Open Access



Genetically determined high activities of the TNF-alpha, IL23/IL17, and NFkB pathways were associated with increased risk of ankylosing spondylitis

Jacob Sode^{1,2,3,4}, Steffen Bank^{5,6*}, Ulla Vogel⁷, Paal Skytt Andersen^{8,9}, Signe Bek Sørensen^{1,5,10}, Anders Bo Bojesen⁵, Malene Rohr Andersen¹¹, Ivan Brandslund¹², Ram Benny Dessau¹³, Hans Jürgen Hoffmann^{14,15}, Bente Glintborg^{16,17}, Merete Lund Hetland^{17,18}, Henning Locht³, Niels Henrik Heegaard^{2,19*} and Vibeke Andersen^{1,5,10,20}

Abstract

Background: Ankylosing spondylitis (AS) results from the combined effects of susceptibility genes and environmental factors. Polymorphisms in genes regulating inflammation may explain part of the heritability of AS.

Methods: Using a candidate gene approach in this case-control study, 51 mainly functional single nucleotide polymorphisms (SNPs) in genes regulating inflammation were assessed in 709 patients with AS and 795 controls. Data on the patients with AS were obtained from the DANBIO registry where patients from all of Denmark are monitored in routine care during treatment with conventional and biologic disease modifying anti-rheumatic drugs (bDMARDs). The results were analyzed using logistic regression (adjusted for age and sex).

Results: Nine polymorphisms were associated with risk of AS (p < 0.05). The polymorphisms were in genes regulating a: the TNF- α pathway (*TNF* -308 G > A (rs1800629), and – 238 G > A (rs361525); *TNFRSF1A* -609 G > T (rs4149570), and *PTPN22* 1858 G > A (rs2476601)), b: the IL23/IL17 pathway (*IL23R* G > A (rs11209026), and *IL18*–137 G > C (rs187238)), or c: the NFkB pathway (*TLR1* 743 T > C (rs4833095), *TLR4* T > C (rs1554973), and *LY96*–1625 C > G (rs11465996)). After Bonferroni correction the homozygous variant genotype of *TLR1* 743 T > C (rs4833095) (odds ratios (OR): 2.59, 95% confidence interval (CI): 1.48–4.51, p = 0.04), and *TNFRSF1A* -609 G > T (rs4149570) (OR: 1.79, 95% CI: 1.31–2.41, p = 0.01) were associated with increased risk of AS and the combined homozygous and heterozygous variant genotypes of *TNF* -308 G > A (rs1800629) (OR: 0.56, 95% CI: 0.44–0.72, p = 0.0002) were associated with reduced risk of AS.

Conclusion: We replicated associations between AS and the polymorphisms in *TNF* (rs1800629), *TNFRSF1A* (rs4149570), and *IL23R* (rs11209026). Furthermore, we identified novel risk loci in *TNF* (rs361525), *IL18* (rs187238), *TLR1* (rs4833095), *TLR4* (rs1554973), and *LY96* (rs11465996) that need validation in independent cohorts. The results suggest that genetically determined high activity of the TNF-a, IL23/IL17, and NFkB pathways increase risk of AS.

Keywords: Ankylosing spondylitis, Single nucleotide polymorphism, SNP, Case-control study

Deceased

Hospital of Southern Jutland, Aabenraa, Denmark

⁶Medical Department, Viborg Regional Hospital, Viborg, Denmark

Full list of author information is available at the end of the article $% \left({{{\rm{I}}_{{\rm{A}}}}} \right)$



© The Author(s). 2018 **Open Access** This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated.

^{*} Correspondence: stb@mb.au.dk

⁵Focused Research Unit for Molecular Diagnostic and Clinical Research,

Background

Ankylosing spondylitis (AS) is a type of spondyloarthritis in which hallmark clinical features are inflammation at entheses and subchondral bone of the pelvic and spinal joints with subsequent abnormal new bone formation at these sites. Ultimately, this leads to ossification of entheses and joints resulting in loss of joint mobility. The incidence varies between 0.1 and 1.8% with the highest incidence in Scandinavia. Onset is typically in young adults with a male predominance. Medications used include non-steroid anti-inflammatory drugs (NSAIDs), and biological diseasemodifying anti-rheumatic drugs (bDMARDs), i.e. tumor necrosis factor- α inhibitors (anti-TNF) and more recently also an interleukin(IL)-17A inhibitor (secukinumab) [1].

The cause of AS is unknown but is believed to involve a combination of genetic and environmental factors [2]. The heritability is polygenic and estimated to exceed 90%, with the HLA-B27 allele as the major contributor accounting for approximately 25% of the heritability of AS [2]. The IL-17/ IL-23 pathway and the TNF- α pathway are central in the pathogenesis of AS and alterations in these pathways have been shown in mouse models to affect development and severity of enthesitis [3, 4].

TNF- α can be activated by Pathogen-Associated Molecular Patterns (PAMPs) such as bacterial or viral DNA, flagellin, or lipopolysaccharide (LPS), through the NFkB pathway. PAMPs can be recognized by Toll-like receptors (TLRs) thereby initiating a kinase cascade which phosphorylates and degrades the NFkB inhibitor IkB α [5]. This releases NFkB which is transported from the cytosol to the nucleus where it initiates expression of pro- and anti-inflammatory cytokines including TNF- α and IL-17 (http://www.bu.edu/nf-kb/gene-resources/targetgenes/). The TNF- α and NFkB pathway are intertwined and TNF- α can feedback stimulate NFkB by binding to TNF receptors (TNFR1 or TNFR2), resulting in a kinase cascade similar to, but distinct from, the pathway induced by TLRs [5].

The IL23/IL17 pathway can also stimulate TNF- α activity. The pro-inflammatory cytokine IL-17 enhances the production of other pro-inflammatory cytokines including TNF- α , and the secretion IL-17 itself can be enhanced by IL-23 [6].

PAMPs can also be recognized by intracellular Nodlike receptors (NLRs). In turn, NLRs can activate proinflammatory cytokines including IL-18 [7]. IL-18 is invloved in the IL23/IL17 pathway and can enhance the production of IL-17 [8].

The aim of this study was to assess whether functional single nucleotide polymorphisms.

(SNPs) in genes involved in the TNF- α , IL23/IL17, NFkB, and other pro- and anti-inflammatory pathways were associated with risk of AS.

Methods

Patients and samples

The DANBIO registry includes prospectively collected clinical data on patients with inflammatory joint diseases including smoking status, disease characteristics e.g. HLA-B27 status, disease activity, treatment, and treatment outcomes. Patients from all of Denmark are monitored in routine care during treatment with conventional and biologic disease modifying anti-rheumatic drugs (bDMARDs) [9].

Screening for tuberculosis before initiation of treatment with biological drugs is routinely performed in Denmark. Left over blood clots (after whole blood analysis for Mycobacterium tuberculosis) were collected from all patients screened for tuberculosis at Statens Serum Institut (Copenhagen, Denmark) from 01.09.2009 to 31.01.2013; the Department of Respiratory Diseases B and the Department of Clinical Microbiology, Aarhus University Hospital (Aarhus, Denmark) from 01.01.2011 to 31.01.2014; the Department of Clinical Biochemistry, Herlev and Gentofte Hospital (Hellerup, Denmark) from 01.03.2012 to 31.01.2014; the Department of Biochemistry, Hospital of Lillebaelt (Vejle, Denmark); and the Department of Biochemistry, Hospital of Slagelse (Slagelse, Denmark) from 01.01.2014 to 31.01.2014. Furthermore, from 01.01.2013 to 31.12.2013 blood samples were collected from all patients with AS treated with or without anti-TNF drugs at the Department of Rheumatology, Frederiksberg Hospital (Frederiksberg, Denmark).

By linking the unique personal identification number of Danish citizens (CPR-number) from each blood sample with the clinical data from DANBIO, 709 patients with AS (ICD-10: M45.9) were identified. The control group consisted of 795 healthy blood donors recruited from Viborg, Denmark.

Genotyping

Fifty-one SNPs in genes involved in the TNF- α , IL23/ IL17, NF κ B, and other pro- and anti-inflammatory pathways were assessed. A list of all SNPs studied and genotype distribution is presented in Table 1 and SNPs associated with AS are summarized in Table 2.

DNA extraction (Maxwell 16 LEV Blood DNA Kit; Promega, Madison, WI, USA) was performed as described by *Bank* et al. [10]. For the healthy controls, DNA was extracted from EDTA-stabilized peripheral blood by either PureGene (Qiagen, Hilden, Germany) or Wizard Genomic (Promega, Madison, Wisconsin, USA) DNA purification kit according to the manufacturers' instructions [11–17]. Competitive Allele-Specific Polymerase chain reaction (KASP[™]), an end-point PCR technology, was used by LGC Genomics for genotyping (LGC Genomics, Hoddesdon, United Kingdom) (http:// www.lgcgenomics.com/).

