) that was strongly induced by pokeweed mitogen and cycloheximide, and enhanced the effect of IL-7 and stem cell factor on pre-B cell colony formation. Intracellular PBEF acts as a dimeric type II phosphoribosyltransferase (nicotinamide phosphoribosyltransferase, nampt) (61). PBEF has been rediscovered recently and was demonstrated to be a novel adipocytokine more abundantly expressed in visceral compared with subcutaneous fat. It was therefore renamed "visfatin," a protein associated with IR in animal models of IR. Visfatin was shown to mimic insulin activity by binding to the insulin receptor (62). Notably, very recently these results published by Fukuhara et al. have been questioned and the paper has been retracted (63). We found primarily proinflammatory activities for this mediator by showing that it dose-dependently upregulated the production of the pro-inflammatory cytokines IL-1 β , IL-6, and TNF- α in human monocytes and in mice challenged with recombinant visfatin (64). Several human studies so far have not demonstrated a convincing association of this pro-inflammatory mediator with IR (65–67).

Retinol-binding protein 4. Serum retinol-binding protein 4 (RBP4) is another characterized adipocytokine (68). Until recently, the function of RBP4 was thought to be the delivery of retinol to tissues. However, in patients with T2D, serum levels of RBP4 are increased (69). Transgenic overexpression of human RBP4 or injection of recombinant RBP4 in normal mice causes IR (68). Therefore, lowering RBP4 could be an interesting strategy for the treatment of individuals with T2D. There is now more evidence that RBP4 might be associated with obesity-related disorders and IR (70–73).

Transcription Factors

Role of the IKKβ/NF-κB pathway. In searching for mechanisms involved in cytokine-induced IR, Yuan et al. (74) identified the IKKβ pathway as a target for TNF- α induced IR. Yin et al. (75) demonstrated in 1998 that aspirin and salicylates inhibit the activity of IKKβ. William Ebstein suggested 130 years ago (76) that high doses of salicylates lower high blood glucose concentrations. Yuan et al. demonstrated in their work that high doses of salicylates reverse hyperglycemia, hyperinsulinemia, and dyslipidemia in fa/fa rats and ob/ob mice, and overexpression of IKKß attenuates insulin signaling in cultured cells. These findings clearly demonstrated the involvement of inflammatory pathways in IR highlighting the important role of IKKβ, a proximal mediator in NF-κB activation (Figure 1).

Two groups have shown the relationship between IKK β expression in the liver and IR (77,78). Cai et al. (77) created a stage of chronic, subacute inflammation in the liver in a transgenic mouse model by selective hepatocellular activation of NF- κ B causing continuous lowlevel expression of IKK β . These mice ex-



Figure 1. Regulation of IR: involved mediators and pathways. In recent years, several inflammatory pathways involved in the generation/regulation of IR have been identified. TNF-α was among the first mediators defined as a key factor linking inflammation, obesity, and IR. Engagement of TNFR by TNF-α induces inhibitory phosphorylation of serine residues of IRS-1 and activates IKKβ/NF-κB and JNK pathways, two major intracellular regulators of IR. Moreover, TNF-α antagonizes adiponectin, an important insulin-sensitizing adipocytokine that signals via adiporeceptors. IL-1 and IL-18 are also able to induce IR. IL-1 has been shown to reduce IRS-1 expression via ERK1/2 and can activate the IKKβ/NF-κB pathway. A role for IL-18 in the regulation of IR has recently been demonstrated in IL-18^{-/-} and IL-18R^{-/-} mice. IL-6 is another cytokine involved in the generation of IR. This cytokine can induce SOCS1 and SOCS3 that link IRS to ubiquitin-mediated degradation. ER stress as well as oxidative stress are both involved in inflammation-associated IR. An important role for the IKKβ/NF-κB pathway has been demonstrated in experiments where knocking out or knocking down of IKKβ or JNK protected mice from IR.

hibited a T2D phenotype with evidence of moderate systemic IR. IR was improved by systemic neutralization of IL-6 or by oral salicylate therapy. Arkan et al. (78) recently presented similar findings in mice lacking either IKK β in hepatocytes or myeloid cells. Liver-specific deletion of IKK β resulted in relative insulin sensitivity in the liver when placed on a high-fat diet or intercrossed with the *ob/ob* model of genetic obesity, but

developed IR in muscle and fat. In contrast, mice deficient in myeloid IKK β exhibited increased insulin sensitivity and were partially protected from IR.

c-Jun N-terminal kinase (JNK). Several serine/threonine kinases are activated by inflammatory stimuli contributing to IR, including JNK, IKK, and others. Activation of these kinases takes place in situations where inflammatory and metabolic pathways are triggered, which is also seen after Toll-like receptor (TLR) activation, for example. JNK has recently emerged as an important regulator of IR in obesity (79). The JNK group belongs to the group of MAPKs and controls many cellular functions through regulation of activator protein-1 (AP-1), including c-Jun and JunB. In obesity, JNK activity is increased in the liver, muscle, and fat tissues probably owing to the increase of FFA and TNF- α , and loss of JNK1 prevents the development of IR in both genetic and dietary models of obesity. Liver-specific knockdown of JNK1 indeed lowers circulating glucose and insulin levels, proving its role in the development of IR (80).

ER stress response. Recent experimental evidence suggests that endoplasmic reticulum (ER) stress is important in the initiation and regulation of inflammation and insulin action as observed in IR (1). Folding, maturation, storage, and transport of most proteins take place in the ER. In case folding is disturbed, an unfolded protein response (UPR) is initiated to restore this organelle, which involves three key molecules: inositolrequiring enzyme-1 (IRE-1), PKR-like endoplasmic-reticulum kinase (PERK), and activating transcription factor 6 (ATF6) (81). Two important pathways in the regulation of IR that have already been discussed, namely NF-κB/IKKβ and JNK-AP-1, are linked to activation of IRE-1 and PERK (82). Indeed and as expected, ER stress is involved in both dietary and genetic models of obesity and regulation of IR (83). ATF6 and X-box binding protein-1 (XBP-1) are critical regulators of ER function and its adaptive responses, as gain- and loss-offunction studies with XPB-1 demonstrated the close interaction with insulin action in vitro and in vivo (83).

