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Identification of genetic variants associated with dengue or West Nile virus disease: a systematic review and meta-analysis

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Abstract

Background: Dengue and West Nile viruses are highly cross-reactive and have numerous parallels in geography, potential vector host (*Aedes* family of mosquitoes), and initial symptoms of infection. While the vast majority (> 80%) of both dengue and West Nile virus infections result in asymptomatic infections, a minority of individuals experience symptomatic infection and an even smaller proportion develop severe disease. The mechanisms by which these infections lead to severe disease in a subset of infected individuals is incompletely understood, but individual host differences including genetic factors and immune responses have been proposed. We sought to identify genetic risk factors that are associated with more severe disease outcomes for both viruses in order to shed light on possible shared mechanisms of resistance and potential therapeutic interventions.

Methods: We applied a search strategy using four major databases (Medline, PubMed, Embase, and Global Health) to find all known genetic associations identified to date with dengue or West Nile virus disease. Here we present a review of our findings and a meta-analysis of genetic variants identified.

Results: We found genetic variations that are significantly associated with infections of these viruses. In particular we found variation within the OAS1 (meta-OR = 0.83, 95% CI: 0.69–1.00) and CCR5 (meta-OR = 1.29, 95% CI: 1.08–1.53) genes is significantly associated with West Nile virus disease, while variation within MICB (meta-OR = 2.35, 95% CI: 1.68–3.29), PLCE1 (meta-OR = 0.55, 95% CI: 0.42–0.71), MBL2 (meta-OR = 1.54, 95% CI: 1.02–2.31), and IFN- γ (meta-OR = 2.48, 95% CI: 1.30–4.71), is associated with dengue disease.

Conclusions: Despite substantial heterogeneity in populations studied, genes examined, and methodology, significant associations with genetic variants were found across studies within both diseases. These gene associations suggest a key role for immune mechanisms in susceptibility to severe disease. Further research is needed to elucidate the role of these genes in disease pathogenesis and may reveal additional genetic factors associated with disease severity.

Keywords: Dengue virus, West Nile virus, Disease severity, Genetic variation, Meta-analysis, Single nucleotide polymorphism

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Background

Dengue (DENV) and West Nile (WNV) viruses are mosquito-borne viruses in the *Flaviviridae* family, which also includes other viruses such as Zika and yellow fever. These viruses can cause disease with substantial public health impact. DENV and WNV are found in similar areas of the world, can be carried by the *Aedes* family of mosquitoes, have similar initial stages of infections and similar symptoms of mild febrile illness, and are highly cross-reactive; however, severe disease manifests differently for these two viruses [1–3]. West Nile Virus was first identified in Uganda in 1937, has been endemic in the United States since 1999 [4], and is estimated to have infected 3 million people [5]. While the majority of infections are asymptomatic, ~20% of infections lead to mild febrile disease in infected individuals and 1% of infected individual experience severe, neurological disease such as meningitis and encephalitis [6]. DENV has a vastly higher disease burden, with an estimated 50 million cases and 25,000 fatalities worldwide annually [7, 8]. The majority of DENV infections can be classified as asymptomatic or mild febrile illness, with approximately <1% progressing to Dengue Hemorrhagic Fever (DHF) or Dengue Shock Syndrome (DSS). DHF is delineated from mild DENV febrile illness by the increase in vascular permeability, while DSS has the additional development of circulatory shock [7, 8].

For both WNV and DENV, known risk factors such as immune-compromised states or advanced age are associated with susceptibility to mild and severe disease [9, 10]. The mechanisms by which an infection leads to severe disease in a subset of all infected individuals is incompletely explained. Differing immune responses to infections, including elevated cytokine responses, have been proposed [11–13] and we have recently shown that geographic location is not a driver of severity of WNV infection in a localized region [14]. In addition to similarities in the early stages of infection [15–17], both viruses induce strong immune responses including chemokines (such as IL-8) and cytokines which up-regulate inflammatory reaction (such as TNF- α , IL-1, IL-6, and IFN- β) [18–21]. Renewed interest in understanding flaviviral infection and disease susceptibility comes as climate change expands the number of individuals at risk of exposure to WNV and DENV [3, 22], and with outbreaks of related flaviviruses, most notably Zika [23, 24].

Genetic differences are additional explanations of individual susceptibility to symptomatic disease, and previous genome-wide association studies (GWAS) and candidate-gene studies have identified genetic factors associated with DENV or WNV disease pathogenesis. To assess the current state of knowledge on genetic variation associated with these flaviviral diseases, and to identify any shared features of anti-viral responses, we

conducted a systematic review and meta-analysis of the published associations to date between genetic variants and development of DENV or WNV disease.

Methods

A systematic review of genetic factors and WNV or DENV disease was conducted using the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) (Additional file 1) [25].

Search strategy

Medline, PubMed, Embase, and Global Health databases were used to search the literature. Search terms included West Nile or DENV and genetic factors; the same set of text words was used for all databases in conjunction with subject headings that were tailored for each database. The text word search specified West Nile or Dengue in the title, a genetic term in the title or abstract, and a human-related term in the title or abstract (Table 1). A sample search strategy is included in the appendix (Additional file 2). Case-control studies which examined at least one genetic factor associated with either viral disease were included. Studies on non-human (e.g., viral, mosquito) genetics and case reports on single patients were excluded. Reports published prior to May 2017 were included in the review. An ancestry search was done of references of selected studies to collect additional potentially relevant references.

Table 1 Text word selection for search of selected databases.

Text words used for the search strategy, with one term from each column required in the title for the viral term, or in the title or abstract for the genetic and human terms

Viral terms	Genetic Factor terms	Human-related terms
<ul style="list-style-type: none"> • West Nile • Dengue 	<ul style="list-style-type: none"> ▪ microsatellite(s) ▪ genetic variation ▪ genetic factor(s) ▪ genetic marker(s) ▪ genetic analysis/analyses ▪ SNP(s) ▪ single nucleotide polymorphism(s) ▪ copy number variant(s) ▪ genetic predisposition ▪ genetic susceptibility ▪ disease susceptibility ▪ GWAS ▪ genome-wide association study/studies ▪ genetic association(s) ▪ genetic association study/studies ▪ candidate gene study/studies ▪ genetic predisposition to disease ▪ susceptibility to disease ▪ genetic variability ▪ gene identity 	<ul style="list-style-type: none"> ▪ human ▪ man/men ▪ woman/women ▪ child/children ▪ teenager(s) ▪ middle-aged ▪ elderly ▪ infant(s) ▪ male(s) ▪ female(s) ▪ patient(s) ▪ participant(s) ▪ citizen(s) ▪ subject(s) ▪ case(s) ▪ control(s)

Study selection and data extraction

Two researchers reviewed the titles and abstracts of all studies and identified potentially relevant articles within Covidence with 98.6% consistency [26]. Discrepancies were resolved through re-review and mutual consensus. Both researchers read the full text of all of the selected potentially relevant articles and identified the final reports to be included in this review. Data sets were extracted without personal identifiers and organized into literature tables. The main fields included authors, year of publication, country, sample size, case and control group definitions, genotyping method, genes and genetic variants analyzed, genotype count data when available, odds ratios (OR), and statistical analysis method.

When two or more studies examined the same variants, we used the raw genotype data to calculate ORs with 95% confidence intervals using the R package EpiTools [27]. When the raw genotype data were not available within the published papers, we requested the data sets from corresponding authors of the studies. Of the six authors contacted, three shared data, two indicated they no longer had access to the data, and one did not respond by date of submission. In order to make comparisons across the different DENV phenotypes used in the studies, we compared asymptomatic DENV infections and controls with all symptomatic infections (DENV fever, DENV hemorrhagic fever, and DENV shock syndrome). Using the genotype data, we calculated ORs for each study under a dominant model, recessive model, homozygote mutant versus homozygote wild-type, and heterozygote versus homozygote wild-type. We meta-analyzed the ORs using RevMan [28]. The genetic model with the most significant meta-OR is presented here. When this model was the homozygote mutant versus homozygote wild or the heterozygote versus homozygote wild, we included both of these models for that particular single nucleotide polymorphism (SNP).

Quality assessment

We assessed the quality of each study with the Newcastle-Ottawa Quality Assessment Scale for Case-Control Studies [29], which assesses each study's selection, comparability, and exposure ascertainment approach.

Results

To identify all published research assessing the role of genetic variation with DENV or WNV disease, we executed the above search strategy and identified 633 published reports (Table 1, Fig. 1). Two researchers independently reviewed the titles and abstracts of these 633 papers and identified 104 papers for further full-text review in this meta-analysis (Additional file 3). One

additional paper from 1987 was identified as pertinent during the ancestry search and was added to the review. The final analysis includes data from 87 of the 105 publications, following exclusion of 18 papers for cause (seven repeats, six conference abstracts, and five with an outcome other than disease severity). Reflecting the higher disease burden and longer research history of DENV virus, of these 87 papers selected, 74 studied DENV-infected populations and 13 focused on WNV.