Gene	Healthy controls	AS	Unadjusted		Adjusted, age & s	iex	Adjusted, age, sex	& smoking
s-number			OR (95 CI)	р	OR (9 5CI)	р	OR (95 CI)	р
<i>'LR1</i> s4833095								
TT	485	415						
TC	261	238	1.07 (0.86–1.33)	0.57	1.03 (0.82–1.29)	0.83	1.05 (0.78–1.42)	0.73
CC	20	43	2.51 (1.45–4.34)	0.00095	2.59 (1.48–4.51)	0.00081	2.86 (1.44–5.68)	0.0026
TC or CC	281	281	1.17 (0.95–1.44)	0.15	1.14 (0.91–1.41)	0.25	1.18 (0.89–1.58)	0.26
<i>"LR2</i> s3804099								
TT	241	197						
TC	393	354	1.10 (0.87–1.40)	0.42	1.07 (0.84–1.37)	0.58	1.02 (0.73–1.42)	0.90
CC	144	142	1.21 (0.89–1.63)	0.22	1.24 (0.91–1.68)	0.17	1.30 (0.87–1.96)	0.20
TC or CC	537	496	1.13 (0.90–1.41)	0.29	1.11 (0.89–1.40)	0.36	1.10 (0.80–1.50)	0.57
<i>"LR2</i> s11938228								
CC	327	314						
CA	368	313	0.89 (0.71–1.10)	0.27	0.86 (0.69–1.07)	0.17	0.80 (0.60–1.08)	0.15
AA	76	69	0.95 (0.66–1.36)	0.76	0.92 (0.63–1.33)	0.66	1.03 (0.62–1.69)	0.92
CA or AA	444	382	0.90 (0.73–1.10)	0.30	0.87 (0.70–1.07)	0.19	0.84 (0.63–1.11)	0.22
"LR2 s4696480								
AA	199	179						
AT	417	348	0.93 (0.72–1.19)	0.55	0.89 (0.69–1.15)	0.38	0.84 (0.60–1.18)	0.31
Π	155	169	1.21 (0.90–1.63)	0.20	1.16 (0.86–1.58)	0.33	1.18 (0.78–1.78)	0.44
AT or TT	572	517	1.00 (0.79–1.27)	0.97	0.97 (0.76–1.23)	0.78	0.92 (0.67–1.27)	0.62
LR4 5030728								
GG	359	322						
GA	323	298	1.03 (0.83–1.28)	0.80	1.01 (0.81–1.27)	0.91	0.93 (0.69–1.25)	0.62
AA	78	70	1.00 (0.70–1.43)	1.00	0.98 (0.68–1.42)	0.93	0.87 (0.53–1.42)	0.57
GA or AA	401	368	1.02 (0.83–1.26)	0.83	1.01 (0.82–1.25)	0.94	0.91 (0.69–1.21)	0.53
<i>"LR4</i> s1554973								
TT	440	395						
TC	272	261	1.07 (0.86–1.33)	0.55	1.06 (0.85–1.32)	0.62	0.98 (0.73–1.32)	0.90
CC	62	33	0.59 (0.38–0.92)	0.02	0.55 (0.34–0.86)	0.01	0.68 (0.38–1.23)	0.20
TC or CC	334	294	0.98 (0.80–1.21)	0.85	0.96 (0.78–1.19)	0.72	0.93 (0.70–1.24)	0.63
"LR4 s12377632								
TT	306	271						
TC	358	319	1.01 (0.81–1.26)	0.96	1.05 (0.84–1.32)	0.66	1.07 (0.78–1.46)	0.67
CC	102	96	1.06 (0.77–1.47)	0.71	1.11 (0.80–1.55)	0.52	1.41 (0.92–2.17)	0.12
TC or CC	460	415	1.02 (0.83–1.26)	0.86	1.06 (0.86–1.32)	0.58	1.14 (0.85–1.53)	0.37
<i>"LR5</i> s5744168								
CC	672	605						

 Table 1 Odds ratios (OR) and 95% confidence interval (95Cl) for genotypes studied among healthy controls and patients with ankylosing spondylitis (AS)

Gene	Healthy controls	AS	Unadjusted		Adjusted, age & s	ex	Adjusted, age, sex	& smoking
rs-number			OR (95 CI)	р	OR (9 5CI)	р	OR (95 CI)	р
CT	94	89	1.05 (0.77–1.43)	0.75	1.05 (0.77–1.45)	0.74	0.89 (0.58–1.37)	0.60
TT	5	2	0.44 (0.09–2.30)	0.33	0.45 (0.08–2.43)	0.35	0.04 (0.00-3.54)	0.16
CT or TT	99	91	1.02 (0.75–1.39)	0.89	1.02 (0.75–1.40)	0.88	0.84 (0.55–1.29)	0.43
<i>TLR5</i> rs5744174								
TT	215	216						
TC	399	337	0.84 (0.66–1.07)	0.15	0.85 (0.67–1.09)	0.20	0.82 (0.60–1.14)	0.24
CC	144	138	0.95 (0.71–1.29)	0.76	1.02 (0.75–1.39)	0.91	0.87 (0.57–1.32)	0.51
TC or CC	543	475	0.87 (0.69–1.09)	0.23	0.90 (0.71–1.13)	0.36	0.84 (0.62–1.14)	0.26
<i>TLR9</i> rs187084								
TT	262	237						
TC	366	335	1.01 (0.80–1.27)	0.92	1.03 (0.82–1.31)	0.78	1.09 (0.79–1.50)	0.60
CC	142	120	0.93 (0.69–1.26)	0.66	0.91 (0.67–1.24)	0.56	1.07 (0.71–1.61)	0.76
TC or CC	508	455	0.99 (0.80–1.23)	0.93	1.00 (0.80–1.25)	0.98	1.08 (0.80–1.46)	0.60
<i>TLR9</i> rs352139								
GG	255	211						
GA	347	324	1.13 (0.89–1.43)	0.32	1.08 (0.85–1.38)	0.52	1.01 (0.73–1.40)	0.93
AA	167	139	1.01 (0.75–1.34)	0.97	0.96 (0.71–1.30)	0.79	0.80 (0.53–1.20)	0.27
GA or AA	514	463	1.09 (0.87–1.36)	0.45	1.04 (0.83–1.31)	0.72	0.94 (0.69–1.27)	0.68
<i>LY96</i> rs11465996								
CC	344	341						
CG	337	298	0.89 (0.72–1.11)	0.30	0.91 (0.73–1.14)	0.42	0.89 (0.66–1.20)	0.45
GG	81	53	0.66 (0.45-0.96)	0.03	0.68 (0.46–1.00)	0.0498	0.65 (0.39–1.10)	0.11
CG or GG	418	351	0.85 (0.69–1.04)	0.11	0.87 (0.70–1.07)	0.18	0.84 (0.63–1.12)	0.24
<i>CD14</i> Rs2569190								
GG	236	194						
GA	360	339	1.15 (0.90–1.46)	0.27	1.18 (0.92–1.51)	0.19	1.27 (0.91–1.78)	0.16
AA	170	157	1.12 (0.84–1.50)	0.43	1.20 (0.89–1.61)	0.24	1.46 (0.98–2.19)	0.06
GA or AA	530	496	1.14 (0.91–1.43)	0.26	1.18 (0.94–1.50)	0.15	1.32 (0.96–1.82)	0.08
<i>TIRAP</i> rs8177374								
CC	556	521						
CT	185	159	0.92 (0.72–1.17)	0.49	0.99 (0.77–1.27)	0.94	1.38 (0.99–1.91)	0.06
TT	21	15	0.76 (0.39–1.49)	0.43	0.76 (0.38–1.53)	0.45	1.31 (0.55–3.12)	0.55
CT or TT	206	174	0.90 (0.71–1.14)	0.39	0.97 (0.76–1.23)	0.81	1.38 (1.00–1.89)	0.047
<i>SUMO4</i> rs237025								
TT	215	195						
TC	362	358	1.09 (0.86–1.39)	0.48	1.08 (0.84–1.38)	0.55	1.04 (0.75–1.44)	0.80
CC	195	136	0.77 (0.57–1.03)	0.08	0.75 (0.55–1.01)	0.06	0.55 (0.36–0.84)	0.01
TC or CC	557	494	0.98 (0.78–1.23)	0.85	0.96 (0.76–1.22)	0.75	0.87 (0.64–1.19)	0.38

 Table 1 Odds ratios (OR) and 95% confidence interval (95Cl) for genotypes studied among healthy controls and patients with ankylosing spondylitis (AS) (Continued)

Gene	Healthy controls	AS	Unadjusted		Adjusted, age & s	Sex	Adjusted, age, sex	& smoking
rs-number			OR (95 CI)	р	OR (9 5CI)	p	OR (95 CI)	р
NFKBIA rs696								
GG	298	259						
GA	366	336	1.06 (0.85–1.32)	0.63	1.06 (0.84–1.33)	0.64	1.02 (0.75–1.39)	0.88
AA	101	90	1.03 (0.74–1.43)	0.88	0.97 (0.69–1.36)	0.86	1.07 (0.67–1.69)	0.78
GA or AA	467	426	1.05 (0.85–1.30)	0.65	1.04 (0.84–1.29)	0.73	1.03 (0.77–1.38)	0.84
<i>NFKB1</i> rs28362491								
Ins/Ins	269	258						
lns/–	376	316	0.88 (0.70–1.10)	0.25	0.89 (0.70–1.12)	0.31	0.74 (0.54–1.01)	0.06
/	122	100	0.85 (0.62–1.17)	0.33	0.82 (0.59–1.13)	0.22	0.78 (0.51–1.19)	0.25
Ins/– or –/–	498	416	0.87 (0.70–1.08)	0.21	0.87 (0.70–1.08)	0.21	0.75 (0.56–1.01)	0.06
<i>TNF</i> rs1800629								
GG	527	549						
GA	223	129	0.56 (0.43–0.71)	0.0000032	0.58 (0.45–0.75)	0.000029	0.63 (0.45–0.89)	0.01
AA	25	9	0.35 (0.16–0.75)	0.01	0.39 (0.18–0.85)	0.02	0.19 (0.04–0.79)	0.02
GA or AA	248	138	0.53 (0.42–0.68)	0.0000030	0.56 (0.44–0.72)	0.0000047	0.59 (0.42–0.82)	0.0018
TNF rs361525								
GG	708	669						
GA	60	30	0.53 (0.34–0.83)	0.01	0.52 (0.32–0.82)	0.0049	0.61 (0.33–1.12)	0.11
AA	3	0	1.00 (1.00-1.00)	1.00	1.00 (1.00-1.00)	1.00	1.00 (1.00-1.00)	1.00
GA or AA	63	30	0.50 (0.32–0.79)	0.0027	0.49 (0.31–0.78)	0.0024	0.58 (0.32-1.05)	0.07
<i>TNFRSF1A</i> rs4149570								
GG	307	217						
GT	355	339	1.35 (1.07–1.70)	0.01	1.33 (1.05–1.68)	0.02	1.46 (1.06–2.00)	0.02
TT	109	132	1.71 (1.26–2.33)	0.00060	1.79 (1.31–2.46)	0.00027	2.26 (1.48–3.47)	0.00017
GT or TT	464	471	1.44 (1.16–1.78)	0.0010	1.44 (1.15–1.80)	0.0013	1.64 (1.21–2.22)	0.0014
<i>TNFAIP3</i> rs6927172								
CC	473	415						
CG	264	245	1.06 (0.85–1.32)	0.61	1.06 (0.85–1.33)	0.61	1.03 (0.76–1.39)	0.85
GG	40	25	0.71 (0.42–1.19)	0.20	0.70 (0.41–1.19)	0.18	0.51 (0.23–1.10)	0.09
CG or GG	304	270	1.01 (0.82–1.25)	0.91	1.01 (0.82–1.26)	0.91	0.95 (0.71–1.27)	0.73
<i>TGFB1</i> rs1800469								
CC	383	344						
CT	297	299	1.12 (0.90–1.39)	0.30	1.08 (0.87–1.35)	0.48	1.28 (0.95–1.71)	0.11
TT	86	53	0.69 (0.47–1.00)	0.047	0.69 (0.47–1.02)	0.06	0.69 (0.40–1.17)	0.17
CT or TT	383	352	1.02 (0.83–1.26)	0.83	1.00 (0.81–1.23)	0.97	1.14 (0.86–1.52)	0.35
<i>PTPN22</i> rs2476601								
GG	588	557						