PPAR-γ. PPARγ is a genetic sensor of fatty acids and a member of the nuclear receptor superfamily of ligand-dependent transcription factors. This transcription factor is required for fat cell development and is the molecular target of TZDs, which exert insulin-sensitizing effects in adipose tissue, skeletal muscle,

and liver. TZDs also negatively regulate the stimulus-dependent production of various pro-inflammatory cytokines that promote IR. Within adipocytes, TZDs suppress the synthesis of IL-6, TNF- α , PAI-1, MCP-1, and angiotensinogen. In macrophages, where PPARy is also expressed, it inhibits TLR- and IFN-ymediated inflammatory responses. As macrophages prominently invade adipose tissue in obesity, macrophagederived PPARy recently gained considerable interest. Hevener et al. (84) presented evidence that macrophage PPARy is essential for normal skeletal muscle and liver insulin sensitivity. They found that inactivation of PPARy in macrophages led to glucose intolerance associated with skeletal and hepatic IR even in lean mice fed a normal diet. IR was even more pronounced when mice lacking macrophage PPARy were fed a high-fat diet, and these mice were only partially responsive to TZD treatment. This indeed suggests that for a proper TZD response, macrophage PPARy expression is essential.

In obesity, adipose tissue is loaded with macrophages resulting in local inflammation, thereby aggravating IR. Different types of macrophages reside in the adipose tissue (85). Whereas resident macrophages present in adipose tissue of lean mice display the alternatively activated phenotype (M2 or alternatively activated macrophages characterized by activated genes for Ym1, arginase 1, and IL-10) (85), pro-inflammatory classically activated macrophages are recruited to sites of tissue damage in the adipose tissue as in obesity (M1 or classically activated macrophages producing enhanced levels of TNF- α and iNOS). Diet-induced obesity influences the state of adipose tissue macrophages from an M2-polarized state in lean animals that protects adipocytes from inflammation to an M1 pro-inflammatory state leading to IR. This obesity-induced phenotypic switch in adipose tissue macrophage polarization has been recently demonstrated as being orchestrated by PPARy (86). Using mice with specific macrophage deletion

of PPARγ, these researchers demonstrated that PPARγ is required for the maturation of alternatively activated macrophages (M2 macrophages).

Because of space limitations, we are unable to provide an extensive overview of an important group of key mediators that also act at the interface of lipid metabolism and inflammatory pathways. These molecules include the LXR family of nuclear hormone receptors (87–91) and the SREBP family of transcription factors (92–95).

Others

Acute-phase proteins. C-reactive protein (CRP), the most important human acute-phase protein, correlates with states of IR (96). CRP in most instances is considered as an inflammatory marker related to atherosclerosis and cardiovascular diseases (97,98). Human CRP, however, might also have some antiinflammatory properties, as it reduces atherosclerosis development in a mouse model with human-like hypercholesterolemia (99). CRP has been demonstrated to upregulate the synthesis of anti-inflammatory cytokines such as IL-1 receptor antagonist (IL-Ra), which could help to explain the above phenomenon (100). Recently it has been suggested that CRP might play a role in leptin resistance by acting as a serum leptin-interacting protein (101).

Adipocyte–fatty-acid binding protein (aP2, FABP4). Fatty acid-binding proteins are a family of 14- to 15-kDa proteins that bind with high affinity to the hydrophobic ligands such as saturated and unsaturated long-chain fatty acids and eicosanoids. Adipocyte-fatty-acid binding protein, aP2 (FABP4), is expressed in adipocytes and macrophages, links inflammatory and metabolic processes, and is mainly regulated by PPARy agonists, insulin, and fatty acids. Deficiency of aP2 protects mice against the development of IR associated with genetic or diet-induced obesity (102). The macrophage is a critical site of FABP action, and total- or macrophage-specific aP2-deficiency leads to a marked protection against early and advanced atherosclerosis in apolipoprotein-deficient mice. Furuhashi et al. (103) have recently demonstrated that an orally active smallmolecule inhibitor of aP2 is an effective therapeutic agent against severe atherosclerosis and T2DM in various mouse models.

Osteopontin. Osteopontin (OPN) is a secreted matrix glycoprotein and pro-inflammatory cytokine playing an important role in cell-mediated immunity. Its ability to interact with integrin surface molecules through an Arg-Gly-Asp sequence and with the CD44 receptor has established this mediator as an important signaling molecule. Indeed, tissue infiltration of macrophages as observed in obesity is dependent on the expression of OPN, which promotes monocyte chemotaxis and motility. Recently, Nomiyama et al. (104) demonstrated that mice after a high-fat diet exhibited increased circulating OPN levels. Obese mice lacking OPN showed improved insulin sensitivity and decreased macrophage infiltration into adipose tissue. These experiments add OPN to a long list of pro-inflammatory pathways involved in the development of IR.

Protein kinase C. Protein kinase C (PKC) is also important in the interactions between inflammatory and metabolic pathways. PKCθ-knockout mice are protected from fat-induced IR (105). Three-day high-fat feeding to rats results in hepatic steatosis and hepatic IR, which is associated with activation of PKCE but not other isoforms. Treatment with an antisense oligonucleotide against PKCE protects rats from fat-induced hepatic IR. Furthermore, this treatment reversed impaired insulin signaling, supporting its role in the development of IR and suggesting another potential therapeutic pathway (106).

iNOS. Inducible nitric oxide synthase (iNOS) and its deletion is associated with improvement of high-fat diet–induced IR. Recently it has been demonstrated that blockade of iNOS by *N*(G)-nitro-L-arginine methyl ester (L-NAME) improved high-fat diet–induced obesity

and glucose intolerance. These effects were accompanied by a reduction of inflammation in the adipose tissue and improved signaling in skeletal muscle (107).