HLA genetic variation associated with disease severity

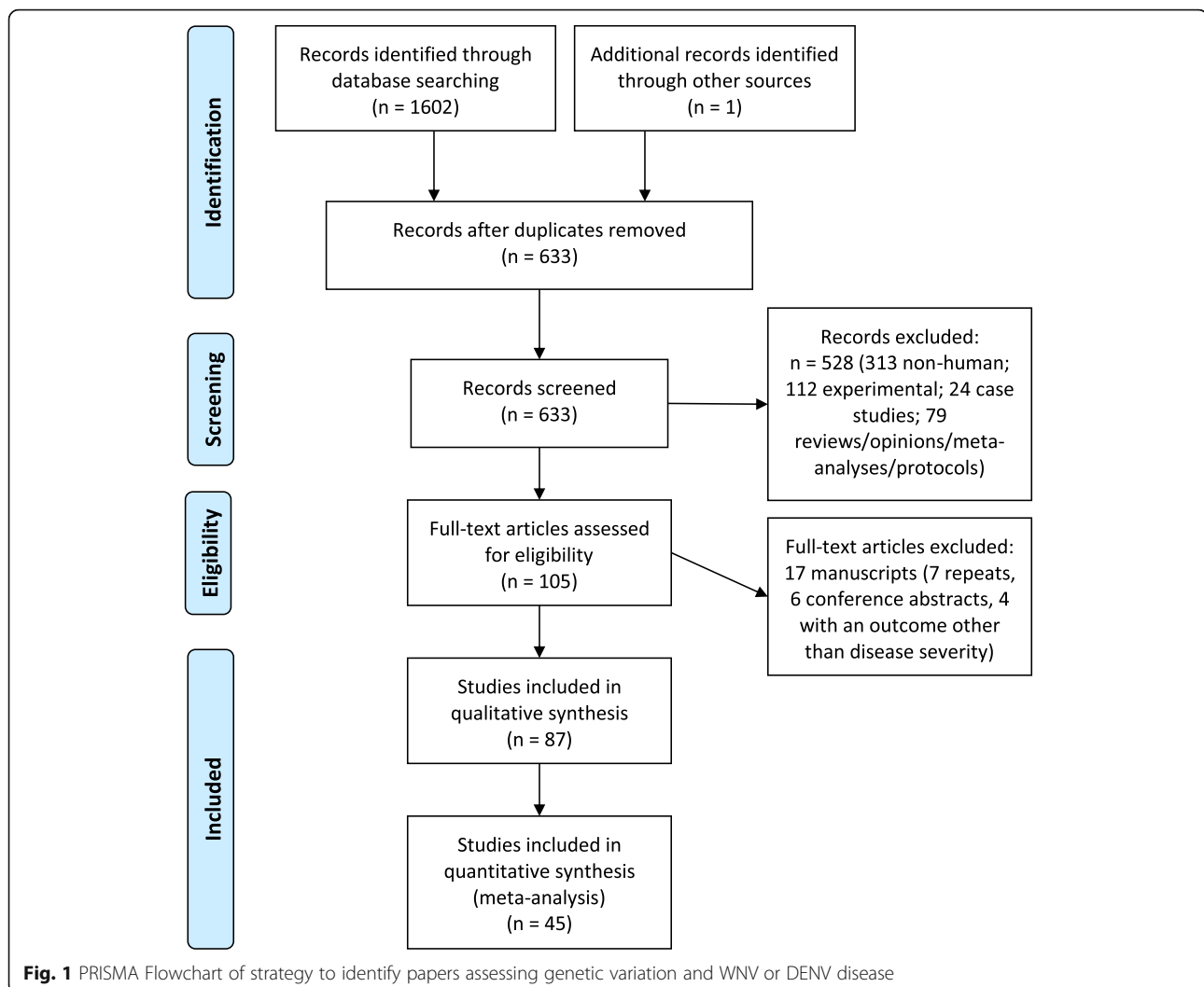
Notably, 27 separate HLA alleles were examined by two or more research groups for an association with severe DENV disease (Additional file 4). Four research groups analyzed HLA alleles for an association with WNV disease (Additional file 5), however there was no overlap in the alleles studied. Although HLA variants show substantial contribution to disease outcome, significant variations in study design, data analysis platforms, data availability and presentation precluded our in-depth meta-analysis of these data.

Multiple genes are associated with severity of WNV infection

Previous reports of genetic associations with WNV disease severity focused on U.S. or Canadian populations and compared severe and non-severe infections. Overall, these studies identified 12 gene variants and significant findings include SNPs of multiple immune-related genes such as RFC1, SCN1A, and IRF3 (Table 2).

OAS1 and CCR5 have significant associations with WNV disease across multiple studies

For genes with genotype count data available for ≥ 2 studies, we conducted a meta-analysis of genetic association to disease severity. Meta-analysis allows recognition of well-established genetic associations and identification of redundant studies for genes with null associations. We found that SNPs in MX1, OASL, OAS1, RFC1, and CCR5 were studied by multiple research groups for an association with WNV disease (Table 2). To assess the overall association of these SNPs with WNV disease, we calculated a combined OR for each gene based on the genotype counts under four different genetic models. Of these, CCR5 and OAS1 meta-ORs were significant under a dominant model, with meta-OR of 0.83 [95% CI: 0.69–1.00] and 1.29 [1.08–1.53], respectively (Fig. 2). The CCR5 meta-OR was also significant under an allelic model with a meta-OR of 1.22 [95% CI: 1.03–1.44]. The CCR5 delta 32 deletion is associated with more severe disease while the OAS1 allele G was associated with less severe disease.



Multiple genes are associated with severity of DENV infection

Seventy-four studies have examined genetic associations with DENV disease severity and more than 30 genes have been implicated in DENV disease (Additional file 3). SNPs that were studied by only a single research group are presented in Table 3. We also include SNPs studied by multiple research groups, but for which genotype data was unavailable or the comparison groups of multiple studies could not be analyzed together.

Significant associations with DENV disease

Among the DENV studies, the same variant within 17 genes was studied by two or more research groups (Table 3). Four genes had significant meta-ORs (Fig. 3). For a SNP in MBL2 (exon 1), we calculated a meta-OR of 1.54 [1.02–2.31] under a dominant model and 1.65 [1.18–2.32] under an allelic model, with alleles other than the A allele being associated with more severe disease. The T allele for SNP rs2430561 in the IFN- γ gene

was associated with severe disease under a recessive model with a meta-OR of 2.48 [0.30–4.71]. For a SNP located within MICB (rs3132468), we found the CC genotype had a significantly greater association with severe disease (meta-OR 2.35 [1.68–3.29]), but the heterozygote genotype showed no significant association with disease severity as compared to the TT genotype (meta-OR = 1.17 [0.86–1.59]). For this SNP, the C allele was also found to be significantly associated with disease as compared to the T allele (meta-OR = 1.35 [1.16–1.57]). For two SNPs located within the PLCE1 gene, every model tested was significant, with the most significant meta-ORs being 0.62 [0.48–0.79] for TT genotype as compared to CC genotype for rs3740360 and 0.55 [0.42–0.71] under a recessive model for rs3765524. TNF- α (rs1800629 and rs361525) was the most studied gene, but none of the models tested provided a significant meta-OR.

Quality scores

Based on the Newcastle Ottawa Scoring System, the average quality score was 5.76 (range: 3–7) for the WNV

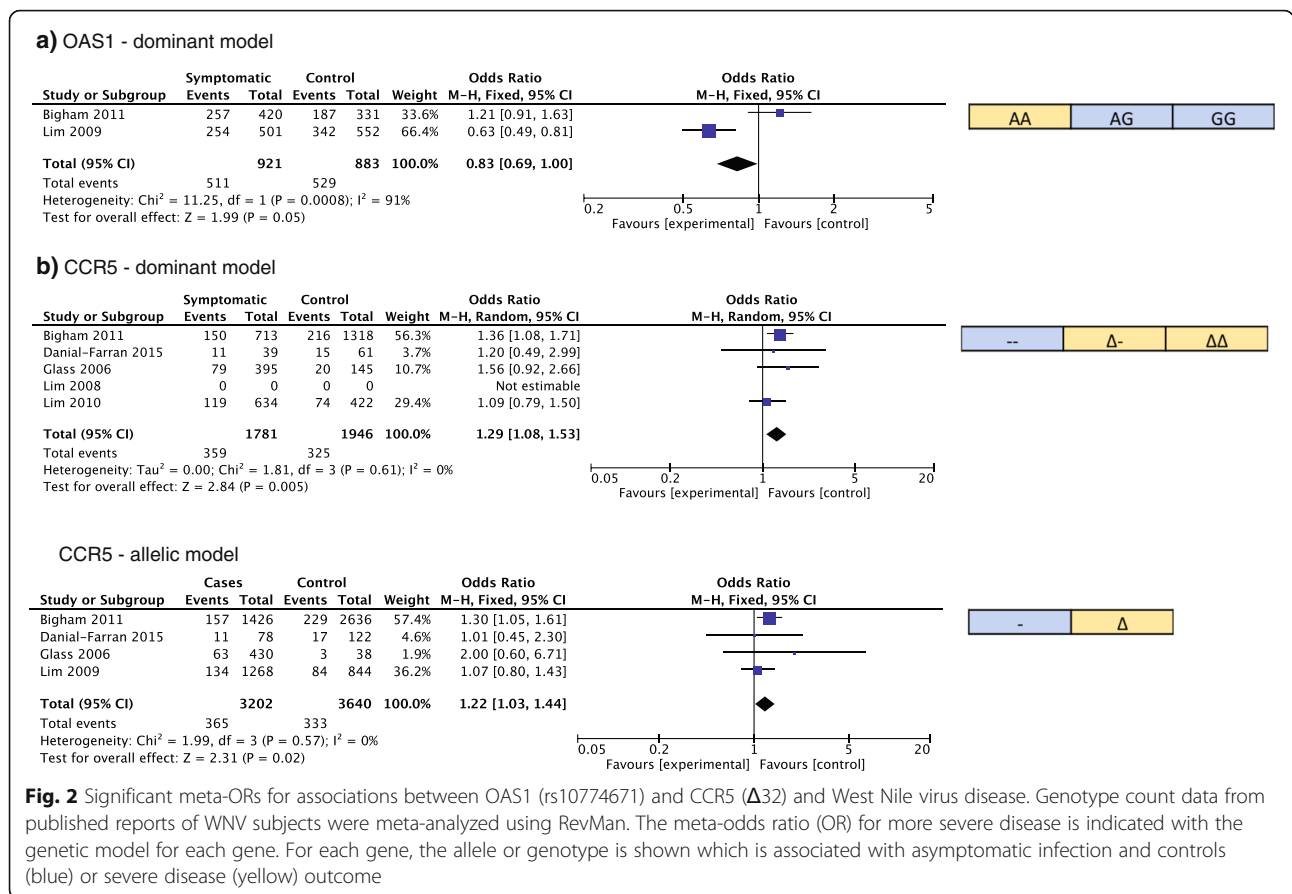
Table 2 Genetic variation significantly associated with West Nile virus disease. We include in this table all variants studied by two or more research groups and variants found to have a significant association by one research group