 Table 1 Odds ratios (OR) and 95% confidence interval (95Cl) for genotypes studied among healthy controls and patients with ankylosing spondylitis (AS) (Continued)

Gene	Healthy controls	AS	Unadjusted		Adjusted, age & s	ex	Adjusted, age, sex	& smoking
rs-number			OR (95 CI)	р	OR (9 5CI)	р	OR (95 CI)	р
GA	166	122	0.78 (0.60–1.01)	0.06	0.77 (0.59–1.00)	0.05	0.75 (0.52–1.09)	0.13
AA	11	6	0.58 (0.21–1.57)	0.28	0.57 (0.20–1.58)	0.28	0.83 (0.21–3.28)	0.80
GA or AA	177	128	0.76 (0.59–0.99)	0.04	0.76 (0.58–0.98)	0.04	0.76 (0.53–1.09)	0.13
<i>PPARG</i> rs1801282								
CC	548	511						
CG	207	167	0.87 (0.68–1.10)	0.23	0.85 (0.66–1.08)	0.18	0.87 (0.63–1.21)	0.42
GG	14	15	1.15 (0.55–2.40)	0.71	1.33 (0.62–2.83)	0.46	1.54 (0.60–3.98)	0.37
CG or GG	221	182	0.88 (0.70-1.11)	0.29	0.88 (0.69–1.11)	0.27	0.91 (0.67–1.26)	0.58
IL1B rs4848306								
GG	246	215						
GA	373	352	1.08 (0.85–1.36)	0.52	1.09 (0.86–1.39)	0.48	1.16 (0.84–1.60)	0.38
AA	151	125	0.95 (0.70-1.28)	0.72	0.96 (0.71–1.31)	0.81	0.88 (0.57–1.34)	0.55
GA or AA	524	477	1.04 (0.83–1.30)	0.72	1.06 (0.84–1.33)	0.64	1.08 (0.79–1.46)	0.64
IL1B rs1143623								
GG	401	365						
GC	316	278	0.97 (0.78–1.20)	0.76	0.98 (0.79–1.22)	0.87	1.07 (0.80–1.44)	0.66
CC	55	52	1.04 (0.69–1.56)	0.85	1.12 (0.74–1.69)	0.59	0.87 (0.48–1.57)	0.64
GC or CC	371	330	0.98 (0.80-1.20)	0.83	1.00 (0.81–1.24)	0.98	1.04 (0.78–1.38)	0.79
<i>IL1B</i> rs1143627								
TT	340	305						
TC	339	305	1.00 (0.81-1.25)	0.98	1.00 (0.79–1.25)	0.97	1.05 (0.78–1.42)	0.75
CC	97	86	0.99 (0.71–1.37)	0.94	1.01 (0.72–1.41)	0.95	0.85 (0.53–1.36)	0.50
TC or CC	436	391	1.00 (0.81-1.23)	1.00	1.00 (0.81–1.24)	1.00	1.00 (0.76–1.34)	0.97
IL1RN rs4251961								
TT	298	247						
TC	360	324	1.09 (0.87–1.36)	0.47	1.04 (0.83–1.32)	0.71	1.22 (0.89–1.67)	0.21
CC	112	105	1.13 (0.83–1.55)	0.44	1.05 (0.76–1.46)	0.76	1.41 (0.92–2.17)	0.12
TC or CC	472	429	1.10 (0.89–1.36)	0.40	1.05 (0.84–1.30)	0.68	1.26 (0.94–1.71)	0.12
<i>L4R</i> rs1805010								
AA	209	201						
AG	410	317	0.80 (0.63–1.02)	0.08	0.79 (0.62–1.02)	0.07	0.73 (0.52–1.02)	0.07
GG	157	133	0.88 (0.65–1.19)	0.41	0.91 (0.67–1.24)	0.55	0.87 (0.58–1.33)	0.53
AG or GG	567	450	0.83 (0.66–1.04)	0.10	0.83 (0.65–1.05)	0.12	0.77 (0.56–1.06)	0.11
<i>L6</i> rs10499563								
TT	476	439						
TC	259	225	0.94 (0.76–1.17)	0.60	0.94 (0.75–1.18)	0.60	0.77 (0.57–1.05)	0.10
CC	35	26	0.81 (0.48–1.36)	0.42	0.72 (0.42–1.25)	0.24	0.80 (0.39–1.63)	0.53
TC or CC	294	251	0.93 (0.75–1.14)	0.48	0.92 (0.74–1.14)	0.43	0.77 (0.57–1.04)	0.09

 Table 1 Odds ratios (OR) and 95% confidence interval (95Cl) for genotypes studied among healthy controls and patients with ankylosing spondylitis (AS) (Continued)

Gene	Healthy controls	AS	Unadjusted		Adjusted, age & s	Sex	Adjusted, age, sex	& smoking
rs-number			OR (95 CI)	р	OR (9 5CI)	р	OR (95 CI)	р
L6R s4537545								
CC	289	247						
CT	369	324	1.03 (0.82–1.29)	0.82	1.05 (0.83–1.32)	0.71	1.07 (0.79–1.47)	0.65
TT	117	113	1.13 (0.83–1.54)	0.44	1.18 (0.86–1.63)	0.30	1.17 (0.76–1.79)	0.48
CT or TT	486	437	1.05 (0.85–1.30)	0.64	1.08 (0.86–1.34)	0.51	1.09 (0.81–1.47)	0.55
L10 s1800872								
CC	482	408						
CA	258	225	1.03 (0.83–1.29)	0.79	1.01 (0.80–1.27)	0.94	0.93 (0.68–1.26)	0.63
AA	35	42	1.42 (0.89–2.26)	0.14	1.35 (0.83–2.18)	0.22	1.47 (0.79–2.73)	0.22
CA or AA	293	267	1.08 (0.87–1.33)	0.50	1.05 (0.84–1.30)	0.67	0.99 (0.74–1.33)	0.95
<i>L10</i> s3024505								
CC	518	467						
CT	221	200	1.00 (0.80–1.26)	0.97	1.01 (0.80–1.28)	0.95	1.19 (0.87–1.61)	0.28
TT	22	24	1.21 (0.67–2.19)	0.53	1.32 (0.72–2.42)	0.37	1.80 (0.79–4.12)	0.16
CT or TT	243	224	1.02 (0.82–1.27)	0.84	1.04 (0.83–1.30)	0.76	1.23 (0.92–1.66)	0.17
<i>L12B</i> s3212217								
GG	499	460						
GC	235	200	0.92 (0.74–1.16)	0.49	0.95 (0.75–1.19)	0.64	0.94 (0.69–1.29)	0.72
CC	25	21	0.91 (0.50–1.65)	0.76	0.94 (0.51–1.72)	0.84	0.57 (0.23–1.41)	0.22
GC or CC	260	221	0.92 (0.74–1.15)	0.47	0.95 (0.76–1.19)	0.63	0.91 (0.67–1.23)	0.53
L12B s6887695								
GG	385	324						
GC	293	301	1.22 (0.98–1.52)	0.07	1.24 (0.99–1.55)	0.06	1.31 (0.97–1.77)	0.07
CC	72	70	1.16 (0.81–1.66)	0.43	1.16 (0.80–1.69)	0.43	0.98 (0.59–1.61)	0.94
GC or CC	365	371	1.21 (0.98–1.49)	0.07	1.22 (0.99–1.51)	0.06	1.24 (0.93–1.64)	0.14
<i>L12RB1</i> s401502								
CC	360	304						
CG	303	311	1.22 (0.98–1.51)	0.08	1.21 (0.96–1.51)	0.10	1.19 (0.88–1.61)	0.26
GG	87	70	0.95 (0.67–1.35)	0.79	0.97 (0.68–1.39)	0.87	1.18 (0.74–1.88)	0.48
CG or GG	390	381	1.16 (0.94–1.42)	0.17	1.15 (0.93–1.43)	0.19	1.19 (0.89–1.58)	0.24
L17A s2275913								
GG	340	307						
GA	336	301	0.99 (0.80–1.24)	0.94	0.98 (0.79–1.23)	0.89	0.90 (0.67–1.22)	0.51
AA	95	84	0.98 (0.70–1.36)	0.90	1.00 (0.71–1.40)	0.98	1.00 (0.63–1.57)	0.99
GA or AA	431	385	0.99 (0.80–1.22)	0.92	0.99 (0.80–1.22)	0.89	0.92 (0.69–1.22)	0.57
L18 rs187238								
GG	387	380						

