CYTOGENETICS (CL MARTIN, SECTION EDITOR)

CNVs in Epilepsy

Heather C. Mefford

Published online: 28 June 2014 © The Author(s) 2014. This article is published with open access at Springerlink.com

Abstract Copy number variants (CNVs) are deletions or duplications of DNA. CNVs have been increasingly recognized as an important source of both normal genetic variation and pathogenic mutation. Technologies for genome-wide discovery of CNVs facilitate studies of large cohorts of patients and controls to identify CNVs that cause increased risk for disease. Over the past 5 years, studies of patients with epilepsy confirm that both recurrent and non-recurrent CNVs are an important source of mutation for patients with various forms of epilepsy. Here, we will review the latest findings and explore the clinical implications.

Keywords Copy number variants · Epilepsy · Microdeletions · Neurodevelopmental disorders

Introduction

Epilepsy is a group of conditions characterized by recurrent, unprovoked seizures that result from abnormal synchronized neuronal firing in the brain. Epilepsy will affect up to 1 in 26 individuals and confers a significant health and economic burden [1]. There are many forms of epilepsy that can be distinguished by various characteristics including age of onset, predominant seizure type(s), and etiology [2]. Three broad classes of epilepsy include genetic generalized epilepsy (GGE; formerly idiopathic generalized epilepsy), focal epilepsy, and epileptic encephalopathy, though it should be noted that there are

H. C. Mefford (\boxtimes)

Division of Genetic Medicine, Department of Pediatrics, University of Washington, RR349A, Box 356320, Seattle, WA, USA e-mail: hmefford@uw.edu many specific epilepsy syndromes within each class (Table 1).

The causes of epilepsy are diverse. Non-genetic or acquired etiologies account for 20-30 % of cases and include stroke, head injury, and tumor. In the remaining cases, genetics is thought to play a significant role. In fact, it has been recognized since the time of Hippocrates that epilepsy is, at least in part, genetic. Modern evidence for genetic factors comes from twin studies, family studies, and the identification of single-gene disorders resulting in epilepsy syndromes. Studies in twins show an excess of disease concordance in monozygotic twins compared to dizygotic twins for most types of epilepsy. In a large Australian cohort, generalized epilepsies showed the highest concordance (~ 80 %); focal epilepsies have a lower (36 %) but still significant concordance [3]. Family studies reveal that the overall recurrence risk for epilepsy in first-degree relatives of affected individuals is 2-5 % [4, 5], and at least one study has shown that there is similar increased recurrence for family members of probands with either generalized or focal epilepsy [6]. Finally, large multiplex families in which epilepsy segregates in an autosomal dominant manner have been used to identify linkage regions and causative genes in several different epilepsy syndromes [7].

Despite longstanding knowledge that epilepsy has a strong genetic component, it was not until 1995 that the first gene for a form of epilepsy was identified: mutations in the alpha 4 subunit of the nicotinic acetylcholine receptor, *CHRNA4*, were identified in a large family with autosomal dominant nocturnal frontal lobe epilepsy (ADNFLE) [8]. Since that time, multiple genes in which mutations cause epilepsy have been discovered [9, 10]. While many epilepsy genes encode ion channel subunits, several non-channel genes encoding proteins important for brain development have been recently discovered as well.

Table 1 Examples of epilepsy syndromes

Major class	Examples of specific syndromes
Genetic generalized epilepsy (GGE)	Juvenile myoclonic epilepsy (JME)
	Childhood absence epilepsy (CAE)
	Generalized epilepsy with febrile seizures plus (GEFS+)
Focal epilepsy	Temporal lobe epilepsy
	Autosomal dominant focal epilepsy with auditory features (ADPEAF)
Epileptic encephalopathy	Ohtahara syndrome
	Dravet syndrome
	West syndrome

Copy Number Variants and Human Disease

Copy number variants (CNVs) are large (>1 kb) deletions or duplications of DNA. CNVs can contain zero, one, or many genes and have been increasingly recognized as an important source of both normal genetic variation and pathogenic mutation. There are two major classes of CNVs: recurrent and non-recurrent. Recurrent CNVs are deletions and duplications that occur as a result of nonallelic homologous recombination (NAHR) at meiosis due to a predisposing sequence architecture: 50 kb to 10 Mb of unique DNA flanked by duplicated blocks of sequence that are >10 kb with >95 % sequence identity [11]. Examples of recurrent CNVs associated with neurological or neurodevelopmental disorders include duplications of 17p12 causing Charcot-Marie-Tooth type IA [12], deletions of 15q11-q13 in Prader–Willi and Angelman syndromes [13, 14], and deletions of 7q11 causing Williams-Beurens syndrome [15]. Because of the mechanism by which they are generated, recurrent CNVs in two unrelated individuals with the same disorder have nearly identical breakpoints.