Gene	Genetic variant	Entrez Gene ID [31]	Major Allele	Minor Allele	Number of Cases	Number in Comparison Group	Country	Key Results	Included in meta-analysis
ANPEP	rs25651	290	T	C	560 severe	950 non-severe infections	US & Canada	0.69 odds of severe disease [59]	No ^a
CACNA1H	rs113802594	8912	A	G	1330 severe	61 controls	Israel	No significant association with disease [60]	No
CCRS5	CCRS5 Δ32	1234	-	Δ32 deletion	560 severe	919 non-severe infections	US & Canada	8.58 odds of encephalitis [61]	No
				Δ32 deletion	39 severe	950 non-Severe infections	US & Canada	No significant association with disease [59]	Yes
				Δ32 deletion	422 symptomatic	61 controls	Israel	No significant association with disease [60]	
				Δ32 deletion	634 infections	331 asymptomatic infections	US & Canada	No significant association with disease [62]	
				Δ32 deletion		422 controls	US	No significant association with disease, but significant association with more severe disease ($p = 0.0016$) [46]	
				Δ32 deletion	395 symptomatic (two cohorts of 247 and 148)	1318 controls	US	Significantly associated with disease in two cohorts (OR = 4.4 [1.6–11.8] and OR = 9.1 [3.4–24.8]), and fatal outcomes in one cohort with OR = 13.2 [1.9–89.9] [63]	
HERC5	rs148556308	51,191	A	G	1330 severe	919 non-severe infections	US & Canada	Significantly associated with severe disease (p -value = 6.5 × 10 ⁻¹⁰) [61]	No
IRF3	rs2304207	3661	G	C	422 severe	331 asymptomatic infections	US	0.52 odds of symptomatic infection under dominant model [62]	No ^a
				C	39 severe	61 controls	Israel	No significant association with disease [60]	
MIF	rs5844572	4282	5 or 6 CATT repeats	7 CATT repeats	518 severe	514 non-severe	US & Canada	1.73 odds of encephalitis among patients with high-expression allele as compared to all other types of WNV disease [64]	No
MX1	rs7280422	4599	C	G	39 severe	61 controls	Israel	4.05 odds of infection associated with variant allele [60]	No ^a
OASL	rs3213545	8638	C	T	422 severe	331 asymptomatic infections	US	0.25 odds of symptomatic infection under a recessive model [62]	Yes
						331 asymptomatic infections	US	No significant association with disease [62]	Yes

Table 2 Genetic variation significantly associated with West Nile virus disease. We include in this table all variants studied by two or more research groups and variants found to have a significant association by one research group (Continued)

Gene	Genetic variant	Entrez Gene ID [31]	Major Allele	Minor Allele	Number of Cases	Number in Comparison Group	Country	Key Results	Included in meta-analysis
OAS1	rs10774671	4938	C	T	39 severe	61 controls	Israel	1.85 (1.03–3.3) odds of infection [60]	
			C	T	33 symptomatic	60 controls	US	Significantly associated with disease ($P < 0.004$) [65]	Yes
			A	G	422 severe	331 asymptomatic infections	US	No significant association with disease [62]	Yes
OAS1	rs34137742	4938	A	G	39 severe	61 controls	Israel	No significant association with disease [60]	
			A	G	501 seropositive	552 controls	US	1.6 [95% CI 1.2–2.0] odds of seroconversion [66]	
			C	T	422 severe	331 asymptomatic infections	US	9.79 [95% CI 3.60–26.61] odds of encephalitis and paralysis [62]	Yes
RFC1	rs2066786	5981	C	T	39 severe	61 controls	Israel	No significant association with disease [60]	
			T	C	560 severe	950 non-severe infections	US & Canada	0.68 odds of severe disease associated with minor allele [59]	No ^a
			G	A	39 severe	61 controls	Israel	2.8 odds under dominant model [60]	
SCN1A	rs2298771	6323	C	T	560 severe	950 non-severe infections	US & Canada	1.47 odds of severe disease associated with minor allele [59]	No ^a
			A	G	39 severe	61 controls	Israel	No significant association with disease [60]	
			A	T	1330 severe	919 non-severe infections	US & Canada	3.57 odds of severe disease and 4.94 odds of Acute Flaccid Paralysis than controls [61]	No

^agenotype data not available for meta-analysis



publications and 5.10 (range: 2–7) for the DENV publications (Additional file 6). We also assessed whether the study authors corrected for multiple testing, and found less than half of both WNV and DENV studies provided corrected *p*-values when appropriate, indicating an inflated type I error rate.

Discussion

We have examined genetic variants that show association with DENV or WNV disease severity. This analysis was undertaken to identify genetic differences that are significant drivers of susceptibility to symptomatic disease that may shed light on mechanisms of immune resistance to these viruses. Among the 87 studies examined, a wide range of genetic targets was found to be significant, with many of the genes unsurprisingly playing a key role in the immune system defense against viral infections (Additional file 7).

Despite the large number of studies, only 27 genes were studied by more than one research group for an association with either disease. Throughout these studies, several key genes rose to the forefront as the most studied and the most significant associations. Many studies focused on the HLA region of the genome, and, although inconsistencies in data presentation preclude a

meta-analyze of these results, there were clear signs of the importance of this area for both diseases.

With the central role of HLA for the immune system, polymorphisms in this region have been well studied for associations with disease. The area is highly polymorphic, however, leading to difficulties for comparing the diverse range of alleles. Adding to this complexity, DENV serotypes interact differently with HLA [30]. The regions identified in this systematic review, including DRB1, DQA1, DQB1, A, B, and C, are among the most diverse regions of the HLA region [31]. A recent study examined some of these regions by supertype, and found the B44 supertype could be protective against DHF during secondary infections and that the A02 and A01/03 superotypes could be associated with more severe disease [32].

KIR genes, which are expressed on the surface of natural killer cells, also have wide genetic variability as noted with HLA genes [33]. While several KIR alleles were studied in DENV-infected populations, only one publication to date has examined KIR genotypes in West Nile virus-infected individuals, and this study had a sample size of four [34]. The results suggested a possible association; this, in conjunction with the results of the DENV research in this area and the genes' highly

Table 3 Genetic variation associated with DENV disease. We include in this table all variants studied by two or more research groups and variants found to have a significant association by one research group

Gene	Genetic Variation	Entrez Gene ID [31]	Major Allele	Minor Allele	Number of Cases	Number in Comparison Group	Country	Key Results	Meta-Analyzed
BAK1	rs5745568	578	G	T	509 DHF/DSS	409 DF	Thailand	1.32 (1.09–1.60) odds of severe disease associated with G allele [67]	No
CCR5	rs333	1234	–	Δ32 deletion	56 DF	91 controls	Australia	No significant association with disease [68]	Yes
CD209/ DCSIGN	rs480803	30,835	A	G	88 DHF/DSS	335 controls	Brazil	No significant association with disease [69]	
			A	G	509 DHF/DSS	409 DF	Thailand	No significant association with disease [67]	Yes
			A	G	88 DHF/DSS	335 controls	Brazil	No significant association with disease [69]	
			A	G	112 symptomatic	104 controls	India	No significant association with disease [70]	
			A	G	103 symptomatic	145 asymptomatic infections	Mexico	No significant association with disease [71]	
			A	G	156 DF and 12 DHF	72 controls	Brazil	No significant association with disease [72]	
			A	G	606 symptomatic	696 controls	Thailand	5.84 (2.77–12.31) odds of DHF compared to DF and 0.204 ($P = 2.0 \times 10^{-6}$) odds of symptomatic infection compared to controls [73]	
			A	G	286 symptomatic	236 asymptomatic infections	Brazil	No significant association with disease [74]	
			A	G	176 DF and 135 DH0046	120 controls	Taiwan	2.36 (1.12–4.97) odds of symptomatic infection compared to controls, 3.68 (1.67–8.09) odds of DHF compared to controls, and 2.46 (1.32–4.59) odds of DHF compared to DF [75]	
CFH	rs3753394	3075	C	T	187 DHF	121 DF	Thailand	No significant association with disease [76]	No ^a
			C	T	87 DHF	34 DF	Brazil	2.53 (1.38–4.69) odds of severe disease compared to mild disease under dominant model [77]	
CLECSA	rs1285933	23,601	T	C	88 DHF/DSS	335 controls	Brazil	2.25 (1.07–4.87) odds of severe disease for TT compared to CC genotype [69]	No
CXCL8/ IL8	rs4973	3576	T	A	45 DHF	108 controls	India	0.43 (0.20–0.93) odds of severe disease [78]	No
DDX58	rs3205166	23,586	T	G	120 DENV positive	109 controls	India	0.66 odds of disease associated with G allele for rs3205166 [79]	No
	rs11795343								
	rs669260								
FCγRII-a	rs1801274	2212	A (H amino acid)	G (R amino acid)	103 symptomatic	145 asymptomatic infections	Mexico	0.51 (0.26–0.98) odds of symptomatic infection and 0.45 (0.21–0.96) odds of severe disease compared to controls [71]	Yes
			T (H)	C (R)	89 DF and 33 DHF	107 controls	India	No significant association with disease [80]	
			T (H)	C (R)	68 DF, 29 DHF/DSS	42 asymptomatic infections	Cuba	10.56 (2.33–54.64) odds of DHF compared to asymptomatic disease for under dominant model [81]	

Table 3 Genetic variation associated with DENV disease. We include in this table all variants studied by two or more research groups and variants found to have a significant association by one research group (Continued)

