## Molecular identification of '*Candidatus* Phytoplasma asteris' associated with little leaf disease of *Chrysanthemum morifolium*

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**Abstract.** Association of *Candidatus* Phytoplasma asteris' with little leaf disease of *Chrysanthemum morifolium* was detected by nested PCR and identified by nucleotide sequence analysis of PCR products for the first time from India.

*Chrysanthemum morifolium* (family Asteraceae) is an important ornamental plant grown in pots and gardens for its beautiful blooms of various colours, sizes and extensive vase life. Various pathogens such as fungi, viruses, viroids and phytoplasma severely affect chrysanthemum cultivation worldwide (Bouwen and van Zaayen 1995). Natural occurrence of little leaf disease on several *C. morifolium* plants was observed in gardens and nurseries at Lucknow, India during winter of 2005–06. The symptoms were excessive proliferation, tiny narrow leaves and shortening of internodes, which altogether gives rise to 'witches'-broom' appearance (Fig. 1). Since chrysanthemums are propagated through suckers or cuttings and the phytoplasma is known to be transmitted by vegetative propagation through cuttings (Bhat *et al.* 2006), the identification of the causal agent of the little leaf disease of *C. morifolium* was attempted.

To investigate the possibility of a phytoplasma causal agent, the total DNA was isolated from leaf tissue of infected and healthy *C. morifolium* plants following the protocol of Ahrens and Seemüller (1992). The initial PCR was performed using P1/P6 universal primers specific to the 16S rRNA gene (Deng and Hiruki 1991). Further, nested PCR was carried out with primers R16F2n/R16R2 (Gundersen and Lee 1996) employing the initial PCR product as template. Agarose gel electrophoresis of direct and nested PCR products obtained with 16S rRNA-gene-specific primers resulted in the expected size DNA fragments of  $\sim 1.5$  kb and  $\sim 1.4$  kb respectively, from infected plant samples but not from healthy samples. The 1.4-kb amplicon obtained from the nested PCR was cloned, sequenced and the data deposited in GenBank (Accession number DQ431842). BLAST search analysis of DQ431842 revealed 99% sequence similarity with the 16S rRNA gene of Ash witches'-broom, Maize bushy stunt, Dogfennel vellows, Epilobium phyllody, Onion vellows and Aster yellows phytoplasmas (Accession numbers AY566302, AY265208, DQ381534, AY101386, AP006628 and AY265209), respectively, belonging to the 'Candidatus Phytoplasma asteris' (16S rI) group.

A literature survey revealed reports of Chrysanthemum witches'-broom phytoplasma on *C. coronarium* in Japan (Okuda *et al.* 1997), Chrysanthemum yellows phytoplasma on *C. frutescens* in Italy (Bertaccini *et al.* 1990) and Aster yellows phytoplasma on *C. coronarium* in China (Zhong and Shen 2004),



Fig. 1. Naturally infected *Chrysanthemum morifolium* plant showing little leaf symptoms (left), compared with a healthy plant (right).

infecting chrysanthemum cultivars, but to the best of our knowledge, the association of '*Candidatus* Phytoplasma asteris' with little leaf disease of chrysanthemum is the first report of this kind from India.

## Acknowledgements

Authors are grateful to the Director, National Botanical Research Institute (NBRI), Lucknow for providing facilities and Council of Scientific and Industrial Research (CSIR), New Delhi for providing fellowship to S. Kumar.

## References

- Ahrens U, Seemüller E (1992) Detection of DNA of plant pathogenic mycoplasma like organism by a polymerase chain reaction that amplifies a sequence of the 16S rRNA gene. *Phytopathology* **82**, 828–832.
- Bertaccini A, Davis RE, Lee IM, Conti M, Dally EL, Douglas SM (1990) Detection of chrysanthemum yellows mycoplasmalike organism by dot hybridization and Southern blot analysis. *Plant Disease* 74, 40–43.
- Bhat AI, Madhubala R, Hareesh PS, Anadaraj M (2006) Detection and characterization of the phytoplasma associated with a phyllody disease of black pepper (*Piper nigrum*) in India. *Scientia Horticulturae* **107**, 200–204. doi: 10.1016/j.scienta.2005.06.013

- Bouwen I, van Zaayen AZ (1995) Chrysanthemum. In 'Virus and viruslike disease of bulb and flower crops'. (Eds G Lobenstein, RH Lawson, AA Brunt) pp. 396–408. (John Wiley and Sons: New York)
- Deng S, Hiruki D (1991) Amplification of 16S rRNA genes from culturable and nonculturable mollicutes. *Journal of Microbiological Methods* 14, 53–61. doi: 10.1016/0167-7012(91)90007-D
- Gundersen DE, Lee IM (1996) Ultrasensitive detection of phytoplasmas by nested-PCR assays using two universal primer pairs. *Phytopathologia Mediterranea* 35, 144–151.
- Okuda S, Prince JP, Davis RE, Dally EL, Lee IM, Morgen B, Kato S (1997) Two groups of phytoplasmas from Japan distinguished on the basis of amplification and restriction analysis of 16S rDNA. *Plant Disease* 81, 301–305.
- Zhong B-X, Shen Y-W (2004) Accumulation of Pathogenesis-related Type-5 Like Proteins in Phytoplasma infected Garland Chrysanthemum Chrysanthemum coronarium. Acta Biochimica et Biophysica Sinica 36, 773–779.

Manucript received 20 January 2007, accepted 29 January 2007