 Table 1 Odds ratios (OR) and 95% confidence interval (95Cl) for genotypes studied among healthy controls and patients with ankylosing spondylitis (AS) (Continued)

Table 1 Odds ratios (OR) and 95% confidence interval (95CI) for genotype	es studied among healthy controls and patients with
ankylosing spondylitis (AS) <i>(Continued)</i>	

Gene	Healthy controls	AS	Unadjusted		Adjusted, age & s	iex	Adjusted, age, sex	& smoking
rs-number			OR (95 CI)	р	OR (9 5CI)	р	OR (95 CI)	р
GC	312	259	0.85 (0.68–1.05)	0.13	0.83 (0.66–1.03)	0.09	0.74 (0.55–1.00)	0.049
CC	64	41	0.65 (0.43–0.99)	0.04	0.69 (0.45–1.06)	0.09	0.58 (0.32–1.04)	0.07
GC or CC	376	300	0.81 (0.66–1.00)	0.0499	0.80 (0.65–0.99)	0.04	0.71 (0.53–0.95)	0.02
<i>IL18</i> rs1946518								
GG	282	259						
GT	363	329	0.99 (0.79–1.24)	0.91	0.96 (0.76–1.21)	0.71	0.89 (0.65–1.21)	0.45
TT	113	97	0.93 (0.68–1.29)	0.68	0.95 (0.68–1.31)	0.74	0.80 (0.51–1.24)	0.32
GT or TT	476	426	0.97 (0.79–1.21)	0.81	0.96 (0.77–1.19)	0.68	0.86 (0.64–1.16)	0.32
<i>IL23R</i> rs11209026								
GG	680	646						
GA	89	50	0.59 (0.41–0.85)	0.0045	0.63 (0.43–0.91)	0.02	0.64 (0.38–1.05)	0.08
AA	5	1	1.00 (1.00-1.00)	1.00	1.00 (1.00-1.00)	1.00	1.00 (1.00-1.00)	1.00
GA or AA	94	51	0.57 (0.40–0.82)	0.0021	0.60 (0.42–0.87)	0.01	0.63 (0.38–1.03)	0.06
<i>IFNG</i> rs2430561								
TT	199	181						
TA	398	369	1.02 (0.80–1.30)	0.88	1.01 (0.79–1.30)	0.92	1.08 (0.77–1.52)	0.65
AA	161	139	0.95 (0.70–1.29)	0.74	0.97 (0.71–1.32)	0.85	1.09 (0.72–1.64)	0.68
TA or AA	559	508	1.00 (0.79–1.26)	0.99	1.00 (0.79–1.27)	0.99	1.08 (0.79–1.50)	0.62
<i>IFNGR1</i> rs2234711								
TT	290	232						
TC	361	348	1.20 (0.96–1.51)	0.11	1.20 (0.95–1.51)	0.12	1.15 (0.84–1.57)	0.40
CC	119	108	1.13 (0.83–1.55)	0.43	1.09 (0.79–1.50)	0.60	1.11 (0.72–1.70)	0.65
TC or CC	480	456	1.19 (0.96–1.47)	0.12	1.17 (0.94–1.46)	0.16	1.14 (0.84–1.53)	0.40
IFNGR2 rs8126756								
TT	553	522						
TC	168	130	0.82 (0.63–1.06)	0.13	0.83 (0.64–1.09)	0.18	0.86 (0.60–1.24)	0.42
CC	18	12	0.71 (0.34–1.48)	0.36	0.69 (0.32–1.49)	0.35	0.53 (0.18–1.54)	0.24
TC or CC	186	142	0.81 (0.63–1.04)	0.09	0.82 (0.64–1.06)	0.13	0.83 (0.59–1.17)	0.28
<i>IFNGR2</i> rs17882748								
CC	199	173						
CT	391	341	1.00 (0.78–1.29)	0.98	1.00 (0.77–1.30)	0.99	1.01 (0.71–1.42)	0.97
TT	153	174	1.31 (0.97–1.76)	0.08	1.31 (0.97–1.78)	0.08	1.16 (0.77–1.73)	0.48
CT or TT	544	515	1.09 (0.86–1.38)	0.48	1.09 (0.86–1.39)	0.48	1.05 (0.76–1.45)	0.76
<i>TBX21</i> rs17250932								
TT	526	497						
TC	210	179	0.90 (0.71–1.14)	0.39	0.94 (0.74–1.19)	0.61	0.84 (0.60–1.17)	0.30
CC	32	19	0.63 (0.35–1.12)	0.12	0.66 (0.36–1.19)	0.17	0.37 (0.14–0.98)	0.046
TC or CC	242	198	0.87 (0.69–1.08)	0.21	0.90 (0.72-1.14)	0.39	0.78 (0.56–1.07)	0.12

Gene	Healthy controls	AS	Unadjusted		Adjusted, age & s	ex.	Adjusted, age, sex	& smoking
rs-number			OR (95 CI)	р	OR (9 5CI)	р	OR (95 CI)	р
NLRP1 rs2670660								
AA	222	202						
AG	390	328	0.92 (0.73–1.18)	0.52	0.96 (0.75–1.23)	0.73	1.12 (0.80–1.56)	0.52
GG	154	154	1.10 (0.82–1.47)	0.53	1.11 (0.82–1.49)	0.51	1.12 (0.75–1.67)	0.59
AG or GG	544	482	0.97 (0.78–1.22)	0.82	1.00 (0.79–1.26)	0.98	1.11 (0.81–1.52)	0.50
NLRP1 rs878329								
GG	217	206						
GC	394	333	0.89 (0.70–1.13)	0.34	0.89 (0.69–1.14)	0.35	0.99 (0.71–1.38)	0.93
CC	155	155	1.05 (0.79–1.41)	0.73	1.05 (0.78–1.41)	0.75	1.03 (0.69–1.54)	0.90
GC or CC	549	488	0.94 (0.75–1.17)	0.57	0.93 (0.74–1.18)	0.56	1.00 (0.73–1.36)	0.98
<i>NLRP3</i> rs10754558								
CC	294	248						
CG	355	324	1.08 (0.86–1.36)	0.50	1.06 (0.84–1.34)	0.61	1.10 (0.81–1.51)	0.54
GG	111	116	1.24 (0.91–1.69)	0.18	1.25 (0.91–1.71)	0.17	1.11 (0.71–1.72)	0.65
CG or GG	466	440	1.12 (0.90–1.39)	0.30	1.11 (0.89–1.38)	0.36	1.11 (0.82–1.49)	0.51
NLRP3 rs4612666								
CC	435	360						
CT	280	277	1.20 (0.96–1.49)	0.11	1.23 (0.99–1.54)	0.07	1.28 (0.95–1.72)	0.10
TT	53	48	1.09 (0.72–1.66)	0.67	1.19 (0.78–1.82)	0.41	1.07 (0.59–1.94)	0.82
CT or TT	333	325	1.18 (0.96–1.45)	0.12	1.23 (0.99–1.52)	0.06	1.24 (0.94–1.65)	0.13
CARD8 rs2043211								
AA	321	298						
AT	342	316	1.00 (0.80–1.24)	0.97	0.98 (0.79–1.23)	0.89	0.90 (0.67–1.22)	0.50
TT	94	78	0.89 (0.64–1.25)	0.52	0.89 (0.63–1.26)	0.50	0.91 (0.57–1.44)	0.68
AT or TT	436	394	0.97 (0.79–1.20)	0.80	0.96 (0.78–1.19)	0.72	0.90 (0.67–1.19)	0.45
<i>JAK2</i> rs12343867								
TT	398	358						
TC	299	263	0.98 (0.79–1.22)	0.84	0.96 (0.76–1.20)	0.69	0.82 (0.61–1.12)	0.21
CC	61	65	1.18 (0.81–1.73)	0.38	1.11 (0.75–1.63)	0.61	1.03 (0.62–1.71)	0.91
TC or CC	360	328	1.01 (0.82–1.25)	0.90	0.98 (0.79–1.21)	0.86	0.86 (0.64–1.14)	0.29

Table 1 Odds ratios (OR) and 95% confidence interval (95CI) for genotypes studied among healthy controls and patients with ankylosing spondylitis (AS) (Continued)

Power calculation

The Genetic Power Calculator was utilized for power analysis of discrete traits (http://zzz.bwh.harvard.edu/gpc/cc2.html). The lowest minor allele frequency (MAF) of the studied SNPs was 0.10. The 'high-risk allele frequency' was set to 0.10, the 'prevalence' was set to 0.0018 [18], D-prime was set to 1, type I error rate was set to 0.05 and number of cases and control:case ratio was 795:709. This cohort study had more than 80%

chance of detecting a dominant effect with an odds ratio (OR) of 1.4 for AS.