Gene	Genetic Variation	Entrez Gene ID [31]	Major Allele	Minor Allele	Number of Cases	Number in Comparison Group	Country	Key Results	Meta-Analyzed
HPA	HPA 1a/1a	Not available	1a antigen	C (R)	302 DHF	238 controls	Vietnam	No significant association with disease [82]	No
			1b antigen	C (R)	40 DF, 30 DHF/DSS	40 asymptomatic infections	Pakistan	3.21 (1.29–7.97) odds of symptomatic disease, 2.82 (1.00–7.97) odds of DF, and 3.90 (1.13–13.07) odds of DHF/DSS over asymptomatic infection [83]	No
IFN-γ	rs2430561	3458	2a antigen	1b antigen	75 DHF	90 DF	India	1.93 odds ($p = 0.006$) of severe disease [84]	No
			2a antigen	2b antigen	75 DHF	90 DF	India	2.8 odds ($p = 0.007$) of severe disease [84]	No
IL-1B	rs16944	3553	A	T	80 symptomatic	100 DEN-negative febrile cases and 99 healthy controls	Brazil	2.23 ($p = 0.0255$) odds compared to DEN-negative and 2.37 ($p = 0.0165$) odds compared to controls [85]	Yes
			A	T	25 DHF	41 DF	Venezuela	No significant association with disease [86]	No
IL-1B	rs1143627	3553	A	T	43 DHF	99 controls	Cuba	No significant association with disease [87]	Yes
			A	G	45 DHF	108 controls	India	No significant association with disease [78]	Yes
IL-1B	rs1143627	3553	C	T	118 symptomatic	80 controls	Brazil	No significant association with disease [88]	No
			C	T	367 secondary DHF and 74 secondary DSS	313 secondary DF	Thailand	3.49 (1.36–8.95) odds of DSS compared to DHF and 2.81 (1.12–7.06) odds of DSS compared to DF under dominant models [89]	No
IL-1RA	86 base pair tandem repeat	3557	4 repeats	2 repeats	367 secondary DHF and 74 secondary DSS	313 secondary DF	Thailand	1.86 (1.05–3.26) odds of DSS compared to DHF and 1.86 (1.05–3.27) odds of DSS compared to DF for the 2/4 genotype [89]	No ^a
IL-6	rs1800795	3569	1 repeat	2–4 repeats	280 DHF	229 controls	Vietnam	No significant association between IL-1RA repeats and DHF [82]	Yes
			G	C	25 DHF	41 DF	Venezuela	No significant association with disease [86]	Yes
IL-6	rs1800795	3569	G	C	43 DHF	99 controls	Cuba	No significant association with disease [87]	Yes
			G	C	118 symptomatic	80 controls	Brazil	No significant association with disease [88]	Yes
IL-10	rs1800871	3586	G	C	200 DF	309 controls	Brazil	0.62 (0.42–0.91) odds of disease among heterozygotes compared to homozygote wild-type [90]	Yes
			C	T	88 DSS	335 controls	Brazil	No significant association with disease [69]	Yes
IL-10	rs1800871	3586	A	G	45 DHF	108 controls	India	No significant association with disease [78]	Yes
			C	T	25 DHF	41 DF	Venezuela	No significant association with disease [86]	Yes
IL-10	rs1800871	3586	C	T	43 DHF	99 controls	Cuba	No significant association with disease [87]	Yes
			C	T	200 DF	309 controls	Brazil	No significant association with disease [90]	Yes
IL-10	rs1800871	3586	T	C	86 DF, 182 DHF, 14 DSS	120 controls	Malaysia	No significant association with disease [91]	Yes

Table 3 Genetic variation associated with DENV disease. We include in this table all variants studied by two or more research groups and variants found to have a significant association by one research group (Continued)

Gene	Genetic Variation	Entrez Gene ID [31]	Major Allele	Minor Allele	Number of Cases	Number in Comparison Group	Country	Key Results	Meta-Analyzed
JAK1	rs1800872		C	T	107 DHF	62 controls	Sri Lanka	No significant association with disease [92]	Yes
			C	A	25 DHF	41 DF	Venezuela	No significant association with disease [86]	Yes
			C	A	43 DHF	99 controls	Cuba	No significant association with disease [87]	
			C	A	200 DF	309 controls	Brazil	No significant association with disease [90]	
			A	C	86 DF, 182 DHF, 14 DSS	120 controls	Malaysia	No significant association with disease [91]	
			A	C	107 DHF	62 controls	Sri Lanka	No significant association with disease [92]	Yes
			A	G	25 DHF	41 DF	Venezuela	No significant association with disease [86]	
			A	G	43 DHF	99 controls	Cuba	No significant association with disease [87]	
			A	G	200 DF	309 controls	Brazil	No significant association with disease [90]	
			A	G	86 DF, 182 DHF, 14 DSS	120 controls	Malaysia	No significant association with disease [91]	
MBL2	rs11208534	3716	A	G	107 DHF	62 controls	Sri Lanka	No significant association with disease [92]	No
			T	C	50 DHF	236 DF	Brazil	4.20 (1.7–10.4) odds of severe disease [74]	No
			G	A	50 DHF	236 DF	Brazil	2.1 (1.1–4.1) odds of severe disease [74]	No
			T	G	50 DHF	236 DF	Brazil	0.4 (0.2–0.7) odds of severe disease [74]	No
MICB	rs310196	4153	A	O	110 symptomatic	150 controls	Brazil	No significant association with disease [93]	Yes
			A	O	57 DHF	104 DF	Brazil	7.24 (1.38–38.02) odds of DHF among OO genotype compared to AA genotype [94]	
PLCE1	rs3740360	51,196	T	C	76 DSS	409 DF, 432 DHF	Thailand	1.58 (1.02–2.40) odds of DSS compared to non-DSS [95]	Yes
			T	C	2008 DSS	2018 controls	Vietnam	1.34 (1.23–1.46) odds of DSS per allele [96]	
			T	C	3961 cases	1068 controls	Vietnam	1.42 (1.20–1.64) odds of DSS per allele [97]	
			A	C	2008 DSS	2018 controls	Vietnam	0.80 (0.75–0.86) odds per allele of DSS [96]	Yes
			A	C	3961 cases	1068 controls	Vietnam	0.77 (0.59–0.99) odds per allele of DSS [97]	
			C	T	76 DSS	409 DF, 432 DHF	Thailand	1.49 (1.00–2.26) odds of DSS compared to non-DSS [95]	Yes
			C	T	2008 DSS	2018 controls	Vietnam	0.80 (0.75–0.86) odds per allele of DSS [96]	
			G	A	60 DHF	137 asymptomatic infections and controls	Cuba	0.36 (0.17–0.77) odds of severe disease [98]	No
			A	C	60 DHF	137 asymptomatic infections and controls	Cuba	0.44 (0.25–0.77) odds of severe disease [98]	
			A	G	60 DHF	137 asymptomatic infections and controls	Cuba	0.41 (0.24–0.72) odds of severe disease [98]	
RXRA	rs4262378	6256	A	G	60 DHF	137 asymptomatic infections and controls	Cuba	0.43 [0.24–0.76] odds of severe disease [98]	
			A	G	60 DHF	137 asymptomatic infections and controls	Cuba	0.43 [0.24–0.76] odds of severe disease [98]	

Table 3 Genetic variation associated with DENV disease. We include in this table all variants studied by two or more research groups and variants found to have a significant association by one research group (Continued)

Gene	Genetic Variation	Entrez Gene ID [31]	Major Allele	Minor Allele	Number of Cases	Number in Comparison Group	Country	Key Results	Meta-Analyzed
	rs62576287		C	A	60 DHF	137 asymptomatic infections and controls	Cuba	0.10 (0.01–0.83) odds of severe disease [98]	
TAP1	amino acid 333	6890	Ile	Val	90 DF, 75 DHF, 32 DSS	100 controls	India	2.58 (p = 0.007) odds of DHF among heterozygotes compared to DF [84]	Yes
TAP2	amino acid 379	6891	Ile	Val	107 DHF	62 controls	Sri Lanka	No significant association with disease [92]	Yes
			Val	Ile	107 DHF	62 controls	Sri Lanka	No significant association with disease [92]	
TGFB1	rs1800471	7040	Val	Ile	90 DF, 75 DHF, 32 DSS	100 controls	India	2.11 (p = 0.001) odds of DHF among heterozygotes [99]	Yes
			G	C	25 DHF	41 DF	Venezuela	No significant association with disease [86]	
TIRAP	rs8177374	114,609	G	C	200 DF	309 controls	Brazil	No significant association with disease [90]	No
			T	C	25 DHF	41 DF	Venezuela	No significant association with disease [86]	
			T	C	43 DHF	99 controls	Cuba	No significant association with disease [87]	
			T	C	200 DF	309 controls	Brazil	No significant association with disease [90]	
TLR3	rs3775291	7098	C	T	33 DHF	109 controls	India	2.64 (1.17–5.99) odds of severe disease among heterozygotes [100]	No
TNF-α	rs361525	7124	C	T	86 DF, 182 DHF, 14 DSS	120 controls	India	0.39 (0.16–0.88) odds of severe disease associated with T allele [100]	No
			G	A	41 DF, 32 DHF	169 controls	Malaysia	4.92 (1.10–21.90) odds of DHF compared to control group for heterozygotes [91]	Yes
TLR4	rs1800629	7099	G	A	80 symptomatic	100 DEN-negative febrile cases and 99 healthy controls	Mexico	0.19 (0.02–0.78) odds of disease with A allele [101]	Yes
			G	A	41 DF	41 DF	Brazil	No significant association with disease [85]	
			G	A	25 DHF	99 controls	Venezuela	2.5 (1.47–4.13) odds of severe disease [86]	
			G	A	43 DHF	99 controls	Cuba	3.51 (1.77–7.00) odds of severe disease [87]	
TLR4	amino acids 299 and 399	7099	G	A	200 DF	309 controls	Brazil	No significant association with disease [90]	Yes
			G	A	86 DF, 182 DHF, 14 DSS	120 controls	Malaysia	0.43 (0.22–0.84) odds of DHF compared to control for heterozygotes [91]	
			G	A	107 DHF	62 controls	Sri Lanka	2.53 (1.10–5.83) odds of disease for GG genotype [92]	
			G	A	85 DF, 29 DHF	110 controls	India	No significant association with disease [102]	
TLR4	amino acids 299 and 399	7099	G	A	19 DF, 82 DHF	106 controls	Thailand	No significant association with disease [103]	Yes
			G	A	85 DF, 45 DHF	163 controls	Mexico	No significant association with disease [101]	
TLR4	amino acids 299 and 399	7099	Asp299, Thr399	Gly299, Ile399	201 DHF	179 controls	Indonesia	No significant association with disease [104]	Yes

Table 3 Genetic variation associated with DENV disease. We include in this table all variants studied by two or more research groups and variants found to have a significant association by one research group (*Continued*)

Gene	Genetic Variation	Entrez Gene ID [31]	Major Allele	Minor Allele	Number of Cases	Number in Comparison Group	Country	Key Results	Meta-Analyzed
			Asp299, Thr399	Gly299, Ile399	63 DF, 57 DHF/DSS	200 controls	India	2.00 (1.17–3.43) odds associated with Gly299 for DF versus controls, and 2.38 (1.16–4.85) associated with Ile399 for DF versus controls [105]	
VDR	rs731236	7421	T	C	302 DHF	238 controls	Vietnam	Associated with more severe disease ($p = 0.033$) [82]	Yes
			T	C	83 DF, 29 DHF	105 controls	India	No significant association with disease [106]	