Non-recurrent CNVs occur throughout the genome, but the breakpoints are not consistent. While recurrent CNVs are generated by aberrant recombination, non-recurrent CNVs are often due to errors at replication. There are several mechanisms for the generation of non-recurrent breakpoints that have been described, many of which involve microhomology-few to several identical base pairs at each breakpoint [16]. Non-recurrent CNVs can be simple, where a stretch of DNA is simply cut out of its original location and the ends rejoined, or complex, in which a deletion may be accompanied by insertion or duplication of DNA at the breakpoints for example. It is rare to find two or more patients with the same nonrecurrent CNV. However, comparison of overlapping CNVs in similarly affected patients often reveals a "smallest region of overlap" that can highlight one or a few genes as primarily responsible for the phenotype. Examples include the discovery of *CHD7* as the gene for CHARGE syndrome [17], *EHMT1* as the critical gene in 9q34 deletions (Kleefstra syndrome) [18], and *MBD5* in 2q23.1 deletions [48].

Genome-wide identification of CNVs became efficient with the introduction of array comparative genomic hybridization (CGH) and single nucleotide polymorphism (SNP) microarrays. These technologies allow the detection of submicroscopic CNVs that are too small to be recognized by routine karyotype analysis. Since the introduction of these technologies, the rate of discovery of submicroscopic rearrangements in both affected and unaffected individuals has increased dramatically. Early application of CGH and SNP arrays in control cohorts produced a "CNV landscape" in unaffected individuals [19-27]. The first large disease cohorts to be systematically studied were patients with intellectual disability (ID), followed by autism and schizophrenia [28, 29]. Comparison of CNVs in patients to those in controls combined with the identification of similar CNVs in multiple affected individuals led to the discovery of novel, disease-associated CNVs. Systematic CNV discovery in patients with epilepsy followed, and there has been steady progress that provides new insight into the genetics of epilepsy and related disorders. The remainder of this review will focus on CNV discovery and characterization in patients with epilepsy and recommendations for clinical testing.

Non-Recurrent CNVs in Epilepsy

The importance of non-recurrent CNVs in epilepsy has actually been known for some time. For example, techniques such as quantitative PCR and multiplex ligation-dependent probe amplification (MLPA) have been used to detect single- and multiple-exon deletions and duplications in known genes such as *SCN1A* [30, 31]. However, qPCR and MLPA are assays directed at specific locations in the genome, which means CNVs elsewhere will not be detected.

Real advances in CNV discovery came from the application of genome-wide investigations using array CGH or SNP microarrays. Heinzen and colleagues performed CNV discovery in a large cohort of 3812 patients with primarily focal epilepsy syndromes and identified an excess of large (>1 Mb) deletions in affected individuals, the majority of which were seen in one individual each [32••]. We performed array CGH studies in 517 patients with various types of epilepsy (primarily generalized); ~5% of patients carried a non-recurrent CNV that affected at least one gene and was not seen in controls [33]. In a study of 102 patients with epilepsy with or without other neurodevelopmental abnormalities, 23/102 individuals had at least one non-polymorphic CNV [34]. Investigation of patients with epileptic encephalopathy syndromes also confirms the role of non-recurrent CNVs in severe epilepsies [35•]. Together, these studies confirm that CNVs are important contributors to the genetics of broad classes of epilepsy.