Oxidative stress. One of the final common mediators of IR seems to be oxidative stress due to generation of reactive oxygen species (ROS) and/or decreased antioxidant defenses (108). In both nonalcoholic steatohepatitis and experimental steatohepatitis, hepatic expression of CYP2E1 is increased, leading to oxidative stress. This enhanced expression has been demonstrated to impair insulin signaling (109). Further supporting the importance of ROS in NAFLD, Xu et al. (110) recently demonstrated a key role for the Nrf1 gene in NASH. Mice with liver-specific deletion of Nrf1, a gene mediating activation of oxidative stress response genes, develop all features of nonalcoholic fatty liver disease including steatosis, apoptosis, necrosis, inflammation, fibrosis, and finally liver cancerhighlighting the importance of oxidative stress in this disease.

ANTI-INFLAMMATORY STRATEGIES TO OVERCOME IR

Anti-TNF Approaches

Infliximab has been demonstrated to reverse steatosis and to improve insulin signaling in a rat model of high-fat diet-induced IR, suggesting that neutralization of this key cytokine improved not only liver inflammation/steatosis/ fibrosis but also insulin signaling (111). Importantly, so far studies using neutralizing anti-TNF antibodies in humans have not shown improved insulin sensitivity (112,113). Single doses of TNF- α antagonists failed to improve IR in diabetic or obese subjects (112,114). Subsequent placebo-controlled studies conducted over a treatment period of 4 weeks provided no improvements in insulin sensitivity in obese, diabetic subjects (115) or in obese, insulin-resistant subjects without diabetes (113). Lo et al. (116) recently demonstrated in a study in metabolic syndrome that etanercept therapy increased total adiponectin concentration, but concentrations of the high-molecular-weight form—which is thought to mediate insulin sensitivity were unchanged. Another recently published study investigated the effect of adalimumab on IR in rheumatoid arthritis patients. These patients with active disease showed marked IR that was not influenced by anti-TNF therapy despite a reduction in systemic inflammation (IL-6, CRP serum levels) during the treatment (117). Thus, at the moment TNFα blockade appears to have no efficacy on IR in humans.

Chemical Chaperones

ER stress is, as mentioned, a key link between obesity, IR, and T2D (1). Pharmaceutical chaperones such as 4-phenyl butyric acid (PBA) or endogenous bile acids and derivates such as ursodeoxycholic acid (UDCA) (including its taurine-conjugated derivates [TUDCA]) are able to modulate ER function and its folding capacity. Indeed, Ozcan et al. (118) have recently demonstrated that treatment of obese and diabetic mice improved hyperglycemia, insulin sensitivity, and fatty liver disease. Interestingly, insulin activity was increased in all target organs such as liver, muscle, and adipose tissues. It has to be mentioned that concentrations used-for example, for UDCA—were approximately ten times higher (118) than those achieved in humans and treatment of patients with nonalcoholic steatohepatitis with UDCA so far has not been successful (119).

Salicylates

The glucose-lowering effects of salicylates were identified more than 130 years ago by William Ebstein (76). Whereas these effects might be mainly attributed to its effect on NF- κ B inhibition (74), recent studies have also demonstrated that that high-dose aspirin (up to 7 g/day) improves glucose tolerance and triglyceride levels (120). These positive effects of high-dose aspirin on IR are, however, limited by its toxicity profile, especially in the gastrointestinal tract. Nonacetylated salicylates with fewer side effects such as salsalate also inhibit NF- κ B through direct inhibition of IKK β (75). These compounds do not prolong bleeding time, and trials investigating the effects of this drug on inflammation and IR have been initiated in patients with T2D.

Statins and Thiazolidinediones

As inflammation is considered crucial in the pathogenesis of IR, several other drugs commonly used in this patient population are studied in this direction. Randomized clinical trials with statins have demonstrated reductions in CRP and cytokine levels, although statins are not able to improve IR or glycemia (121,122). The PPARy ligands pioglitazone or rosiglitazone are used to improve insulin sensitivity in patients with T2D. As macrophages are of key importance in the pathogenesis of IR, targeting PPARy in these cells might represent one major mechanism of these powerful drugs. Recently some caution on this drug class has been raised, as metaanalyses demonstrated an increased rate of cardiovascular events in certain patient groups (123,124).

IL-1 Receptor Antagonist

IL-1Ra is markedly upregulated in the serum of obese patients, is correlated with BMI and IR, and is overexpressed in the white adipose tissue of obese humans (125). Treatment of T2D patients with recombinant human IL-1Ra improves glycemic control, clearly highlighting the role of inflammation in T2D and IR (126). This is the first and important evidence that an anti-inflammatory strategy might indeed improve glycemic control and IR.

CONCLUSIONS

It became evident in the last year in many in vitro and animal studies that various pro-inflammatory cytokines, adipocytokines, and transcription factors are critically involved in the pathogenesis of IR. This intriguing concept is also supported by many clinical observations in patients with T2D and NAFLD, where IR is correlated with a state of low-grade chronic inflammation. Based on these findings, scientists have started to examine whether anti-inflammatory strategies in humans are able to revert IR, and thereby are demonstrating that indeed inflammation significantly drives the development of IR. These studies, however, are still in their infancy, and currently we have only limited data in hand showing that anti-inflammatory strategies may improve IR.

ACKNOWLEDGMENTS

This work was supported by the Christian Doppler Society and a grant from the Austrian Science Foundation (P17447).