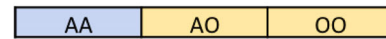
^a genotype data not available for meta-analysis

a) MBL2 - dominant model

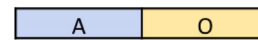
Study or Subgroup	Symptomatic		Asymptomatic + Control		Total Weight	Odds Ratio	
	Events	Total	Events	Total		M-H, Random, 95% CI	M-H, Random, 95% CI
Acioili-Santos 2008	28	57	34	104	37.8%	1.99	[1.03, 3.85]
Figueiredo 2016	42	110	48	150	62.2%	1.31	[0.78, 2.20]
Total (95% CI)	167	254	100.0%	1.54	[1.02, 2.31]		

MBL2 - allelic model

Study or Subgroup	Symptomatic		Asymptomatic + Control		Total Weight	Odds Ratio	
	Events	Total	Events	Total		M-H, Random, 95% CI	M-H, Random, 95% CI
Acioili-Santos 2008	49	200	55	200	60.5%	1.45	[0.94, 2.23]
Figueiredo 2016	34	114	36	208	39.5%	2.03	[1.19, 3.48]
Total (95% CI)	83	314	91	508	100.0%	1.65	[1.18, 2.32]



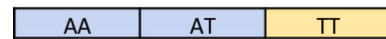
where O = alleles other than A



where O = alleles other than A

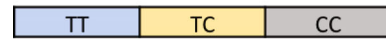
b) IFN-γ - recessive model

Study or Subgroup	Symptomatic		Asymptomatic + Control		Total Weight	Odds Ratio	
	Events	Total	Events	Total		M-H, Fixed, 95% CI	M-H, Fixed, 95% CI
Felton 2016	14	80	5	99	31.5%	3.99	[1.37, 11.61]
Perez 2010	13	43	18	92	68.5%	1.78	[0.78, 4.09]
Total (95% CI)	27	123	23	191	100.0%	2.48	[1.30, 4.71]



c) MICB - heterozygote vs homozygote wild

Study or Subgroup	Symptomatic		Asymptomatic + Control		Total Weight	Odds Ratio	
	Events	Total	Events	Total		M-H, Random, 95% CI	M-H, Random, 95% CI
Dang 2014	125	176	215	821	18.1%	0.63	[0.35, 1.13]
Khor 2011	594	1953	471	1986	45.3%	1.41	[1.22, 1.62]
Whitehorn 2013	81	284	1411	5876	36.6%	1.26	[0.97, 1.64]
Total (95% CI)	690	2320	2095	8683	100.0%	1.17	[0.86, 1.59]



MICB - homozygote mutant vs homozygote wild

Study or Subgroup	Symptomatic		Asymptomatic + Control		Total Weight	Odds Ratio	
	Events	Total	Events	Total		M-H, Random, 95% CI	M-H, Random, 95% CI
Dang 2014	5	83	19	627	11.0%	2.35	[0.85, 6.50]
Khor 2011	56	1415	31	1546	57.3%	2.01	[1.29, 3.14]
Whitehorn 2013	13	216	92	4557	31.7%	3.11	[1.71, 5.65]
Total (95% CI)	74	1704	142	6730	100.0%	2.35	[1.68, 3.29]



MICB - allelic model

Study or Subgroup	Symptomatic		Asymptomatic + Control		Total Weight	Odds Ratio	
	Events	Total	Events	Total		M-H, Random, 95% CI	M-H, Random, 95% CI
Dang 2014	125	176	215	1680	10.2%	0.94	[0.60, 1.47]
Khor 2011	706	4018	533	4034	57.9%	1.40	[1.24, 1.58]
Whitehorn 2013	107	594	1595	13936	31.9%	1.42	[1.15, 1.77]
Total (95% CI)	4788	17650	100.0%	1.35	[1.16, 1.57]		



d) PLCE1 rs3740360 - heterozyte vs homozygote wild

Study or Subgroup	Symptomatic		Asymptomatic + Control		Total Weight	Odds Ratio	
	Events	Total	Events	Total		M-H, Random, 95% CI	M-H, Random, 95% CI
Khor 2011	683	1910	823	1883	78.2%	0.72	[0.61, 0.82]
Whitehorn 2013	105	281	2414	5569	21.8%	0.78	[0.61, 1.00]
Total (95% CI)	2191	7452	100.0%	0.73	[0.65, 0.82]		



PLCE1 rs3740360 - homozygote mutant vs homozygote wild

Study or Subgroup	Symptomatic		Asymptomatic + Control		Total Weight	Odds Ratio	
	Events	Total	Events	Total		M-H, Random, 95% CI	M-H, Random, 95% CI
Khor 2011	99	1326	136	1196	81.6%	0.63	[0.48, 0.83]
Whitehorn 2013	13	189	418	3573	18.4%	0.56	[0.31, 0.99]
Total (95% CI)	112	1515	554	4769	100.0%	0.62	[0.48, 0.79]



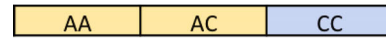
PLCE1 rs3740360 - allelic model

Study or Subgroup	Symptomatic		Asymptomatic + Control		Total Weight	Odds Ratio	
	Events	Total	Events	Total		M-H, Random, 95% CI	M-H, Random, 95% CI
Khor 2011	881	4018	1095	2943	51.0%	0.47	[0.43, 0.53]
Whitehorn 2013	131	588	3250	11974	48.4%	0.77	[0.63, 0.94]
Total (95% CI)	1012	4606	4345	14917	100.0%	0.60	[0.37, 0.96]



e) PLCE1 rs3765524 - recessive model

Study or Subgroup	Symptomatic		Asymptomatic + Control		Total Weight	Odds Ratio	
	Events	Total	Events	Total		M-H, Random, 95% CI	M-H, Random, 95% CI
Dang 2014	6	76	93	839	9.7%	0.69	[0.29, 1.63]
Khor 2011	125	2009	93	839	90.3%	0.53	[0.40, 0.71]
Total (95% CI)	131	2085	186	1678	100.0%	0.55	[0.42, 0.71]



PLCE1 rs3765524 - allelic model

Study or Subgroup	Symptomatic		Asymptomatic + Control		Total Weight	Odds Ratio	
	Events	Total	Events	Total		M-H, Random, 95% CI	M-H, Random, 95% CI
Dang 2014	36	152	530	1678	6.0%	0.67	[0.46, 0.99]
Khor 2011	1002	4018	1213	4038	94.0%	0.77	[0.70, 0.85]
Total (95% CI)	1038	4170	1743	5716	100.0%	0.77	[0.70, 0.84]

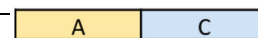


Fig. 3 (See legend on next page.)

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Fig. 3 Meta-analyzed genetic variation associated with DENV disease. Genotype count data from published reports of WNV subjects were meta-analyzed using RevMan. The meta-odds ratio (OR) for more severe disease is indicated with the genetic model for each gene: MBL2 (**a**), IFN- γ (**b**), MICB (**c**), PLCE1 (**d** and **e**). If multiple models were significant, we present the most significant model. The alleles or genotypes associated with asymptomatic infection and controls (blue) or with severe disease (yellow) outcome are shown for each gene

polymorphic nature, could be an area that should be explored further. Infection with WNV has been shown to lead to diversification of KIR receptor expression [35]. In addition to the research outlined above, researchers have examined the association of KIR genotypes with DENV infection in vitro. Within the in vitro research, the timing of natural killer cell activation has been linked to disease severity and interactions between KIR and HLA have been suggested [36–38].

Another key non-HLA gene identified to be associated with WNV disease was OASL, which codes for an enzyme that is induced by type 1 interferon and viruses [39]. OASL was first identified to have a potentially critical role in WNV disease pathogenesis in 2002, when researchers found that mice with a truncated form of the gene were more susceptible to disease [40]. Elevated activity of the OAS genes has also been associated with more severe DENV infection in vitro [41]. This data and the significance of variation within the OAS genes for WNV outcomes highlight the importance of the interferon pathways in response to flavivirus infections and suggest a need for further in depth examination the association of genetic variability within OAS and DENV severity.

CCR5 Δ 32 was the only gene studied by two or more research groups for each disease. CCR5 was first identified as a co-receptor for HIV in 1996, and CCR5 deficiency, or a homozygous genotype of CCR5 Δ 32, was found to be protective against HIV infection [42–45]. In West Nile, CCR5 deficiency is not associated with incidence of infection, but is associated with severity of disease for infected individuals [46]. Subsequent research showed that CCR5 specifically plays a role in the ability of cortical neurons to combat West Nile virus infection of the brain [47]. In DENV, CCR5 deficiency has been linked with increased viral load and disease severity [48]. The study also found that the CCR5 receptor in macrophages is necessary for replication of DENV serotype 2, an early step in the infection process [49]. Given the similarities of these flaviviral diseases and the significant association of CCR5, the only gene looked at by research groups for both diseases, further research could be beneficial to further understanding the role of genetic variation in the development of severe flaviviral disease [50].

When we meta-analyzed the DENV studies, we found significant associations between DENV disease and genetic variation in MBL2, PLCE1, IFN- γ , and MICB. The role of many of these genes in disease pathogenesis has

been characterized through in vivo and in vitro studies. MBL2, the mannose-binding lectin 2 gene, encodes a protein with a role in innate immunity and complement pathway, while PLCE1 encodes an enzyme critical to the generation of the inositol 1,4,5-triphosphate (IP3) and diacylglycerol (DAG) messengers [51]. MICB and IFN- γ are both critical in the immune response, and thus variations within these genes could have strong effects on the initial response to the viral infection and the subsequent disease pathogenesis [51].

Our study is limited by several factors, most notably by the available literature. To ensure we found as many papers as possible, we constructed a search strategy that involved multiple databases, used both subject headings and text words, was not limited to English articles, and included an ancestry search [52]. Despite our focus on significant results and genes studied by at least two research groups, the wide heterogeneity among the populations studied limited our ability to interpret the meta-analyzed results. Lack of diversity in genetic studies is well-documented [53], and the absence of certain affected populations, particularly in Africa, among the identified studies further demonstrates this unmet research need [54]. The diversity of results among studies that examined the same SNP could be due to population heterogeneity, as well as to differences in study approach, including selection of control and comparison groups. Additionally, previous exposure history, DENV serotype, and WNV or DENV genotype are all factors that can affect disease severity, but were not accounted for in the included studies [4]. The number and type of genes examined varied greatly between studies, and we were limited by what genes researchers chose to sequence and include in publications. The unavailability of comparable genotype data and the incomparability of research groups across some studies preclude a more in depth analysis at present.