Statistical analysis

Logistic regression was used to compare genotype distributions among patients with AS versus healthy controls. Crude odds ratio, odds ratio adjusted for age and sex, and odds ratio adjusted for age, sex, and smoking status were assessed (Table 1). A chi-square test was

.is	
Ę	
ģ	
ğ	
SO	
p	1
Sil	
3	
¥	
a C	
jţ	
>	
e e	
iat	
ğ	
asso	
.0 .0	•
SNP	
S	,
SC	
SL	
Ę	
ő	
Ē	
6	
0	
- Ŭ	
ğ	
9	
Ď	
n alpr	
g	1
SII:	
in of the sin	
ft	
0	
<u> </u>	
ati	
ret	
d L	
Jte	
<u>i</u> G	
bC	1
0	
le 2 Biologia	
N	

Gene	Rs-number	Pathway	Model	OR (95% CI)	P-value / Bonferroni ^a Effect of minor-allele	Effect of minor-allele	Biological interpretation
TLR1	rs4833095	Pathogen recognition CC vs TT		2.59 (1.48-4.51) 0.00081 / 0.04	0.0081 / 0.04	743C increase TLR1 level in PBMC [56]	Increased TLR1 level was associated with increased risk of AS. This could indicate that a genetically determined high activity of the NFkB pathway, and thus high TNF-a and IL-17 activity, was associated with increased risk of AS.
TLR4	rs1554973	Pathogen recognition CC vs Π		0.55 (0.34-0.86) 0.010 / 0.51	0.010 / 0.51	Unknown [67]	1
9671	rs11465996	Pathogen recognition	GG vs CC	0.68 (0.46–1.00)	0.049 / 1.00	-1 625G increase MD-2 and TNF-a levels in human U937 cells and whole blood leukocytes [57]	Increased MD-2 and TNF-a level was associated with a reduced risk of AS. In contrast to the other results this indicate that genetically determined high TNF-driven inflammatory response was associated with reduced risk of AS.
TNF	rs1800629	Cytokines	GA or AA vs GG	0.56 (0.44–0.72)	0.56 (0.44-0.72) 0.0000047 / 0.00024	-308A increase expression in jurkat cells [65], reduce mRNA level in PBMC and serum [48] or no association was found [49]	Reduced TNF-a mRNA level was associated with reduced risk of AS. This could indicate that genetically determined high TNF-driven inflammatory response was associated with increased risk of AS.
TNF	rs361525	Cytokines	GA or AA vs GG	0.49 (0.31–0.78) 0.0024 / 0.12	0.0024 / 0.12	-238A reduce expression in PBMC [49]	Reduced TNF-a expression was associated with r educed risk of AS. This indicates that genetically determined high TNF-driven inflammatory response was associated with increased risk of AS.
TNFRSF1A	TNFRSF1A rs4149570	Cytokines	GT or TT vs GG	1.44 (1.15–1.80) 0.0013 / 0.066 ^b	0.0013 / 0.066 ^b	-609 T increase expression in PBMC [50]	Increased TNF-a receptor 1 expression was associated with increased risk of AS. This indicates that genetically determined high TNF-driven inflammatory response was associated with increased risk of AS.
PTPN22	rs2476601	Immune response	GA or AA vs GG	0.76 (0.58–0.98)	0.037 / 1.00	1858A reduce TNF-a level in serum [51]	Reduced TNF-a level was associated with reduced risk of AS. This indicates that genetically determined high TNF-driven inflammatory response was associated with increased risk of AS.
И. 18	rs187238	Cytokines	GC or CC vs GG	0.80 (0.65-0.99) 0.044 / 1.00	0.044 / 1.00	-137C reduce IL-18 level in serum [53] and expression in PBMC [54]	Reduced IL-18 expression, and thus reduced IL-17 and TNF-a activity, was associated with reduced risk of AS. This indicates that a genetically determined high activity of the IL23/IL17 pathway was associated with increased risk of AS.
IL23R	rs11209026 Cytokines	Cytokines	GA or AA vs GG	0.60 (0.42-0.87) 0.0071 / 0.36	0.0071 / 0.36	rs11209026A reduce IL-17 level in PBMC [52]	Reduced IL-17 level was associated with reduced risk of AS. This indicates that a genetically determined high activity of the IL23/IL17 pathway was associated with increased risk of AS.
<i>OR</i> Odds ratio 95% <i>Cl</i> 95% cc <i>PBMC</i> peripher ^a The Bonferror ^b The <i>TNFRSF1</i> ,	OR Odds ratio 95% CI 95% confidence interval PBMC peripheral blood mononu The Bonferroni calculations wei ^b The TNFRSF1A (rs4149570) TT v	OR Odds ratio 95% Cl 95% confidence interval PBMC peripheral blood mononuclear cell $^{\text{The Bonferroni}}$ calculations were based on the 51 SNPs assessed in this study bThe TNFRSF1A (rs4149570) TT vs GG: OR: 1.79, 95% Cl: 1.31–2.41, p = 0.00027, Bonferroni = 0.014	SNPs assessed in thi CI: 1.31–2.41, <i>p</i> = 0.0	s study 00027, Bonferroni	= 0.014		

Haplotype	Haplotypes			N _{AS} (%)	N _{Control} (%)	OR ^a	(95% CI)	P-value
combinations	rs4696480 A > T	rs11938228 C > A	rs3804099 T>C ^b					
11	T:T	A:A	T:T	69 (11)	76 (10)	1.00	-	-
22	A:A	C:C	C:C	72 (11)	74 (10)	1.07	0.68–1.70	0.82
33	A:A	C:C	T:T	28 (4)	34 (5)	0.91	0.50–1.65	0.76
44	T:T	C:C	C:C	14 (2)	10 (1)	1.52	0.64-3.70	0.38
12	T:A	C:A	C:T	158 (24)	197 (27)	0.88	0.60-1.30	0.55
13	T:A	C:A	T:T	76 (12)	103 (14)	0.81	0.52-1.26	0.37
14	T:T	C:A	C:T	59 (9)	49 (7)	1.33	0.80-2.19	0.31
23	A:A	C:C	C:T	77 (12)	89 (12)	0.95	0.61-1.49	0.91
24	T:A	C:C	C:C	52 (8)	55 (8)	1.04	0.63-1.72	0.90
34	T:A	C:C	C:T	51 (8)	44 (6)	1.28	0.76-2.14	0.43

Table 3 Association of the *TLR2* haplotype combinations and risk of ankylosing spondylitis (AS). The haplotype combinations in *TLR2* described 93% of the genotypes observed

OR Odds ratio

^aOR was calculated for each haplotype combination by using the haplotype 11 as reference group

^bThe variant allele of rs3804099T T > C has been shown to decrease TNF- α , IL-1 β & IL-6 level [68]

used to test for deviation from Hardy-Weinberg equilibrium in the healthy controls and for haplotype analysis (Tables 3, 4, 5 and 6).

Statistical analyses were performed using STATA version 15 (StataCorp LP, College Station, TX, USA).

Results

Study population

Among the patients with AS the median age was 32 years (SD: 11.5) and 68% (483/709) were males. The healthy controls had a median age of 43 years (SD: 11.5) and 52% (411/384) were males. Among the patients 37% (118/323), 23% (73/323), and 41% (132/323) and among the controls 26% (207/788), 24% (189/788), and 50% (392/788) were current smokers, former smokers and never smokers,

respectively. HLA-B27 staus was available for 498 patients of which 83% (411/498) were positive. Sixty percent (427/709) of the patients were treated with anti-TNF.

The genotype distributions among the healthy controls deviated from Hardy-Weinberg equilibrium for *TLR1* (743 T > C (rs4833095)) (p = 0.03), *TLR2* (-16,934 A > T (rs4696480)) (p = 0.02), *TLR4* (rs1554973 T > C) (p = 0.03), *TLR9* (1174 G > A (rs352139)) (p = 0.02) and *TGFB1* (-509 C > T (rs1800469)) (p = 0.02). After correction for multiple testing, all SNPs studied were in Hardy-Weinberg equilibrium.

Polymorphisms associated with susceptibility of AS

In the age and sex adjusted analysis, the homozygous variant genotype of *TLR1* 743 T > C (rs4833095) (OR: 2.59,

Table 4 Association between *TLR4* haplotype combinations and risk of ankylosing spondylitis (AS). The haplotype combinations in *TLR4* described 94% of the genotypes observed

Haplotype	Haplotypes			N _{AS} (%)	N _{Control} (%)	OR ^a	(95% CI)	P-value
combinations	rs12377632 T > C	rs1554973 T > C	rs5030728 G > A					
11	C:C	T:T	G:G	95 (14)	101 (14)	1.00	_	-
22	T:T	T:T	A:A	69 (10)	74 (10)	0.99	0.64–1.53	1.00
33	T:T	C:C	G:G	29 (4)	57 (8)	0.54	0.32-0.92	0.03
44	T:T	T:T	G:G	3 (0)	5 (1)	0.64	0.15-2.74	0.72
12	T:C	T:T	G:A	154 (23)	188 (25)	0.87	0.61-1.24	0.47
13	T:C	T:C	G:G	126 (19)	129 (17)	1.04	0.72-1.51	0.85
14	T:C	T:T	G:G	30 (5)	32 (4)	1.00	0.56-1.77	1.00
23	T:T	T:C	G:A	99 (15)	106 (14)	0.99	0.67-1.47	1.00
24	T:T	T:T	G:A	31 (5)	24 (3)	1.37	0.75-2.51	0.36
34	T:T	T:C	G:G	28 (4)	26 (4)	1.14	0.63-2.09	0.76

OR Odds ratio

The biological effect of the three polymorphisms in TLR4 was unknown

^aOR was calculated for each haplotype combination by using the haplotype 11 as reference group

Haplotype combinations	Haplotypes			N _{AS} (%)	N _{Control} (%)	OR^{a}	(95% CI)	P-value
	rs4848306 -3737G > A [69, 70]	rs1143623 -1464G > C [69, 71]	rs1143627 -31 T > C [69, 71, 72]					
11	A:A	G:G	T:T	125 (18)	148 (20)	1.00	-	-
22	G:G	C:C	C:C	52 (8)	54 (7)	1.14	0.73-1.79	0.65
33	G:G	G:G	T:T	32 (5)	41 (5)	0.92	0.55-1.55	0.79
44	G:G	G:G	C:C	5 (1)	3 (0)	1.97	0.46-8.42	0.48
12	A:G	G:C	T:C	163 (24)	185 (24)	1.04	0.76-1.43	0.81
13	A:G	G:G	T:T	141 (20)	147 (19)	1.14	0.82-1.58	0.50
14	A:G	G:G	T:C	44 (6)	38 (5)	1.37	0.84-2.25	0.26
23	G:G	C:G	C:T	84 (12)	92 (12)	1.08	0.74–1.58	0.70
24	G:G	C:G	C:C	28 (4)	34 (4)	0.98	0.56-1.70	1.00
34	G:G	G:G	T:C	14 (2)	16 (2)	1.04	0.49-2.21	1.00