REFERENCES

- Hotamisligil GS. (2006) Inflammation and metabolic disorders. *Nature* 444:860–7.
- Wellen KE, Hotamisligil GS. (2005) Inflammation, stress, and diabetes. J. Clin. Invest. 115:1111–9.
- Rosen ED, Spiegelman BM. (2006) Adipocytes as regulators of energy balance and glucose homeostasis. *Nature* 444:847–53.
- Pirola L, Johnston AM, Van Obberghen E. (2004) Modulation of insulin action. *Diabetologia* 47:170–84.
- Hotamisligil GS, Shargill NS, Spiegelman BM. (1993) Adipose expression of tumor necrosis factor-alpha: direct role in obesity-linked insulin resistance. *Science* 259:87–91.
- Uysal KT, Wiesbrock SM, Marino MW, Hotamisligil GS. (1997) Protection from obesity-induced insulin resistance in mice lacking TNF-alpha function. *Nature* 389:610–4.
- Kern PA, Saghizadeh M, Ong JM, Bosch RJ, Deem R, Simsolo RB. (1995) The expression of tumor necrosis factor in human adipose tissue: regulation by obesity, weight loss, and relationship to lipoprotein lipase. *J. Clin. Invest.* 95:2111–9.
- Dandona P, Weinstock R, Thusu K, Abdel-Rahman E, Aljada A, Wadden T. (1998) Tumor necrosis factor-alpha in sera of obese patients: fall with weight loss. J. Clin. Endocrinol. Metab. 83:2907–10.
- Paz K, Hemi R, LeRoith D, Karasik A, Elhanany E, Kanety H, Zick Y. (1997) A molecular basis for insulin resistance: elevated serine/threonine phosphorylation of IRS-1 and IRS-2 inhibits their binding to the juxtamembrane region of the insulin receptor and impairs their ability to undergo insulin-induced tyrosine phosphorylation. *J. Biol. Chem.* 272:29911–8.
- Aguirre V, Uchida T, Yenush L, Davis R, White MF. (2000) The c-Jun NH(2)-terminal kinase promotes insulin resistance during association with insulin receptor substrate-1 and phosphorylation of Ser(307). J. Biol. Chem. 275:9047–54.
- 11. Dinarello CA. (2005) Blocking IL-1 in systemic inflammation. J. Exp. Med. 201:1355–9.

- Matsuki T, Horai R, Sudo K, Iwakura Y. (2003) IL-1 plays an important role in lipid metabolism by regulating insulin levels under physiological conditions. J. Exp. Med. 198:877–88.
- Spranger J, et al. (2003) Inflammatory cytokines and the risk to develop type 2 diabetes: results of the prospective population-based European Prospective Investigation into Cancer and Nutrition (EPIC)-Potsdam Study. *Diabetes* 52:812–7.
- Jager J, Gremeaux T, Cormont M, Marchand-Brustel Y, Tanti JF. (2007) Interleukin-1beta-induced insulin resistance in adipocytes through down-regulation of insulin receptor substrate-1 expression. *Endocrinology* 148:241–51.
- Kishimoto T, Akira S, Narazaki M, Taga T. (1995) Interleukin-6 family of cytokines and gp130. *Blood* 86:1243–54.
- Kopp HP, et al. (2003) Impact of weight loss on inflammatory proteins and their association with the insulin resistance syndrome in morbidly obese patients. *Arterioscler. Thromb. Vasc. Biol.* 23:1042–7.
- Fontana L, Eagon JC, Trujillo ME, Scherer PE, Klein S. (2007) Visceral fat adipokine secretion is associated with systemic inflammation in obese humans. *Diabetes* 56:1010–3.
- Klover PJ, Clementi AH, Mooney RA. (2005) Interleukin-6 depletion selectively improves hepatic insulin action in obesity. *Endocrinology* 146:3417–27.
- Wallenius V, et al. (2002) Interleukin-6-deficient mice develop mature-onset obesity. *Nat. Med.* 8:75–9.
- Di Gregorio GB, Hensley L, Lu T, Ranganathan G, Kern PA. (2004) Lipid and carbohydrate metabolism in mice with a targeted mutation in the IL-6 gene: absence of development of age-related obesity. Am. J. Physiol. Endocrinol. Metab. 287:E182–7.
- Flier JS. (2004) Obesity wars: molecular progress confronts an expanding epidemic. *Cell* 116:337–50.
- Wernstedt I, Edgley A, Berndtsson A, Faldt J, Bergstrom G, Wallenius V, Jansson JO. (2006) Reduced stress- and cold-induced increase in energy expenditure in interleukin-6-deficient mice. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 291:R551–7.
- Febbraio MA. (2007) gp130 receptor ligands as potential therapeutic targets for obesity. J. Clin. Invest. 117:841–9.
- (1996) A double-blind placebo-controlled clinical trial of subcutaneous recombinant human ciliary neurotrophic factor (rHCNTF) in amyotrophic lateral sclerosis. ALS CNTF Treatment Study Group. Neurology 46:1244–9.
- Watt MJ, et al. (2006) CNTF reverses obesityinduced insulin resistance by activating skeletal muscle AMPK. *Nat. Med.* 12:541–8.
- Dinarello CA. (2007) Interleukin-18 and the pathogenesis of inflammatory diseases. *Semin. Nephrol.* 27:98–114.
- Zirlik A, et al. (2007) Interleukin-18, the metabolic syndrome, and subclinical atherosclerosis: results from the Dallas Heart Study. *Arterioscler. Thromb. Vasc. Biol.* 27:2043–9.