Conclusions

The genes found to be significantly associated with WNV or DENV disease pathogenesis varied in function, with most being linked to the immune response. As the regions of the world affected by WNV, DENV, and related viruses such as Zika, continue to expand due in part to climate change, an improved understanding of the association between genetic variation and disease severity will be valuable for all potentially affected populations [55–58]. Based on the growing incidence of these

diseases, the paucity of consistency in the associations found, and the limited overlap in genetic targets studied to date, there is need for continued and deeper studies examining the role of genetic factors in WNV and DENV disease severity. In addition to conducting new studies such as whole-exome sequencing within larger population samples, further analyses could be conducted of existing data, to glean novel findings such as gene-gene or gene-environment interactions, rare and low frequency variants, and pathways of significant determinants of anti-viral resistance.

Additional files

Additional file 1: PRISMA Checklist. This systematic review of genetic factors and WNV or dengue disease was conducted using the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) guidelines outlined in this checklist. (DOC 95 kb)

Additional file 2: Sample Search Strategy for the Embase Database. Medline, PubMed, Embase, and Global Health databases were used to search the literature. Search terms included West Nile or Dengue and genetic factors; the same set of text words was used for all databases in conjunction with subject headings that were tailored for each database. As an example, this search strategy for the Embase database is provided. These subject headings, in conjunction with the text search (Table 1), were used to find relevant literature in Embase. (DOCX 13 kb)

Additional file 3: Included Papers. All publications included in this review are listed, with study details on authors, year of publication, country, sample size, case and control group definitions, and genotyping method. (XLSX 54 kb)

Additional file 4: HLA Associations with DENV Disease Severity. HLA alleles studied by two or more research groups for association with DENV disease severity. (DOCX 311 kb)

Additional file 5: HLA Associations with WNV Disease Severity. All examined targets from the analyzed publications are listed, with significant associations shown in bold. (XLSX 9 kb)

Additional file 6: Quality Scores. The quality of each study was evaluated with the Newcastle-Ottawa Quality Assessment Scale for Case-Control Studies, which assesses each study's selection, comparability, and exposure ascertainment approach. (XLSX 16 kb)

Additional file 7: Extracted Data from Included Studies. All genotype data extracted from the manuscripts or collected from study authors is provided. (XLSX 54 kb)

Abbreviations

DENV: Dengue virus; DHF: Dengue Hemorrhagic Fever; DSS: Dengue Shock Syndrome; GWAS: genome-wide association studies; PRISMA: Preferred Reporting Items for Systematic Reviews and Meta-Analyses; SNP: single nucleotide polymorphism; WNV: West Nile virus

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Availability of data and materials

All data generated or analyzed during this study are included in this published article and its supplementary information files.

Authors' contributions

Conceptualization (MEC, ATD, RRM); project administration (MEC, RRM); data curation (MEC, SC); methodology (MEC, SC, ATD); formal analysis (MEC); writing – original draft preparation (MEC); writing – review and editing (MEC, SC, ATD, RRM). All authors read and approved the final manuscript.

Ethics approval and consent to participate

Not applicable. Data sets were extracted without personal identifiers and organized into literature tables.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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References

- Papa A, Karabaxoglou D, Kansouzidou A. Acute West Nile virus neuroinvasive infections: cross-reactivity with dengue virus and tick-borne encephalitis virus. *J Med Virol*. 2011;83(10):1861–5.
- Hua RH, Chen NS, Qin CF, Deng YQ, Ge JY, Wang XJ, Qiao ZJ, Chen WY, Wen ZY, Liu WX, et al. Identification and characterization of a virus-specific continuous B-cell epitope on the PrM/M protein of Japanese encephalitis virus: potential application in the detection of antibodies to distinguish Japanese encephalitis virus infection from West Nile virus and dengue virus infections. *Virology*. 2010;7:249.
- Daep CA, Munoz-Jordan JL, Eugenin EA. Flaviviruses, an expanding threat in public health: focus on dengue, West Nile, and Japanese encephalitis virus. *J Neuro-Oncol*. 2014;20(6):539–60.
- Gubler DJ. The continuing spread of West Nile virus in the western hemisphere. *Clin Infect Dis*. 2007;45(8):1039–46.
- Petersen LR, Brault AC, Nasci RS. West Nile virus: review of the literature. *JAMA*. 2013;310(3):308–15.
- Symptoms & Treatments [<http://www.cdc.gov/westnile/symptoms/index.html>].
- Coffey LL, Mertens E, Brehin AC, Fernandez-Garcia MD, Amara A, Despres P, Sakuntabhai A. Human genetic determinants of dengue virus susceptibility. *Microbes Infect*. 2009;11(2):143–56.
- Kalayanarooj S. Clinical manifestations and Management of Dengue/DHF/ DSS. *Trop Med Health*. 2011;39(4 Suppl):83–7.
- Montgomery RR, Murray KO. Risk factors for West Nile virus infection and disease in populations and individuals. *Expert Rev Anti-Infect Ther*. 2015; 13(3):317–25.
- Whitehorn J, Hubbard S, Anders KL, Quyen NTH, Simmons C. Dengue pathogenesis: host factors. *Dengue and Dengue Hemorrhagic Fever*, 2nd Edition. 2014:214–28.
- James EA, Gates TJ, LaFond RE, Yamamoto S, Ni C, Mai D, Gersuk VH, O'Brien K, Nguyen QA, Zeitner B, et al. Neuroinvasive West Nile infection elicits elevated and atypically polarized T cell responses that promote a pathogenic outcome. *PLoS Pathog*. 2016;12(1):e1005375.
- Qian F, Thakar J, Yuan X, Nolan M, Murray KO, Lee WT, Wong SJ, Meng H, Fikrig E, Kleinstein SH, et al. Immune markers associated with host susceptibility to infection with West Nile virus. *Viral Immunol*. 2014;27(2):39–47.
- Yao Y, Montgomery RR. Role of immune aging in susceptibility to West Nile virus. *Methods Mol Biol*. 2016;1435:235–47.