Table 5 Association between *IL1B* haplotype combinations and risk of ankylosing spondylitis (AS). The haplotype combinations in *IL1B* described 97% of the genotypes observed

OR Odds ratio

The variant allele of -3737 G > A [69], -1464 G > C [70] and -31 T > C [71, 72] have been shown to decrease IL-1 β level [69–72]

^aOR was calculated for each haplotype combination by using the haplotype 11 as reference group

95% CI: 1.48–4.51, *p* = 0.0008) and the combined homozygous and the heterozygous variant genotypes of TNFRSF1A -609 G > T (rs4149570) (OR: 1.44, 95% CI: 1.15-1.80, p = 0.001) were associated with increased risk of AS. The homozygous variant genotype of TLR4 T > C (rs1554973) (OR: 0.55, 95% CI: 0.34-0.86, p = 0.01) and LY96-1625 C > G (rs11465996) (OR: 0.68, 95% CI: 0.46-1.00, p = 0.05), and the combined homozygous and the heterozygous variant genotypes of TNF -308 G > A(rs1800629) (OR: 0.56, 95% CI: 0.44–0.72, p = 0.000005), TNF -238 G > A (rs361525) (OR: 0.49, 95% CI: 0.31-0.78, p = 0.002), *PTPN22* 1858 G > A (rs2476601) (OR: 0.76, 95% CI: 0.58–0.98, *p* = 0.04), *IL18*–137 G > C (rs187238) (OR: 0.80, 95% CI: 0.65–0.99, *p* = 0.04), and *IL23R* G > A (rs11209026) (OR: 0.60, 95% CI: 0.42–0.87, *p* = 0.01) were associated with reduced risk of AS (Table 1).

After Bonferroni correction for multiple testing the homozygous variant genotype of *TLR1* 743 T > C (rs4 833095) (OR: 2.59, 95% CI: 1.48–4.51, p = 0.04) and *TNFRSF1A* -609 G > T (rs4149570) (OR: 1.79, 95% CI:

1.31–2.41, p = 0.01) were associated with increased risk of AS and the combined homozygous and the heterozygous variant genotypes of *TNF* -308 G > A (rs1800629) (OR: 0.56, 95% CI: 0.44–0.72, p = 0.0002) were associated with reduced risk of AS (Table 2).

SNPs associated with AS and the biological effect of the SNPs are summarized in Table 2.

Haplotype analysis

Haplotype analyses of *TLR2*, *TLR4*, *IL1B* and *TNF* are shown in Tables 3, 4, 5 and 6, respectively.

The *TLR4* haplotype combination 33 (rs12377632TT, rs1554973CC and rs5030728GG) was associated with reduced risk of AS (OR: 0.54, 95% CI: 0.32–0.92, p = 0.03) compared to the haplotype combination 11. In *TNF* all haplotype combinations were associated with reduced risk of AS compared to the haplotype combination 11 (rs361525GG and rs1800629GG).

No associations were found for haplotype combinations of *TLR2* or *IL1B*.

Table 6 Association of the *TNF* haplotype combinations and risk of ankylosing spondylitis (AS). The haplotype combinations in *TNF* described 97% of the genotypes observed

Haplotype combinations	Haplotypes		N _{AS} (%)	N _{Control} (%)	OR ^a	(95% CI)	P-value
	rs361525 G>A ^b	rs1800629 G>A ^c					
11	G:G	G:G	523 (76)	469 (61)	1.00	_	=
22	G:G	A:A	9 (1)	25 (3)	0.32	(0.15–0.70)	0.005
12	G:G	G:A	125 (18)	210 (28)	0.53	(0.41–0.69)	< 0.0001
13	G:A	G:G	26 (4)	47 (6)	0.50	(0.30–0.81)	0.007
14	G:A	G:A	4 (1)	12 (2)	0.30	(0.10–0.93)	0.05

OR Odds ratio

^aOR was calculated for each haplotype combination by using the haplotype 11 as reference group

^bThe variant allele of TNF -238A rs361525A G > A has been shown to reduce expression of TNF- α [49]

^cThe variant allele of TNF -308A rs1800629 G > A has been shown to reduce mRNA level [48]

Discussion

In this case-control study, polymorphisms in a: the TNF- α (*TNF* (rs1800629 and rs361525), *TNFRSF1A* (rs4149570), and *PTPN22* (rs2476601)), b: the IL23/IL17 (*IL23R* (rs11 209026), and *IL18* (rs187238)), or c: the NFkB (*TLR1* (rs4833095), *TLR4* (rs1554973), and *LY96* (rs11465996)) pathways were associated with risk of AS.

The found associations for *TNF* (rs1800629) [19–22], *TNFRSF1A* (rs4149570) [23], and *IL23R* (rs11209026) [24–33] are in agreement with other case-control studies. Furthermore, Zhao et al. found that the variant allele of *NLRP3* (rs4612666) was associated with increased risk of AS in Chinese patients [23]. In our study we found a trend for associations of the variant allele of *NLRP3* (rs4612666) with increased risk of AS (p = 0.06). However, our results are in contrast to a meta-analysis of the *PTPN22* (rs2476601) polymorphism that did not find an association with AS [34]. Finally, we identified novel risk loci in *TNF* (rs361525), *IL18* (rs187238), *TLR1* (rs48 33095), *TLR4* (rs1554973), and *LY96* (rs11465996) that need validation in independent cohorts.

Most of the SNPs assessed in our study have known biological effects thus allowing a biological interpretation of the observed associations based on increased or reduced gene activity as summarized in Table 2 [35-47]. The associations observed for the TNF (rs1800629 and rs361525) polymorphisms suggest that reduced TNF- α mRNA level and expression of TNF-a was associated with reduced risk of AS [48, 49]. This is supported by our haplotype analysis which also suggests that the variant alleles of TNF rs1800629 and rs361525 were associated with reduced risk of AS. Likewise, the associations observed for the TNFRSF1A (rs4149570) polymorphism indicates that increased expression of the TNF-a receptor 1 was associated with increased risk of AS [50]. Furthermore, the associations observed for the PTPN22 (rs2476601) polymorphism suggests that reduced TNF- α serum level was associated with reduced risk of AS [51]. Taken together, this suggests that genetically determined high activity of the TNF- α pathway was associated with increased risk of AS.

IL-17 is known to induce the production of many cytokines including TNF-α [6]. IL-18 is a pro-inflammatory cytokine known to enhance the production of IL-17, TNF-α, and IL-1β [8]. In this study, the association observed for the *IL23R* (rs11209026) polymorphism suggests that reduced IL-17 serum level, and thus reduced TNF-α activity, was associated with reduced risk of AS [52]. Furthermore, the associations observed for the *IL18* (rs187238) polymorphism indicates that reduced IL-18 expression, and thus reduced IL-17 and TNF-α activity, was associated with reduced risk of AS [53, 54]. The associations found in the *IL23R* (rs11209026) and the *IL18* (rs187238) polymorphisms thus suggest that a genetically determined high activity of the IL23/IL17 pathway was associated with increased risk of AS. The two SNPs furthermore support that genetically determined high activity of the TNF- α pathway was associated with increased risk of AS. The observed associations between the polymorphisms in *IL23R* and *IL18* and risk of AS are in line with previous studies pointing out the IL23/IL17 pathway as central to the pathophysiology of AS [3, 4, 55].

This study also suggests that the NFkB pathway may be involved in the etiology of AS. The associations observed for the TLR1 (rs4833095) polymorphism suggests that increased TLR1 level was associated with increased risk of AS [56]. High level of TLR1 may lead to increased NFkB activation and thus increased TNF- α and IL-17 activity, which is in line with the other results. However, in contrast to the other results, the associations observed for the LY96 (rs11465996) polymorphism suggests that increased MD-2 (LY96) and TNF- α level was associated with a reduced risk of AS [57]. Finally, the TLR4 (rs1554973) polymorphism was associated with reduced risk of AS which was supported by the haplotype results (Table 4). The biological effect of the TLR4 (rs1554973) polymorphism is unknown, however, the result supports the notion that the NFkB pathway may be involved in the etiology of AS.

Both TNF- α [58] and interleukin-17 inhibitors [59] have been shown to reduce inflammation and improve symptoms in patients with AS [60]. Furthermore, increased levels of TNF- α , IL-17, IL-23, IL-1 β , and IL-6 have been found in sera and synovial fluid from AS patients [61–64]. The genetic associations between AS and the polymorphisms in *TLR1*, *TLR4*, *LY96*, *TNF*, *TNFRSF1A*, *IL18*, and *IL23R* found in this study, could potentially – in part – explain this altered cytokine milieu present in AS patients.

There are aspects of this study which should be interpreted with care. Conflicting results have been reported for the TNF (rs1800629) polymorphism [48, 49, 65]. Furthermore, the TNF polymorphisms, as well as the HLA-B27 locus, are located on chromosome 6, and there is a risk that even a minor linkage disequilibrium could have confounded our results [2]. TLR1 (rs4833095), TLR2 (rs4696480), TLR4 (rs1554973), TLR9 (rs352139), and TGFB1 (rs1800469) were not in Hardy-Weinberg equilibrium among the healthy controls. Due to the number of polymorphisms analyzed this is probably a type II error. The polymorphisms do not deviate from Hardy-Weinberg equilibrium when corrected for multiple testing. We cannot exclude that some of our positive findings may be due to chance due to the obtained p-values and the number of statistical tests performed. When the results were corrected for multiple testing only the variant allele of TLR1 (rs4833095) and TNFRSF1A (rs4149570) were associated with increased risk of AS and the variant allele of *TNF* (rs1800629) was associated with reduced risk of AS.