- 28. Kim HJ, et al. (2007) Effects of rosiglitazone and metformin on inflammatory markers and adipokines: decrease in interleukin-18 is an independent factor for the improvement of homeostasis model assessment-beta in type 2 diabetes mellitus. *Clin. Endocrinol. (Oxf.)* 66:282–9.
- Netea MG, et al. (2006) Deficiency of interleukin-18 in mice leads to hyperphagia, obesity and insulin resistance. *Nat. Med.* 12:650–6.
- Kanda H, et al. (2006) MCP-1 contributes to macrophage infiltration into adipose tissue, insulin resistance, and hepatic steatosis in obesity. *J. Clin. Invest.* 116:1494–505.
- Weisberg SP, et al. (2006) CCR2 modulates inflammatory and metabolic effects of high-fat feeding. J. Clin. Invest. 116:115–24.
- Inouye KE, Shi H, Howard JK, Daly CH, Lord GM, Rollins BJ, Flier JS. (2007) Absence of CC chemokine ligand 2 does not limit obesity-associated infiltration of macrophages into adipose tissue. *Diabetes* 56:2242–50.
- Berg AH, Combs TP, Scherer PE. (2002) ACRP30/ adiponectin: an adipokine regulating glucose and lipid metabolism. *Trends Endocrinol. Metab.* 13:84–9.
- Wolf AM, et al. (2006) Up-regulation of the antiinflammatory adipokine adiponectin in acute liver failure in mice. J. Hepatol. 44:537–43.
- Arita Y, et al. (1999) Paradoxical decrease of an adipose-specific protein, adiponectin, in obesity. *Biochem. Biophys. Res. Commun.* 257:79–83.
- Maeda N, et al. (2002) Diet-induced insulin resistance in mice lacking adiponectin/ACRP30. *Nat. Med.* 8:731–7.
- Bruun JM, Lihn AS, Verdich C, Pedersen SB, Toubro S, Astrup A, Richelsen B. (2003) Regulation of adiponectin by adipose tissue-derived cytokines: in vivo and in vitro investigations in humans. *Am. J. Physiol. Endocrinol. Metab.* 285:E527–33.
- Maeda N, et al. (2001) PPARgamma ligands increase expression and plasma concentrations of adiponectin, an adipose-derived protein. *Diabetes* 50:2094–9.
- Ouchi N, et al. (1999) Novel modulator for endothelial adhesion molecules: adipocytederived plasma protein adiponectin. *Circulation* 100:2473–6.
- Kubota N, et al. (2002) Disruption of adiponectin causes insulin resistance and neointimal formation. J. Biol. Chem. 277:25863–6.
- Ma K, et al. (2002) Increased beta-oxidation but no insulin resistance or glucose intolerance in mice lacking adiponectin. J. Biol. Chem. 277:34658–61.
- Yamauchi T, et al. (2003) Cloning of adiponectin receptors that mediate antidiabetic metabolic effects. *Nature* 423:762–9.
- Yamauchi T, et al. (2007) Targeted disruption of AdipoR1 and AdipoR2 causes abrogation of adiponectin binding and metabolic actions. *Nat. Med.* 13:332–9.
- Kim JY, et al. (2007) Obesity-associated improvements in metabolic profile through expansion of adipose tissue. J. Clin. Invest. 117:2621–37.
- 45. Xu A, Wang Y, Keshaw H, Xu LY, Lam KS, Cooper

GJ. (2003) The fat-derived hormone adiponectin alleviates alcoholic and nonalcoholic fatty liver diseases in mice. *J. Clin. Invest* 112:91–100.

- La Cava A, Matarese G. (2004) The weight of leptin in immunity. *Nat. Rev. Immunol.* 4:371–9.
- Howard JK, Flier JS. (2006) Attenuation of leptin and insulin signaling by SOCS proteins. *Trends Endocrinol. Metab.* 17:365–71.
- Pelleymounter MA, Cullen MJ, Baker MB, Hecht R, Winters D, Boone T, Collins F. (1995) Effects of the obese gene product on body weight regulation in ob/ob mice. *Science* 269:540–3.
- Shimomura I, Hammer RE, Ikemoto S, Brown MS, Goldstein JL. (1999) Leptin reverses insulin resistance and diabetes mellitus in mice with congenital lipodystrophy. *Nature* 401:73–6.
- Oral EA, et al. (2002) Leptin-replacement therapy for lipodystrophy. N. Engl. J. Med. 346:570–8.
- Heymsfield SB, et al. (1999) Recombinant leptin for weight loss in obese and lean adults: a randomized, controlled, dose-escalation trial. *JAMA* 282:1568–75.
- Minokoshi Y, Kim YB, Peroni OD, Fryer LG, Muller C, Carling D, Kahn BB. (2002) Leptin stimulates fatty-acid oxidation by activating AMP-activated protein kinase. *Nature* 415:339–43.
- Kamohara S, Burcelin R, Halaas JL, Friedman JM, Charron MJ. (1997) Acute stimulation of glucose metabolism in mice by leptin treatment. *Nature* 389:374–7.
- Covey SD, et al. (2006) The pancreatic beta cell is a key site for mediating the effects of leptin on glucose homeostasis. *Cell Metab* 4:291–302.
- 55. Steppan CM, et al. (2001) The hormone resistin links obesity to diabetes. *Nature* 409:307–12.
- Utzschneider KM, et al. (2005) Resistin is not associated with insulin sensitivity or the metabolic syndrome in humans. *Diabetologia* 48:2330–3.
- Kusminski CM, McTernan PG, Kumar S. (2005) Role of resistin in obesity, insulin resistance and Type II diabetes. *Clin. Sci. (Lond.)* 109:243–56.
- Kawanami D, et al. (2004) Direct reciprocal effects of resistin and adiponectin on vascular endothelial cells: a new insight into adipocytokineendothelial cell interactions. *Biochem. Biophys. Res. Commun.* 314:415–9.
- Muse ED, Lam TK, Scherer PE, Rossetti L. (2007) Hypothalamic resistin induces hepatic insulin resistance. J. Clin. Invest. 117:1670–8.
- Samal B, Sun Y, Stearns G, Xie C, Suggs S, Mc-Niece I. (1994) Cloning and characterization of the cDNA encoding a novel human pre-B-cell colony-enhancing factor. *Mol. Cell Biol.* 14:1431–7.
- Rongvaux A, Shea RJ, Mulks MH, Gigot D, Urbain J, Leo O, Andris F. (2002) Pre-B-cell colony-enhancing factor, whose expression is upregulated in activated lymphocytes, is a nicotinamide phosphoribosyltransferase, a cytosolic enzyme involved in NAD biosynthesis. *Eur. J. Immunol.* 32:3225–34.
- Fukuhara A, et al. (2005) Visfatin: a protein secreted by visceral fat that mimics the effects of insulin. *Science* 307:426–30.