14. Cahill ME, Yao Y, Nock D, Armstrong PM, Andreadis TG, Diuk-Wasser MA, Montgomery RR. West Nile Virus Seroprevalence, Connecticut, USA, 2000–2014. *Emerg Infect Dis*. 2017;23(4):708–10.
15. Krishnan MN, Sukumaran B, Pal U, Agaisse H, Murray JL, Hodge TW, Fikrig E. Rab 5 is required for the cellular entry of dengue and West Nile viruses. *J Virol*. 2007;81(9):4881–5.
16. Campos RK, Wong B, Xie XP, Lu YF, Shi PY, Pompon J, Garcia-Blanco MA, Bradrick SS. RPLP1 and RPLP2 are essential flavivirus host factors that promote early viral protein accumulation. *J Virol*. 2017;91(4)
17. Gack MU, Diamond MS. Innate immune escape by dengue and West Nile viruses. *Curr Opin Virol*. 2016;20:119–28.
18. Chaturvedi UC, Agarwal R, Elbishbishi EA, Mustafa AS. Cytokine cascade in dengue hemorrhagic fever: implications for pathogenesis. *FEMS Immunol Med Microbiol*. 2000;28(3):183–8.
19. Kumar M, Verma S, Nerurkar VR. Pro-inflammatory cytokines derived from West Nile virus (WNV)-infected SK-N-SH cells mediate neuroinflammatory markers and neuronal death. *J Neuroinflammation*. 2010;7:73.
20. Rossini G, Landini MP, Gelsomino F, Sambri V, Varani S. Innate host responses to West Nile virus: implications for central nervous system immunopathology. *World J Virol*. 2013;2(2):49–56.
21. Sprague AH, Khalil RA. Inflammatory cytokines in vascular dysfunction and vascular disease. *Biochem Pharmacol*. 2009;78(6):539–52.
22. Mackenzie JS, Gubler DJ, Petersen LR. Emerging flaviviruses: the spread and resurgence of Japanese encephalitis, West Nile and dengue viruses. *Nat Med*. 2004;10(12):S98–S109.
23. Hotez PJ, Murray KO. Dengue, West Nile virus, chikungunya, Zika - and now Mayaro? *PLoS Negl Trop Dis*. 2017;11(8): e0005462.
24. Yun SJ, Lee YM. Zika virus: An emerging flavivirus. *J Microbiol*. 2017;55(3):204–19.
25. Moher D, Liberati A, Tetzlaff J, Altman DG, Group P. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. *PLoS Med*. 2009;6(7):e1000097.
26. Covidence systematic review software. www.covidence.org.
27. Tomas J, Aragon MPF, Daniel Wollschlaeger, Adam Omidpanah: epitools: Epidemiology Tools. In.; 2017.
28. Review Manager (RevMan). In., 5.3 EDN Copenhagen: The Nordic Cochrane Centre, The Cochrane Collaboration; 2014.
29. Wells G, Shea B, O'Connell D, Peterson J, Welch V, Losos M, Tugwell P: The Newcastle-Ottawa Scale (NOS) for assessing the quality of nonrandomised studies in meta-analyses. In.; 2000.
30. Gan CS, Yusof R, Othman S. Different serotypes of dengue viruses differently regulate the expression of the host cell antigen processing machinery. *Acta Trop*. 2015;149:8–14.
31. Shiina T, Hosomichi K, Inoko H, Kulski JK. The HLA genomic loci map: expression, interaction, diversity and disease. *J Hum Genet*. 2009;54(1):15–39.
32. Vejbaeysa S, Thongpradit R, Kalayanaroj S, Luangtrakool K, Luangtrakool P, Gibbons RV, Srinak D, Ngammthaworn S, Apisawes K, Yoon IK, et al. HLA class I Supertype associations with clinical outcome of secondary dengue virus infections in ethnic Thais. *J Infect Dis*. 2015;212(6):939–47.
33. Uhrberg M. The KIR gene family: life in the fast lane of evolution. *Eur J Immunol*. 2005;35(1):10–5.
34. Spiroski M, Milenkovic Z, Petlichkovski A, Ivanovski L, Topuzovska IK, Djulejic E. Killer cell immunoglobulin-like receptor genes in four human West Nile virus infections reported 2011 in the republic of Macedonia. *Hum Immunol*. 2013;74(3):389–94.
35. Strauss-Albee DM, Fukuyama J, Liang EC, Yao Y, Jarrell JA, Drake AL, Kinuthia J, Montgomery RR, John-Stewart G, Holmes S, et al. Human NK cell repertoire diversity reflects immune experience and correlates with viral susceptibility. *Sci Transl Med*. 2015;7(297):297ra115.
36. Azeredo EL, De Oliveira-Pinto LM, Zagne SM, Cerqueira DI, Nogueira RM, Kubelka CF. NK cells, displaying early activation, cytotoxicity and adhesion molecules, are associated with mild dengue disease. *Clin Exp Immunol*. 2006;143(2):345–56.
37. Townsley E, O'Connor G, Cosgrove C, Woda M, Co M, Thomas SJ, Kalayanaroj S, Yoon IK, Nisalak A, Srikiatkachorn A, et al. Interaction of a dengue virus NS1-derived peptide with the inhibitory receptor KIR3DL1 on natural killer cells. *Clin Exp Immunol*. 2016;183(3):419–30.
38. Yao Y, Strauss-Albee DM, Zhou JQ, Malawista A, Garcia MN, Murray KO, Blish CA, Montgomery RR. The natural killer cell response to West Nile virus in young and old individuals with or without a prior history of infection. *PLoS One*. 2017;12(2):e0172625.
39. OASL - 2'-5'-oligoadenylate synthase-like protein. <http://www.uniprot.org/uniprot/Q15646>.
40. Mashimo T, Lucas M, Simon-Chazottes D, Frenkiel MP, Montagutelli X, Ceccaldi PE, Deubel V, Guenet JL, Despres P. A nonsense mutation in the gene encoding 2'-5'-oligoadenylate synthetase/L1 isoform is associated with West Nile virus susceptibility in laboratory mice. *Proc Natl Acad Sci U S A*. 2002;99(17):11311–6.
41. Simon-Lorieri E, Lin RJ, Kalayanaroj SM, Chuansumrit A, Casademont I, Lin SY, Yu HP, Lert-Itthiporn W, Chaiyaratana W, Tangthawornchaikul N, et al. High anti-dengue virus activity of the OAS gene family is associated with increased severity of dengue. *J Infect Dis*. 2015;212(12):2011–20.
42. Alkhatib G, Combadiere C, Broder CC, Feng Y, Kennedy PE, Murphy PM, Berger EA. CC CKR5: a RANTES, MIP-1alpha, MIP-1beta receptor as a fusion cofactor for macrophage-tropic HIV-1. *Science*. 1996;272(5270):1955–8.
43. Deng H, Liu R, Ellmeier W, Choe S, Unutmaz D, Burkhart M, Di Marzio P, Marmon S, Sutton RE, Hill CM, et al. Identification of a major co-receptor for primary isolates of HIV-1. *Nature*. 1996;381(6584):661–6.
44. Dragic T, Litwin V, Allaway GP, Martin SR, Huang Y, Nagashima KA, Cayanan C, Maddon PJ, Koup RA, Moore JP, et al. HIV-1 entry into CD4+ cells is mediated by the chemokine receptor CC-CKR-5. *Nature*. 1996;381(6584):667–73.
45. Liu R, Paxton WA, Choe S, Ceradini D, Martin SR, Horuk R, MacDonald ME, Stuhlmann H, Koup RA, Landau NR. Homozygous defect in HIV-1 coreceptor accounts for resistance of some multiply-exposed individuals to HIV-1 infection. *Cell*. 1996;86(3):367–77.
46. Lim JK, McDermott DH, Lisco A, Foster GA, Krysztof D, Follmann D, Stramer SL, Murphy PM. CCR5 deficiency is a risk factor for early clinical manifestations of West Nile virus infection but not for viral transmission. *J Infect Dis*. 2010;201(2):178–85.
47. Durrant DM, Daniels BP, Pasiaka T, Dorsey D, Klein RS. CCR5 limits cortical viral loads during West Nile virus infection of the central nervous system. *J Neuroinflammation*. 2015;12:233.
48. Marques RE, Guabiraba R, Del Sarto JL, Rocha RF, Queiroz AL, Cisalpino D, Marques PE, Pacca CC, Fagundes CT, Menezes GB, et al. Dengue virus requires the CC-chemokine receptor CCR5 for replication and infection development. *Immunology*. 2015;145(4):583–96.
49. Chen YC, Wang SY. Activation of terminally differentiated human monocytes/macrophages by dengue virus: productive infection, hierarchical production of innate cytokines and chemokines, and the synergistic effect of lipopolysaccharide. *J Virol*. 2002;76(19):9877–87.
50. Loeb M. Genetic susceptibility to West Nile virus and dengue. *Public Health Genomics*. 2013;16(1–2):4–8.
51. Gene. <https://www.ncbi.nlm.nih.gov/gene/>.
52. Akobeng AK. Understanding systematic reviews and meta-analysis. *Arch Dis Child*. 2005;90(8):845–8.
53. Popejoy AB, Fullerton SM. Genomics is failing on diversity. *Nature*. 2016;538(7624):161–4.
54. Were F. The dengue situation in Africa. *Paediatr Int Child Health*. 2012;32(Suppl 1):18–21.
55. Ebi KL, Nealon J. Dengue in a changing climate. *Environ Res*. 2016;151:115–23.
56. Paz S. Climate change impacts on West Nile virus transmission in a global context. *Philos Trans R Soc Lond Ser B Biol Sci*. 2015;370(1665)
57. Carlson CJ, Dougherty ER, Getz W. An ecological assessment of the pandemic threat of Zika virus. *PLoS Negl Trop Dis*. 2016;10(8):e0004968.
58. Lessler J, Chaisson LH, Kucirka LM, Bi Q, Grantz K, Salje H, Carcelen AC, Ott CT, Sheffield JS, Ferguson NM, et al. Assessing the global threat from Zika virus. *Science*. 2016;353(6300):aaf8160.
59. Loeb M, Eskandarian S, Rupp M, Fishman N, Gasink L, Patterson J, Bramson J, Hudson TJ, Lemire M. Genetic variants and susceptibility to neurological complications following West Nile virus infection. *J Infect Dis*. 2011;204(7):1031–7.
60. Danial-Farran N, Eghbaria S, Schwartz N, Kra-Oz Z, Bisharat N. Genetic variants associated with susceptibility of Ashkenazi Jews to West Nile virus infection. *Epidemiology & Infection*. 2015;143(4):857–63.
61. Long D, Deng X, Singh P, Loeb M, Lauring AS, Seielstad M. Identification of genetic variants associated with susceptibility to West Nile virus neuroinvasive disease. *Genes & Immunity*. 2016;17(5):298–304.
62. Bigham AW, Buckingham KJ, Husain S, Emond MJ, Bofferding KM, Gildersleeve H, Rutherford A, Astakhova NM, Perelygin AA, Busch MP et al: Host genetic risk factors for West Nile virus infection and disease progression. *PLoS One* 2011, 6 (9) (no pagination)(e24745).
63. Glass WG, McDermott DH, Lim JK, Lekhong S, Yu SF, Frank WA, Pape J, Cheshier RC, Murphy PM. CCR5 deficiency increases risk of symptomatic West Nile virus infection. *J Exp Med*. 2006;203(1):35–40.