A major strength of this study was that the cohort was rather large including 709 patients with AS and 795 healthy controls and the associations that we report were biologically plausible. Also, the validity of the diagnosis is expected to be high, since the patients were identified via a clinical database that the rheumatologist use for prospective monitoring of patients as part of routine care [66].

Conclusions

In conclusion, we replicated associations between AS and the polymorphism *TNF* (rs1800629), *TNFRSF1A* (rs4149570), and *IL23R* (rs11209026). Furthermore, we identified novel risk loci in *TNF* (rs361525), *IL18* (rs187238), *TLR1* (rs4833095), *TLR4* (rs1554973), and *LY96* (rs11465996) that need validation in independent cohorts. The results suggest that genetically determined high activity of the TNF- α , IL23/IL17, and NFkB pathways increase the risk of AS.

Acknowledgments

We thank Ewa Kogutowska and Mette Errebo Rønne, Statens Serum Institut, for laboratory support; and Niels Steen Krogh, Zitelab Aps, Copenhagen, Denmark for database management. We also thank Department of Medicine, Viborg Regional Hospital, Denmark and OPEN (Odense Patient data Explorative Network), Odense University Hospital, Denmark for supporting this work. In memory of Niels Henrik Heegaard:

Co-author Niels H.H. Heegaard, Professor, MD, DMSc, DNatSc, died unexpectedly on September 26, 2017, at age 57. As director of the Department of Autoimmunology and Biomarkers, Statens Serum Institut, Copenhagen, Dr. Heegaard advanced research in autoimmunology and neurodegenerative disease. He had an extensive international research network and published more than 200 papers in scientific journals, focusing on biomarkers such as autoantibodies, microRNA, and microparticle proteins. He was a patient and unpretentious collaborator who always sought to highlight the work of other collaborators and co-workers. Dr. Heegaard was characterized

by humor, kindness, and optimism. He is survived by his wife and 2 children.

Funding

This study was funded by the Danish Rheumatism Association (A1923, A3037, and A3570 - www. Gigtforeningen.dk) and Region of Southern Denmark's PhD Fund, 12/7725 (www. Regionsyddanmark.dk).

Availability of data and materials

The datasets used during the current study are available from the corresponding author on reasonable request.

Authors' contributions

JS, SB, UV, PSA, SBS, HL, NHH and VA designed the research study and PSA, ABB, MRA, IB, RBD, HJH, BG and MLH collected the materials. JS and SB analysed the data and wrote the first draft. UV, PSA, SBS, ABB, MRA, IB, RBD, HJH, BG, MLH, HL and VA critically revised the manuscript. All authors agreed to be accountable for all aspects of the work and approved the final version of the manuscript.

Ethics approval and consent to participate

The study was conducted in accordance with the Declaration of Helsinki and was approved by the Regional Ethics Committees of Central (M20100153) and Southern (S-20120113) Denmark and the Danish Data Protection Agency of Central (RM: J. 2010–41-4719) and Southern (RSD: 2008–58-035) Denmark. For blood samples collected after routine TB screening, the Ethics Committees gave exemption from informed consent requirements because samples were taken as part of routine care and data were not identifiable. Written informed consent was obtained from patients donating blood samples at Frederiksberg Hospital as this involved collecting additional samples from patients.

Consent for publication

Not applicable.

Competing interests

VA receives compensation as a consultant and for being member of an advisory board for MSD and Janssen. BG has recived research funding from AbbVie, Biogen, Pfizer. The other authors declare no conflicts of interest.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Author details

Institute of Regional Health Research, University of Southern Denmark, Odense, Denmark. ²Department of Autoimmunology and Biomarkers, Statens Serum Institut, Copenhagen, Denmark. ³Department of Rheumatology, Frederiksberg Hospital, Frederiksberg, Denmark. ⁴Department of Rheumatology, Skåne University Hospital, Lund, Sweden. ⁵Focused Research Unit for Molecular Diagnostic and Clinical Research, Hospital of Southern Jutland, Aabenraa, Denmark. ⁶Medical Department, Viborg Regional Hospital, Viborg, Denmark. ⁷National Research Centre for the Working Environment, Copenhagen, Denmark. ⁸Microbiology and Infection Control, Statens Serum Institut, Copenhagen, Denmark. ⁹Veterinary Disease Biology, University of Copenhagen, Copenhagen, Denmark. ¹⁰Institute of Molecular Medicine, University of Southern Denmark, Odense, Denmark. ¹¹Department of Clinical Biochemistry, Herlev and Gentofte Hospital, Hellerup, Denmark. ²Department of Biochemistry, Hospital of Lillebaelt, Vejle, Denmark. ¹³Department of Clinical Microbiology, Slagelse Hospital, Slagelse, Denmark. ¹⁴Department of Clinical Medicine, Aarhus University, Aarhus, Denmark. ¹⁵Department of Respiratory Diseases B, Aarhus University Hospital, Aarhus, Denmark. ¹⁶Department of Rheumatology, Gentofte and Herlev Hospital, Hellerup, Denmark. ¹⁷The DANBIO Registry, Copenhagen Center for Arthritis Research, Center for Rheumatology and Spine Diseases, Rigshospitalet, Glostrup, Denmark. ¹⁸Department of Clinical Medicine, Faculty of Health and Medical Sciences, University of Copenhagen, Copenhagen, Denmark. ¹⁹Clinical Biochemistry, Clinical Institute, University of Southern Denmark, Odense, Denmark. ²⁰OPEN Odense Patient Data Explorative Network, Odense University Hospital, Odense, Denmark.

Received: 17 May 2018 Accepted: 3 September 2018 Published online: 12 September 2018

References

- Khan MA. Ankylosing spondylitis. In: Oxford University press; 2009. https:// global.oup.com/academic/product/ankylosing-spondylitis-9780195368079?cc=dk&lang=en&#
- Brown MA, Kenna T, Wordsworth BP. Genetics of ankylosing spondylitis-insights into pathogenesis. Nat Rev Rheumatol. 2016;12(2):81–91.
- Yago T, et al. IL-23 and Th17 disease in inflammatory arthritis. J Clin Med. 2017;6(9):E81.
- 4. Raychaudhuri SP, Raychaudhuri SK. Mechanistic rationales for targeting interleukin-17A in spondyloarthritis. Arthritis Res Ther. 2017;19(1):51.
- Verstrepen L, et al. TLR-4, IL-1R and TNF-R signaling to NF-kappaB: variations on a common theme. Cell Mol Life Sci. 2008;65(19):2964–78.
- 6. Hoeve MA, et al. Divergent effects of IL-12 and IL-23 on the production of IL-17 by human T cells. Eur J Immunol. 2006;36(3):661–70.
- Aguilera M, Darby T, Melgar S. The complex role of inflammasomes in the pathogenesis of inflammatory bowel diseases - lessons learned from experimental models. Cytokine Growth Factor Rev. 2014;25(6):715–30.
- Dinarello CA, et al. Interleukin-18 and IL-18 binding protein. Front Immunol. 2013;4:289.
- Hetland ML. DANBIO–powerful research database and electronic patient record. Rheumatology (Oxford). 2011;50(1):69–77.
- Bank S, et al. High-quality and -quantity DNA extraction from frozen archival blood clots for genotyping of single-nucleotide polymorphisms. Genet Test Mol Biomarkers. 2013;17(6):501–3.
- 11. Andersen V, et al. Polymorphisms in NF-kappaB, PXR, LXR, PPARgamma and risk of inflammatory bowel disease. World J Gastroenterol. 2011;17(2):197–206.
- 12. Ernst A, et al. Common polymorphisms in the microsomal epoxide hydrolase and N-acetyltransferase 2 genes in association with inflammatory

bowel disease in the Danish population. Eur J Gastroenterol Hepatol. 2011; 23(3):269–74.