- 63. Fukuhara A, et al. (2007) Retraction. *Science* 318:565.
- Moschen AR, Kaser A, Enrich B, Mosheimer B, Theurl M, Niederegger H, Tilg H. (2007) Visfatin, an adipocytokine with pro-inflammatory and immunomodulating properties. *J. Immunol.* 178:1748–58.
- 65. Takebayashi K, Suetsugu M, Wakabayashi S, Aso Y, Inukai T. (2007) Association between plasma visfatin and vascular endothelial function in patients with type 2 diabetes mellitus. *Metabolism* 56:451–8.
- Axelsson J, et al. (2007) Circulating levels of visfatin/pre-B-cell colony-enhancing factor 1 in relation to genotype, GFR, body composition, and survival in patients with CKD. Am. J. Kidney Dis. 49:237–44.
- Varma V, et al. (2007) Human visfatin expression: relationship to insulin sensitivity, intramyocellular lipids, and inflammation. *J. Clin. Endocrinol. Metab.* 92:666–72.
- Yang Q, et al. (2005) Serum retinol binding protein 4 contributes to insulin resistance in obesity and type 2 diabetes. *Nature* 436:356–62.
- Graham TE, et al. (2006) Retinol-binding protein 4 and insulin resistance in lean, obese, and diabetic subjects. N. Engl. J. Med. 354:2552–63.
- Stefan N, et al. (2007) High circulating retinolbinding protein 4 is associated with elevated liver fat but not with total, subcutaneous, visceral, or intramyocellular fat in humans. *Diabetes Care* 30:1173–8.
- Broch M, Vendrell J, Ricart W, Richart C, Fernandez-Real JM. (2007) Circulating retinol-binding protein 4, insulin sensitivity, insulin secretion and insulin disposition index in obese and nonobese subjects. *Diabetes Care* 30:1802–6.
- Gavi S, Stuart LM, Kelly P, Melendez MM, Mynarcik DC, Gelato MC, McNurlan MA. (2007) Retinol-binding protein 4 is associated with insulin resistance and body fat distribution in nonobese subjects without type 2 diabetes. J. Clin. Endocrinol. Metab. 92:1886–90.
- Haider DG, Schindler K, Prager G, Bohdjalian A, Luger A, Wolzt M, Ludvik B. (2007) Serum retinol-binding protein 4 is reduced after weight loss in morbidly obese subjects. J. Clin. Endocrinol. Metab. 92:1168–71.
- Yuan M, Konstantopoulos N, Lee J, Hansen L, Li ZW, Karin M, Shoelson SE. (2001) Reversal of obesity- and diet-induced insulin resistance with salicylates or targeted disruption of Ikkbeta. *Sci*ence 293:1673–7.
- Yin MJ, Yamamoto Y, Gaynor RB. (1998) The antiinflammatory agents aspirin and salicylate inhibit the activity of I(kappa)B kinase-beta. *Nature* 396:77–80.
- Ebstein W. (1876) Zur Therapie des Diabetes mellitus, insbesondere über die Anwendung des salicylsauren Natron bei demselben. *Berliner Klinische Wochenschrift* 13:337–40.
- 77. Cai D, Yuan M, Frantz DF, Melendez PA, Hansen L, Lee J, Shoelson SE. (2005) Local and systemic insulin resistance resulting from hepatic activa-

tion of IKK-beta and NF-kappaB. *Nat. Med.* 11:183–90.

- Arkan MC, et al. (2005) IKK-beta links inflammation to obesity-induced insulin resistance. *Nat. Med.* 11:191–8.
- 79. Hirosumi J, et al. (2002) A central role for JNK in obesity and insulin resistance. *Nature* 420:333–6.
- Yang R, et al. (2007) Liver-specific knockdown of JNK1 up-regulates proliferator-activated receptor gamma coactivator 1 beta and increases plasma triglyceride despite reduced glucose and insulin levels in diet-induced obese mice. J. Biol. Chem. 282:22765–74.
- Harding HP, Zhang Y, Ron D. (1999) Protein translation and folding are coupled by an endoplasmicreticulum-resident kinase. *Nature* 397:271–4.
- Urano F, Wang X, Bertolotti A, Zhang Y, Chung P, Harding HP, Ron D. (2000) Coupling of stress in the ER to activation of JNK protein kinases by transmembrane protein kinase IRE1. *Science* 287:664–6.
- Ozcan U, et al. (2004) Endoplasmic reticulum stress links obesity, insulin action, and type 2 diabetes. *Science* 306:457–61.
- Hevener AL, et al. (2007) Macrophage PPAR gamma is required for normal skeletal muscle and hepatic insulin sensitivity and full antidiabetic effects of thiazolidinediones. J. Clin. Invest. 117:1658–69.
- Lumeng CN, Bodzin JL, Saltiel AR. (2007) Obesity induces a phenotypic switch in adipose tissue macrophage polarization. *J. Clin. Invest.* 117:175–84.
- Odegaard JI, et al. (2007) Macrophage-specific PPARgamma controls alternative activation and improves insulin resistance. *Nature* 447:1116–20.
- Joseph SB, Castrillo A, Laffitte BA, Mangelsdorf DJ, Tontonoz P. (2003) Reciprocal regulation of inflammation and lipid metabolism by liver X receptors. *Nat. Med.* 9:213–9.
- Ide T, et al. (2004) SREBPs suppress IRS-2-mediated insulin signaling in the liver. *Nat. Cell Biol.* 6:351–7.
- Laffitte BA, et al. (2003) Activation of liver X receptor improves glucose tolerance through coordinate regulation of glucose metabolism in liver and adipose tissue. *Proc. Natl. Acad. Sci. U. S. A.* 100:5419–24.
- Commerford SR, et al. (2007) Dissection of the insulin-sensitizing effect of liver X receptor ligands. *Mol. Endocrinol.* 21:3002–12.
- Kalaany NY, et al. (2005) LXRs regulate the balance between fat storage and oxidation. *Cell Metab.* 1:231–44.
- Nakayama H, et al. (2007) Transgenic mice expressing nuclear sterol regulatory element-binding protein 1c in adipose tissue exhibit liver histology similar to nonalcoholic steatohepatitis. *Metabolism* 56:470–5.
- Yahagi N, et al. (2002) Absence of sterol regulatory element-binding protein-1 (SREBP-1) ameliorates fatty livers but not obesity or insulin resistance in Lep(ob)/Lep(ob) mice. J. Biol. Chem. 277:19353–7.