Because non-recurrent CNVs are rare and often unique, it can be difficult to interpret the clinical significance of any given CNV. Several criteria can be used, including size, gene content, presence or absence in control studies, and inheritance [36]. While a CNV in a single patient may be difficult to interpret, by collecting multiple patients with overlapping non-recurrent CNVs, it becomes possible to determine a critical region and, sometimes, a single critical gene for a given condition. This approach has been successful in highlighting several novel epilepsy genes. For example, Depienne and colleagues [37] performed array CGH on a series of patient with a Dravet-like syndrome and identified a single patient with a deletion involving PCDH19 on chromosome X. Sequence analysis of the gene in additional patients revealed sequence mutations and established the gene as a cause of epilepsy restricted to females with ID. Similarly, rare reports of deletions involving the GRIN2A gene highlighted the gene as a potentially important gene [38]. Indeed, we and others have identified mutations in GRIN2A in 5-20 % of patients with epilepsy syndromes associated with language deficits (epilepsy aphasia syndromes) [39, 40, 41•]. We recently identified a patient with a large deletion of 15q26 (reported in [42•]). Comparing the deletion in our patient to other 15q26 deletions in the literature highlighted a single gene in the smallest region of overlap: CHD2. Using high-throughput targeted sequencing of CHD2 in 500 patients with epileptic encephalopathy, we identified de novo pathogenic mutations in 1.2 % of cases [43]. Additional studies of overlapping rearrangements published in large disease cohorts, as well as smaller case reports, are likely to yield important causative genes.

Recurrent Deletions in Epilepsy

A major—and somewhat surprising—advance in the field of epilepsy genetics has been the discovery of recurrent CNVs in patients. The importance of recurrent CNVs as causes of ID syndromes, such as Prader–Willi, Angelman, Smith– Magenis, and velocardiofacial syndromes, has been known since the 1980s. More recently, genome-wide CGH in large cohorts of patients with ID led to the identification of several novel recurrent microdeletion syndromes [29, 44]. The first study that highlighted the importance of CNVs in the genetic etiology of epilepsy was the discovery of recurrent 15q13.3 deletions in patients with generalized epilepsy [45••]. The 15q13.3 microdeletion (chr15: 31,000,000–32,500,000, hg19) was first described in patients with ID, but it was noted that most patients also suffered from seizures [46]. This observation led to a collaborative effort to determine the frequency of the deletion in a cohort of 1,223 patients with generalized epilepsy, most of whom did not have ID or other neurodevelopmental abnormalities. Indeed, 12/1,223 (1 %) patients carried a 15q13.3 deletion compared to 0/3,699 control individuals [45••]. Several subsequent studies confirmed this finding [33, 47, 48], establishing the deletion as one of the most prevalent genetic risk factors for GGE with an estimated odds ratio of 68 (29–181) [47].

Two other recurrent deletions have been firmly associated with epilepsy. De Kovel and colleagues investigated a cohort of 1,234 individuals with GGE and 3,022 controls for recurrent CNVs [49]. In addition to 15q13.3 deletions, they found recurrent deletions at 16p13.11 (chr16: 15,500,000-16,300,000, hg19) and 15q11.2 (chr15: 22,800,000-23,100,000, hg19) in 0.5 and 1 % of patients, respectively, representing a significantly increased frequency compared to controls. Heinzen and colleagues performed CNV genotyping in 3,812 individuals with epilepsy, most of which presented with a focal epilepsy syndrome [32...]. Deletions of 16p13.11 were also enriched in patients compared to controls (23/3,812 vs 0/1299). While 15q11.2 deletions were identified in 24/3,812 patients, there was not a significant enrichment compared to the frequency in controls (3/1, 299). In a study of 517 patients with various types of epilepsy, we identified five patients each with deletions at 15q11.2, 15q11.3, and 16p13.11, again emphasizing the importance of each of these as frequent genetic susceptibility factors in epilepsy [33]. Of note, in an investigation of 315 patients with epileptic encephalopathy, there were no occurrences of 15q13.3, 16p13.11, or 15q11.2 deletions [35•], suggesting a different genetic architecture for this class of disorders.