64. Das R, Loughran K, Murchison C, Qian F, Leng L, Song Y, Montgomery RR, Loeb M, Bucala R. Association between high expression macrophage migration inhibitory factor (MIF) alleles and West Nile virus encephalitis. *Cytokine*. 2016;78:51–4.
65. Yakub I, Lillibridge KM, Moran A, Gonzalez OY, Belmont J, Gibbs RA, Tweardy DJ. Single nucleotide polymorphisms in genes for 2'-5'-oligoadenylate synthetase and RNase L in patients hospitalized with West Nile virus infection. *J Infect Dis*. 2005;192(10):1741–8.
66. Lim JK, Lisco A, McDermott DH, Huynh L, Ward JM, Johnson B, Johnson H, Pape J, Foster GA, Krysztof D, et al. Genetic variation in OAS1 is a risk factor for initial infection with West Nile virus in man. *PLoS Pathog*. 2009;5(2):e1000321.
67. Dang TN, Naka I, Sa-Ngasang A, Anantapreecha S, Wichukhinda N, Sawanpanyalert P, Patarapotikul J, Tsuchiya N, Ohashi J. Association of BAK1 single nucleotide polymorphism with a risk for dengue hemorrhagic fever. *BMC Medical Genetics*. 2016;17(1):43.
68. Brestovac B, Halicki LA, Harris RP, Sampson I, Speers DJ, Mamotte C, Williams D. Primary acute dengue and the deletion in chemokine receptor 5 (CCR5DELTA32). *Microbes & Infection*. 2014;16(6):518–21.
69. Xavier-Carvalho C, Gibson G, Brasil P, Ferreira RX, de Souza Santos R, Goncalves Cruz O, de Oliveira SA, de Sa Carvalho M, Pacheco AG, Kubelka CF, et al. Single nucleotide polymorphisms in candidate genes and dengue severity in children: a case-control, functional and meta-analysis study. *Infection, Genetics & Evolution*. 2013;20:197–205.
70. Alagarasu K, Damle IM, Bachal RV, Mulay AP, Shah PS, Dayaraj C. Association of promoter region polymorphisms of CD209 gene with clinical outcomes of dengue virus infection in western India. *Infection, Genetics & Evolution*. 2013;17:239–42.
71. Noecker CA, Amaya-Larios IY, Galeana-Hernandez M, Ramos-Castaneda J, Martinez-Vega RA. Contrasting associations of polymorphisms in FcgammaRIIIa and DC-SIGN with the clinical presentation of dengue infection in a Mexican population. *Acta Trop*. 2014;138:15–22.
72. Oliveira LF, Lima CP, Azevedo Rdo S, Mendonca DS, Rodrigues SG, Carvalho VL, Pinto EV, Maia AL, Maia MH, Vasconcelos JM, et al. Polymorphism of DC-SIGN (CD209) promoter in association with clinical symptoms of dengue fever. *Viral Immunol*. 2014;27(5):245–9.
73. Sakuntabhai A, Turbpaiboon C, Casademont I, Chuansumrit A, Lowhnoo T, Kajaste-Rudnitski A, Kalayanaraj SM, Tangnararatchakit K, Tangthawornchaikul N, Vasanawathana S, et al. A variant in the CD209 promoter is associated with severity of dengue disease. *Nat Genet*. 2005;37(5):507–13.
74. Silva LK, Blanton RE, Parrado AR, Melo PS, Morato VG, Reis EA, Dias JP, Castro JM, Vasconcelos PF, Goddard KA, et al. Dengue hemorrhagic fever is associated with polymorphisms in JAK1. *Eur J Hum Genet*. 2010;18(11):1221–7.
75. Wang L, Chen RF, Liu JW, Lee IK, Lee CP, Kuo HC, Huang SK, Yang KD. DC-SIGN (CD209) promoter -336 a/G polymorphism is associated with dengue hemorrhagic fever and correlated to DC-SIGN expression and immune augmentation. *PLoS Neglected Tropical Diseases* [electronic resource]. 2011; 5(1):e934.
76. Kraivong R, Vasanawathana S, Limpitkul W, Malasit P, Tangthawornchaikul N, Botto M, Sreanont GR, Mongkolsapaya J, Pickering MC. Complement alternative pathway genetic variation and dengue infection in the Thai population. *Clinical & Experimental Immunology*. 2013;174(2):326–34.
77. Pastor AF, Rodrigues Moura L, Neto JW, Nascimento EJ, Calzavara-Silva CE, Gomes AL, Silva AM, Cordeiro MT, Braga-Neto U, Crovella S, et al. Complement factor H gene (CFH) polymorphisms C-257T, G257A and haplotypes are associated with protection against severe dengue phenotype, possible related with high CFH expression. *Hum Immunol*. 2013;74(9):1225–30.
78. Alagarasu K, Bachal RV, Tillu H, Mulay AP, Kakade MB, Shah PS, Cecilia D. Association of combinations of interleukin-10 and pro-inflammatory cytokine gene polymorphisms with dengue hemorrhagic fever. *Cytokine*. 2015;74(1):130–6.
79. Alagarasu K, Memane RS, Shah PS. Polymorphisms in the retinoic acid-1 like-receptor family of genes and their association with clinical outcome of dengue virus infection. *Arch Virol*. 2015;160(6):1555–60.
80. Alagarasu K, Bachal RV, Damle I, Shah PS, Cecilia D. Association of FCGR2A p.R131H and CCL2 c.-2518 A>G gene variants with thrombocytopenia in patients with dengue virus infection. *Hum Immunol*. 2015;76(11):819–22.
81. Garcia G, Sierra B, Perez AB, Aguirre E, Rosado I, Gonzalez N, Izquierdo A, Pupo M, Danay Diaz DR, Sanchez L, et al. Asymptomatic dengue infection in a Cuban population confirms the protective role of the RR variant of the FcgammaRIIIa polymorphism. *American Journal of Tropical Medicine & Hygiene*. 2010;82(6):1153–6.
82. Loke H, Bethell D, Phuong CX, Day N, White N, Farrar J, Hill A. Susceptibility to dengue hemorrhagic fever in Vietnam: evidence of an association with variation in the vitamin d receptor and fc gamma receptor IIa genes. *American Journal of Tropical Medicine & Hygiene*. 2002;67(1):102–6.
83. Mohsin SN, Mahmood S, Amar A, Ghafoor F, Raza SM, Saleem M. Association of FcgammaRIIIa polymorphism with clinical outcome of dengue infection: first insight from Pakistan. *American Journal of Tropical Medicine & Hygiene*. 2015;93(4):691–6.
84. Soundravally R, Hoti SL. Immunopathogenesis of dengue hemorrhagic fever and shock syndrome: role of TAP and HPA gene polymorphism. *Hum Immunol*. 2007;68(12):973–9.
85. Feitosa RNM, Vallinoto ACR, Vasconcelos PFDC, Azevedo RDS, Azevedo VN, MacHado LFA, Lima SS, Ishak MDOG, Ishak R. Gene polymorphisms and serum levels of pro- and anti-inflammatory markers in dengue viral infections. *Viral Immunol*. 2016;29(7):379–88.
86. Fernandez-Mestre MT, Gendzekhadze K, Rivas-Vetencourt P, Layrisse Z. TNF-alpha-308A allele, a possible severity risk factor of hemorrhagic manifestation in dengue fever patients. *Tissue Antigens*. 2004;64(4):469–72.
87. Perez AB, Sierra B, Garcia G, Aguirre E, Babel N, Alvarez M, Sanchez L, Valdes L, Volk HD, Guzman MG. Tumor necrosis factor-alpha, transforming growth factor-beta1, and interleukin-10 gene polymorphisms: implication in protection or susceptibility to dengue hemorrhagic fever. *Hum Immunol*. 2010;71(11):1135–40.
88. Cansancoa IF, do Carmo AP, Leite RD, Portela RD, de Sa Leitao Paiva Junior S, de Queiroz Balbino V, Rabenhorst SH. Association of genetic polymorphisms of IL1beta -511 C>T, IL1RN VNTR 86 bp, IL6-174 G>C, IL10-819 C>T and TNFalpha -308 G>A, involved in symptomatic patients with dengue in Brazil. *Inflamm Res*. 2016;65(11):925–32.
89. Sa-Ngasang A, Ohashi J, Naka I, Anantapreecha S, Sawanpanyalert P, Patarapotikul J. Association of IL1B -31C/T and IL1RA variable number of an 86-bp tandem repeat with dengue shock syndrome in Thailand. *J Infect Dis*. 2014;210(1):138–45.
90. Moreira ST, Cardoso DM, Visentainer JE, Fonzar UJV, Moliterno RA. The possible protective role of the Il6^{-174GC} genotype in Dengue Fever. *Open Tropical Medicine Journal*. 2008;1:87–91.
91. Sam SS, Teoh BT, Chinna K, AbuBakar S. High producing tumor necrosis factor alpha gene alleles in protection against severe manifestations of dengue. *Int J Med Sci*. 2015;12(2):177–86.
92. Fernando AN, Malavige GN, Perera KL, Premawansa S, Ogg GS, De Silva AD. Polymorphisms of transporter associated with antigen presentation, tumor necrosis factor-alpha and Interleukin-10 and their implications for protection and susceptibility to severe forms of dengue fever in patients in Sri Lanka. *J Global Infect Dis*. 2015;7(4):157–64.
93. Acioli-Santos B, Segat L, Dhalia R, Brito CA, Braga-Neto UM, Marques ET, Crovella S. MBL2 gene polymorphisms protect against development of thrombocytopenia associated with severe dengue phenotype. *Hum Immunol*. 2008;69(2):122–8.
94. Figueiredo GG, Cezar RD, Freire NM, Teixeira VG, Baptista P, Cordeiro M, Carmo RF, Vasconcelos LR, Moura P. Mannose-binding lectin gene (MBL2) polymorphisms related to the mannose-binding lectin low levels are associated to dengue disease severity. *Hum Immunol*. 2016;77(7):571–5.
95. Dang TN, Naka I, Sa-Ngasang A, Anantapreecha S, Chanama S, Wichukhinda N, Sawanpanyalert P, Patarapotikul J, Tsuchiya N, Ohashi J. A replication study confirms the association of GWAS-identified SNPs at MICB and PLCE1 in Thai patients with dengue shock syndrome. *BMC Medical Genetics*. 2014;15:58.
96. Khor CC, Chau TN, Pang J, Davila S, Long HT, Ong RT, Dunstan SJ, Wills B, Farrar J, Van Tram T, et al. Genome-wide association study identifies susceptibility loci for dengue shock syndrome at MICB and PLCE1. *Nat Genet*. 2011;43(11):1139–41.
97. Whitehorn J, Chau TN, Nguyet NM, Kien DT, Quyen NT, Trung DT, Pang J, Wills B, Van Vinh Chau N, Farrar J, et al. Genetic variants of MICB and PLCE1 and associations with non-severe dengue. *PLoS ONE* [Electronic Resource]. 2013;8(3):e59067.
98. Sierra B, Triska P, Soares P, Garcia G, Perez AB, Aguirre E, Oliveira M, Cavadas B, Regnault B, Alvarez M, et al. OSBPL10, RXRA and lipid metabolism confer African-ancestry protection against dengue haemorrhagic fever in admixed Cubans. *PLoS Pathog*. 2017;13(2):e1006220.
99. Soundravally R, Hoti SL. Significance of transporter associated with antigen processing 2 (TAP2) gene polymorphisms in susceptibility to dengue viral infection. *J Clin Immunol*. 2008;28(3):256–62.

100. Alagarasu K, Bachal RV, Memane RS, Shah PS, Cecilia D. Polymorphisms in RNA sensing toll like receptor genes and its association with clinical outcomes of dengue virus infection. *Immunobiology*. 2015;220(1):164–8.
101. Garcia-Trejo AR, Falcon-Lezama JA, Juarez-Palma L, Granados J, Zuniga-Ramos J, Rangel H, Barquera R, Vargas-Alarcon G, Ramos C. Tumor necrosis factor alpha promoter polymorphisms in Mexican patients with dengue fever. *Acta Trop*. 2011;120(1–2):67–71.
102. Alagarasu K, Mulay AP, Singh R, Gavade VB, Shah PS, Cecilia D. Association of HLA-DRB1 and TNF genotypes with dengue hemorrhagic fever. *Hum Immunol*. 2013;74(5):610–7.
103. Chuansumrit A, Anantasil N, Sasanakul W, Chaiyaratana W, Tangnaratchakit K, Butthep P, Chunhakan S, Yoksan S. Tumour necrosis factor gene polymorphism in dengue infection: association with risk of bleeding. *Paediatrics and International Child Health*. 2013;33(2):97–101.
104. Djamiatun K, Ferwerda B, Netea MG, van der Ven AJ, Dolmans WM, Faradz SM. Toll-like receptor 4 polymorphisms in dengue virus-infected children. *American Journal of Tropical Medicine & Hygiene*. 2011;85(2):352–4.
105. Swati S, Singh SK, Kavita K, Nikky N, Dhole TN, Rajesh K, Saba H. Analysis of TLR4 (Asp299 Gly and Thr399Ile) gene polymorphisms and mRNA level in patients with dengue infection: a case-control study. *Infect Genet Evol*. 2016;43:412–7.
106. Alagarasu K, Honap T, Mulay AP, Bachal RV, Shah PS, Cecilia D. Association of vitamin D receptor gene polymorphisms with clinical outcomes of dengue virus infection. *Hum Immunol*. 2012;73(11):1194–9.

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