- Takahashi A, et al. (2005) Transgenic mice overexpressing SREBP-1a under the control of the PEPCK promoter exhibit insulin resistance, but not diabetes. *Biochim. Biophys. Acta* 1740:427–33.
- Shimano H. (2007) SREBP-1c and TFE3, energy transcription factors that regulate hepatic insulin signaling. J. Mol. Med. 85:437–44.
- Yoneda M, et al. (2007) High-sensitivity C-reactive protein is an independent clinical feature of nonalcoholic steatohepatitis (NASH) and also of the severity of fibrosis in NASH. J. Gastroenterol. 42:573–82.
- Ridker PM, Hennekens CH, Buring JE, Rifai N. (2000) C-reactive protein and other markers of inflammation in the prediction of cardiovascular disease in women. N. Engl. J. Med. 342:836–43.
- Lind L. (2003) Circulating markers of inflammation and atherosclerosis. *Atherosclerosis* 169:203–14.
 Kovacs A, Tornvall P, Nilsson R, Tegner J,
- Hamsten A, Bjorkegren J. (2007) Human C-reactive protein slows atherosclerosis development in a mouse model with human-like hypercholesterolemia. *Proc. Natl. Acad. Sci. U. S.* A. 104:13768–73.
- 100. Tilg H, Vannier E, Vachino G, Dinarello CA, Mier JW. (1993) Antiinflammatory properties of hepatic acute phase proteins: preferential induction of interleukin 1 (IL-1) receptor antagonist over IL-1 beta synthesis by human peripheral blood mononuclear cells. J. Exp. Med. 178:1629–36.
- Chen K, et al. (2006) Induction of leptin resistance through direct interaction of C-reactive protein with leptin. *Nat. Med.* 12:425–32.
- 102. Newberry EP, Xie Y, Kennedy SM, Luo J, Davidson NO. (2006) Protection against Western diet-induced obesity and hepatic steatosis in liver fatty acid-binding protein knockout mice. *Hepatology* 44:1191–205.
- Furuhashi M, et al. (2007) Treatment of diabetes and atherosclerosis by inhibiting fatty-acidbinding protein aP2. *Nature* 447:959–65.
- Nomiyama T, et al. (2007) Osteopontin mediates obesity-induced adipose tissue macrophage infiltration and insulin resistance in mice. J. Clin. Invest. 117:2877–88.
- Kim JK, et al. (2004) PKC-theta knockout mice are protected from fat-induced insulin resistance. J. Clin. Invest. 114:823–7.
- 106. Samuel VT, et al. (2007) Inhibition of protein kinase Cepsilon prevents hepatic insulin resistance in nonalcoholic fatty liver disease. J. Clin. Invest. 117:739–45.
- 107. Tsuchiya K, Sakai H, Suzuki N, Iwashima F, Yoshimoto T, Shichiri M, Hirata Y. (2007) Chronic blockade of nitric oxide synthesis reduces adiposity and improves insulin resistance in high fat-induced obese mice. *Endocrinology* 148:4548–56.
- Sanyal AJ, et al. (2001) Nonalcoholic steatohepatitis: association of insulin resistance and mitochondrial abnormalities. *Gastroenterology* 120:1183–92.
- 109. Schattenberg JM, Wang Y, Singh R, Rigoli RM, Czaja MJ. (2005) Hepatocyte CYP2E1 overexpres-

sion and steatohepatitis lead to impaired hepatic insulin signaling. J. Biol. Chem. 280:9887–94.