- Andersen V, et al. Cyclooxygenase-2 (COX-2) polymorphisms and risk of inflammatory bowel disease in a Scottish and Danish case-control study. Inflamm Bowel Dis. 2011;17(4):937–46.
- Andersen V, et al. The polymorphism rs3024505 proximal to IL-10 is associated with risk of ulcerative colitis and Crohns disease in a Danish case-control study. BMC Med Genet. 2010;11:82.
- Ernst A, et al. Genetic variants of glutathione S-transferases mu, theta, and pi display no susceptibility to inflammatory bowel disease in the Danish population. Scand J Gastroenterol. 2010;45(9):1068–75.
- 16. Ostergaard M, et al. Cyclooxygenase-2, multidrug resistance 1, and breast cancer resistance protein gene polymorphisms and inflammatory bowel disease in the Danish population. Scand J Gastroenterol. 2009;44(1):65–73.
- Ernst A, et al. Mutations in CARD15 and smoking confer susceptibility to Crohn's disease in the Danish population. Scand J Gastroenterol. 2007;42(12):1445–51.
- Exarchou S, et al. The prevalence of clinically diagnosed ankylosing spondylitis and its clinical manifestations: a nationwide register study. Arthritis Res Ther. 2015;17:118.
- Manolova I, et al. Association of single nucleotide polymorphism at position –308 of the tumor necrosis factor-alpha gene with ankylosing spondylitis and rheumatoid arthritis. Biotechnol Biotechnol Equip. 2014;28(6):1108–14.
- Hohler T, et al. Association of different tumor necrosis factor alpha promoter allele frequencies with ankylosing spondylitis in HLA-B27 positive individuals. Arthritis Rheum. 1998;41(8):1489–92.
- McGarry F, et al. The –308.1 polymorphism in the promoter region of the tumor necrosis factor gene is associated with ankylosing spondylitis independent of HLA-B27. J Rheumatol. 1999;26(5):1110–6.
- Milicic A, et al. Interethnic studies of TNF polymorphisms confirm the likely presence of a second MHC susceptibility locus in ankylosing spondylitis. Genes Immun. 2000;1(7):418–22.
- 23. Zhao S, et al. The association of NLRP3 and TNFRSF1A polymorphisms with risk of ankylosing spondylitis and treatment efficacy of etanercept. J Clin Lab Anal. 2017;31(6). https://doi.org/10.1002/jcla.22138. Epub 23 Jan 2017.
- 24. Abdollahi E, et al. Protective role of R381Q (rs11209026) polymorphism in IL-23R gene in immune-mediated diseases: a comprehensive review. J Immunotoxicol. 2016;13(3):286–300.
- Roberts AR, et al. Investigation of a possible extended risk haplotype in the IL23R region associated with ankylosing spondylitis. Genes Immun. 2017; 18(2):105–8.
- Rueda B, et al. The IL23R Arg381GIn non-synonymous polymorphism confers susceptibility to ankylosing spondylitis. Ann Rheum Dis. 2008;67(10):1451–4.
- 27. Rahman P, et al. Association of interleukin-23 receptor variants with ankylosing spondylitis. Arthritis Rheum. 2008;58(4):1020–5.
- Karaderi T, et al. Association between the interleukin 23 receptor and ankylosing spondylitis is confirmed by a new UK case-control study and meta-analysis of published series. Rheumatology (Oxford). 2009; 48(4):386–9.
- Safrany E, et al. Variants of the IL23R gene are associated with ankylosing spondylitis but not with Sjogren syndrome in Hungarian population samples. Scand J Immunol. 2009;70(1):68–74.
- Duan Z, et al. Interleukin-23 receptor genetic polymorphisms and ankylosing spondylitis susceptibility: a meta-analysis. Rheumatol Int. 2012;32(5):1209–14.
- Lee YH, et al. Associations between interleukin-23R polymorphisms and ankylosing spondylitis susceptibility: a meta-analysis. Inflamm Res. 2012; 61(2):143–9.
- Brionez TF, Reveille JD. The contribution of genes outside the major histocompatibility complex to susceptibility to ankylosing spondylitis. Curr Opin Rheumatol. 2008;20(4):384–91.
- Burton PR, et al. Association scan of 14,500 nonsynonymous SNPs in four diseases identifies autoimmunity variants. Nat Genet. 2007;39(11):1329–37.
- 34. Wang W, et al. Association between protein tyrosine phosphatase nonreceptor type 22 (PTPN22) polymorphisms and risk of Ankylosing spondylitis: a meta-analysis. Med Sci Monit. 2017;23:2619–24.
- Bank, S, et al. Polymorphisms in the inflammatory pathway genes TLR2, TLR4, TLR9, LY96, NFKBIA, NFKB1, TNFA, TNFRSF1A, IL6R, IL10, IL23R, PTPN22, and PPARG are associated with susceptibility of inflammatory bowel disease in a Danish cohort. PLoS One. 2014;9(6):e98815.
- Bank, S, et al. Polymorphisms in the toll-like receptor and the IL-23/IL-17 pathways were associated with susceptibility to inflammatory bowel disease in a Danish cohort. PLoS One. 2015;10(12):e0145302.

- 37. Bank S, et al. Associations between functional polymorphisms in the NFkappaB signaling pathway and response to anti-TNF treatment in Danish patients with inflammatory bowel disease. Pharmacogenomics J. 2014;14(6):526–34.
- Bank S, et al. Genetically determined high activity of IL-12 and IL-18 in ulcerative colitis and TLR5 in Crohns disease were associated with nonresponse to anti-TNF therapy. Pharmacogenomics J. 2018;18(1):87–97.
- Bank S, et al. Effectiveness of anti-tumour necrosis factor-alpha therapy in Danish patients with inflammatory bowel diseases. Dan Med J. 2015;62(3): A4994.
- Bank S. A cohort of anti-TNF treated Danish patients with inflammatory bowel disease, used for identifying genetic markers associated with treatment response. Dan Med J. 2015;62(5):85087.
- Sode J, et al. Anti-TNF treatment response in rheumatoid arthritis patients is associated with genetic variation in the NLRP3-inflammasome. PLoS One. 2014;9(6):e100361.
- Sode J, et al. Genetic variations in pattern recognition receptor loci are associated with anti-TNF response in patients with rheumatoid arthritis. PLoS One. 2015;10(10):e0139781.
- Sode J, et al. Confirmation of an IRAK3 polymorphism as a genetic marker predicting response to anti-TNF treatment in rheumatoid arthritis. Pharmacogenomics J. 2018;18(1):81–6.
- Loft ND, et al. Associations between functional polymorphisms and response to biological treatment in Danish patients with psoriasis. Pharmacogenomics J. 2018;18(3):494–500.
- Bek S, et al. Systematic review: genetic biomarkers associated with anti-TNF treatment response in inflammatory bowel diseases. Aliment Pharmacol Ther. 2016;44(6):554–67.
- Bek S, et al. Systematic review and meta-analysis: pharmacogenetics of anti-TNF treatment response in rheumatoid arthritis. Pharmacogenomics J. 2017; 17(5):403–11.
- Loft ND, et al. Genetic polymorphisms associated with psoriasis and development of psoriatic arthritis in patients with psoriasis. PLoS One. 2018; 13(2):e0192010.
- Oliveira JM, et al. The –308 bp TNF gene polymorphism influences tumor necrosis factor expression in leprosy patients in Bahia state, Brazil. Infect Genet Evol. 2016;39:147–54.
- Kaluza W, et al. Different transcriptional activity and in vitro TNF-alpha production in psoriasis patients carrying the TNF-alpha 238A promoter polymorphism. J Invest Dermatol. 2000;114(6):1180–3.
- Wang GB, et al. A regulatory polymorphism in promoter region of TNFR1 gene is associated with Kawasaki disease in Chinese individuals. Hum Immunol. 2011;72(5):451–7.
- Kariuki SN, Crow MK, Niewold TB. The PTPN22 C1858T polymorphism is associated with skewing of cytokine profiles toward high interferon-alpha activity and low tumor necrosis factor alpha levels in patients with lupus. Arthritis Rheum. 2008;58(9):2818–23.
- Oosting M, et al. Role of interleukin-23 (IL-23) receptor signaling for IL-17 responses in human Lyme disease. Infect Immun. 2011;79(11):4681–7.
- Jaiswal PK, et al. Association of IL-12, IL-18 variants and serum IL-18 with bladder cancer susceptibility in north Indian population. Gene. 2013;519(1):128–34.
- Dziedziejko V, et al. The impact of IL18 gene polymorphisms on mRNA levels and interleukin-18 release by peripheral blood mononuclear cells. Postepy Hig Med Dosw (Online). 2012;66:409–14.
- Sherlock JP, et al. IL-23 induces spondyloarthropathy by acting on ROR-gammat+ CD3+CD4-CD8- entheseal resident T cells. Nat Med. 2012;18(7):1069–76.
- Uciechowski P, et al. Susceptibility to tuberculosis is associated with TLR1 polymorphisms resulting in a lack of TLR1 cell surface expression. J Leukoc Biol. 2011;90(2):377–88.
- Gu W, et al. Functional significance of gene polymorphisms in the promoter of myeloid differentiation-2. Ann Surg. 2007;246(1):151–8.
- Maxwell LJ, et al. TNF-alpha inhibitors for ankylosing spondylitis. Cochrane Database Syst Rev. 2015;18(4):Cd005468.
- Pavelka K, et al. Efficacy, safety, and tolerability of secukinumab in patients with active ankylosing spondylitis: a randomized, double-blind phase 3 study, MEASURE 3. Arthritis Res Ther. 2017;19(1):285.
- 60. Cheung PP. Anti-IL17A in axial Spondyloarthritis-where are we at? Front Med (Lausanne). 2017;4:1.
- Jandus C, et al. Increased numbers of circulating polyfunctional Th17 memory cells in patients with seronegative spondylarthritides. Arthritis Rheum. 2008;58(8):2307–17.

- Singh R, Aggarwal A, Misra R. Th1/Th17 cytokine profiles in patients with reactive arthritis/undifferentiated spondyloarthropathy. J Rheumatol. 2007; 34(11):2285–90.
- Xueyi L, et al. Levels of circulating Th17 cells and regulatory T cells in ankylosing spondylitis patients with an inadequate response to anti-TNF-alpha therapy. J Clin Immunol. 2013;33(1):151–61.
- 64. Londono J, et al. The association between serum levels of potential biomarkers with the presence of factors related to the clinical activity and poor prognosis in spondyloarthritis. Rev Bras Reumatol. 2012;52(4):536–44.
- Karimi M, et al. A critical assessment of the factors affecting reporter gene assays for promoter SNP function: a reassessment of -308 TNF polymorphism function using a novel integrated reporter system. Eur J Hum Genet. 2009;17(11):1454–62.
- 66. Ibfelt EH, et al. Validity and completeness of rheumatoid arthritis diagnoses in the nationwide DANBIO clinical register and the Danish National Patient Registry. Clin Epidemiol. 2017;9:627–32.
- Gast A, et al. Association of inherited variation in toll-like receptor genes with malignant melanoma susceptibility and survival. PLoS One. 2011;6(9): e24370.
- 68. Zhang F, et al. Polymorphisms in toll-like receptors 2, 4 and 5 are associated with legionella pneumophila infection. Infection. 2013;41(5):941–8.
- Chen H, et al. Single nucleotide polymorphisms in the human interleukin-1B gene affect transcription according to haplotype context. Hum Mol Genet. 2006;15(4):519–29.
- Yoshida M, et al. Haplotypes in the expression quantitative trait locus of interleukin-1beta gene are associated with schizophrenia. Schizophr Res. 2012;140(1–3):185–91.
- Wen AQ, et al. Clinical relevance of IL-1beta promoter polymorphisms (-1470, -511, and -31) in patients with major trauma. Shock. 2010;33(6): 576–82.
- Lind H, Haugen A, Zienolddiny S. Differential binding of proteins to the IL1B -31 T/C polymorphism in lung epithelial cells. Cytokine. 2007;38(1):43–8.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