- 110. Xu Z, Chen L, Leung L, Yen TS, Lee C, Chan JY. (2005) Liver-specific inactivation of the Nrf1 gene in adult mouse leads to nonalcoholic steatohepatitis and hepatic neoplasia. *Proc. Natl. Acad. Sci. U. S. A.* 102:4120–5.
- 111. Barbuio R, Milanski M, Bertolo MB, Saad MJ, Velloso LA. (2007) Infliximab reverses steatosis and improves insulin signal transduction in liver of rats fed a high-fat diet. J. Endocrinol. 194:539–50.
- 112. Ofei F, Hurel S, Newkirk J, Sopwith M, Taylor R. (1996) Effects of an engineered human anti-TNF-alpha antibody (CDP571) on insulin sensitivity and glycemic control in patients with NIDDM. *Diabetes* 45:881–5.
- Bernstein LE, Berry J, Kim S, Canavan B, Grinspoon SK. (2006) Effects of etanercept in patients with the metabolic syndrome. *Arch. Intern. Med.* 166:902–8.
- 114. Paquot N, Castillo MJ, Lefebvre PJ, Scheen AJ. (2000) No increased insulin sensitivity after a single intravenous administration of a recombinant human tumor necrosis factor receptor: Fc fusion protein in obese insulin-resistant patients. J. Clin. Endocrinol. Metab. 85:1316–9.
- 115. Dominguez H, et al. (2005) Metabolic and vascular effects of tumor necrosis factor-alpha blockade with etanercept in obese patients with type 2 diabetes. J. Vasc. Res. 42:517–25.
- 116. Lo J, Bernstein LE, Canavan B, Torriani M, Jackson MB, Ahima RS, Grinspoon SK. (2007) Effects of TNF-{alpha} neutralization on adipocytokines and skeletal muscle adiposity in the metabolic syndrome. *Am. J. Physiol. Endocrinol. Metab.* 293:E102–9.
- 117. Rosenvinge A, Krogh-Madsen R, Baslund B, Pedersen BK. (2007) Insulin resistance in patients with rheumatoid arthritis: effect of anti-TNFalpha therapy. Scand. J. Rheumatol. 36:91–6.
- Ozcan U, et al. (2006) Chemical chaperones reduce ER stress and restore glucose homeostasis in a mouse model of type 2 diabetes. *Science* 313:1137–40.
- Lindor KD, et al. (2004) Ursodeoxycholic acid for treatment of nonalcoholic steatohepatitis: results of a randomized trial. *Hepatology* 39:770–8.
- Hundal RS, Petersen KF, Mayerson AB, Randhawa PS, Inzucchi S, Shoelson SE, Shulman GI. (2002) Mechanism by which high-dose aspirin improves glucose metabolism in type 2 diabetes. J. Clin. Invest. 109:1321–6.
- 121. Forst T, et al. (2007) Effect of simvastatin and/or pioglitazone on insulin resistance, insulin secretion, adiponectin, and proinsulin levels in nondiabetic patients at cardiovascular risk—the PIOSTAT Study. *Metabolism* 56:491–6.
- 122. Hanefeld M, et al. (2007) Anti-inflammatory effects of pioglitazone and/or simvastatin in high cardiovascular risk patients with elevated high sensitivity C-reactive protein: the PIOSTAT Study. J. Am. Coll. Cardiol. 49:290–7.
- 123. Nissen SE, Wolski K. (2007) Effect of rosiglita-

zone on the risk of myocardial infarction and death from cardiovascular causes. *N. Engl. J. Med.* 356:2457–71.

- 124. Lago RM, Singh PP, Nesto RW. (2007) Congestive heart failure and cardiovascular death in patients with prediabetes and type 2 diabetes given thiazolidinediones: a meta-analysis of randomised clinical trials. *Lancet* 370:1129–36.
- 125. Somm E, et al. (2006) Interleukin-1 receptor antagonist is upregulated during diet-induced obesity and regulates insulin sensitivity in rodents. *Diabetologia* 49:387–93.
- Larsen CM, et al. (2007) Interleukin-1-receptor antagonist in type 2 diabetes mellitus. N. Engl. J. Med. 356:1517–26.
- 127. Yang RZ, et al. (2006) Acute-phase serum amyloid A: an inflammatory adipokine and potential link between obesity and its metabolic complications. *PLoS. Med.* 3:e287.
- 128. van Exel E, Gussekloo J, de Craen AJ, Frolich M, Bootsma-Van Der Wiel A, Westendorp RG. (2002) Low production capacity of interleukin-10 associates with the metabolic syndrome and type 2 diabetes: the Leiden 85-Plus Study. *Diabetes* 51:1088–92.
- 129. den Boer MA, et al. (2006) Endogenous interleukin-10 protects against hepatic steatosis but does not improve insulin sensitivity during highfat feeding in mice. *Endocrinology* 147:4553–8.
- Sell H, Dietze-Schroeder D, Eckardt K, Eckel J. (2006) Cytokine secretion by human adipocytes is differentially regulated by adiponectin, AICAR, and troglitazone. *Biochem. Biophys. Res. Commun.* 343:700–6.
- 131. Wu H, et al. (2007) T-cell accumulation and regulated on activation, normal T cell expressed and secreted upregulation in adipose tissue in obesity. *Circulation* 115:1029–38.
- 132. Song Y, et al. (2007) Circulating levels of endothelial adhesion molecules and risk of diabetes in an ethnically diverse cohort of women. *Diabetes* 56:1898–904.
- Matsumoto K, Miyake S, Yano M, Ueki Y, Tominaga Y. (2000) High serum concentrations of soluble E-selectin in patients with impaired glucose tolerance with hyperinsulinemia. *Atherosclerosis* 152:415–20.
- De Pergola G, et al. (2007) sP-selectin plasma levels in obesity: association with insulin resistance and related metabolic and prothrombotic factors. *Nutr. Metab Cardiovasc. Dis.* Mar 30 [Epub ahead of print]
- 135. Kent JW Jr, et al. (2004) Intercellular adhesion molecule-1 concentration is genetically correlated with insulin resistance, obesity, and HDL concentration in Mexican Americans. *Diabetes* 53:2691–5.
- Kim F, et al. (2007) Toll-like receptor-4 mediates vascular inflammation and insulin resistance in diet-induced obesity. *Circ. Res.* 100:1589–96.
- 137. Shi H, Kokoeva MV, Inouye K, Tzameli I, Yin H, Flier JS. (2006) TLR4 links innate immunity and fatty acid-induced insulin resistance. J. Clin. Invest 116:3015–25.