Blurring the Lines: Shared Genetics of Neurodevelopmental Disorders

Each of the three epilepsy-associated recurrent deletions must be regarded as a risk factor for disease. In each case, the deletion may be de novo or inherited from an affected or unaffected parent. Importantly, all three deletions also confer risk for other neurodevelopmental disorders. As described above, deletions of 15q13.3 were first identified in patients from an ID cohort [46]. The deletion is also enriched in patients with schizophrenia [50, 51••] and is seen in patients with autism spectrum disorder and nonspecific developmental delays [52, 53]. Similarly, deletions of 16p13.11 and 15q11.2 are also associated with a wide range of neurodevelopmental and neuropsychiatric conditions [32••, 54–57]. Interestingly, the 15q13.3 deletion appears to confer risk specifically for generalized forms of epilepsy—though present in 0.5-1 % of most GGE cohorts, it was not reported in >3,000 patients with focal epilepsy syndromes [32••].

Perhaps not surprisingly, patients with one of the epilepsy-associated recurrent deletions may present with a more severe phenotype than expected. This is especially true for GGE, which is not typically characterized by other neurocognitive deficits. Muhle and colleagues [58] identified 4/570 patients with various types of epilepsy who carried a 15q13.3 deletion. Detailed phenotype analysis revealed that all patients had absence epilepsy as well as some degree of ID. Similar features were described in two other families segregating the 15q13.3 deletion [59]. More recently, a systematic comparison of the frequency of recurrent CNVs in patients with GGE compared to patients with GGE and ID showed that patients with "dual disability" are more likely to carry one of the three epilepsyassociated CNVs than patients with GGE without other features [42•]: 10 % of the GGE + ID patients had one of the three recurrent CNVS compared to ~ 3 % of patients with GGE but no ID. Other recurrent CNVs that are associated with neurodevelopmental disorders are also found in patients with epilepsy, though not as frequently as the three deletions discussed above [28, 29, 60]. Examples include deletions and duplications of 1q21.1, 22q11.2, and 16p11.2.

Who Should be Tested?

The role of CNVs in the genetic etiology of epilepsy has been clearly established, and diagnostic testing by chromosome microarray should be considered in this population. There is a clear consensus that chromosome microarray testing should be the first-line test in the diagnosis of patients with neurodevelopmental disorders or multiple congenital anomalies [36]. Given the overlapping genetic etiologies of a broad range of neurodevelopmental disorders, patients with epilepsy that is associated with other findings such as ID, autistic features, or developmental delays should be tested. Indeed, for GGE with ID, the diagnostic yield will be 10 % or greater. Similarly, patients with brain malformations or other congenital abnormalities and patients with epileptic encephalopathy without a clear diagnosis should undergo CNV testing. The yield of CNV testing in patients with epilepsy and no other features may be lower, but a CNV involving a known epilepsy gene would be an important diagnostic finding.

Conclusions

The role of CNVs in the genetic etiology of epilepsy is now well established. Both recurrent and non-recurrent CNVs have been identified in most major classes of epilepsy. In some cases, CNVs are highlighting the shared genetic susceptibility for a range of neurodevelopmental and neuropsychiatric conditions. It is clear that the identification of disease-associated CNVs will lead to improved diagnosis and prognosis counseling. Recurrence risk counseling will remain complicated for CNVs with broad effects but is nevertheless an important consideration for families. Finally, as patients with shared genetic etiologies of epilepsy are identified, studies of genotype–phenotype correlation, natural history, and therapeutic response to specific anti-epileptic drugs can be performed, which will lead to improved long-term care and outcomes for patients.

Disclosure HC Mefford declares no conflicts of interest.

Human and Animal Rights and Informed Consent All studies by HC Mefford involving animal and/or human subjects were performed after approval by the appropriate institutional review boards. When required, written informed consent was obtained from all participants.

Open Access This article is distributed under the terms of the Creative Commons Attribution License which permits any use, distribution, and reproduction in any medium, provided the original author(s) and the source are credited.

References

Papers of particular interest, published recently, have been highlighted as:

- Of importance
- •• Of major importance
- 1. England MJ, et al., editors. Epilepsy across the spectrum: promoting health and understanding. Washington: The National Academies Press; 2012.
- Berg AT, Scheffer IE. New concepts in classification of the epilepsies: entering the 21st century. Epilepsia. 2011;52(6): 1058–62.
- Berkovic SF, et al. Epilepsies in twins: genetics of the major epilepsy syndromes. Ann Neurol. 1998;43(4):435–45.
- Annegers JF, et al. The risks of seizure disorders among relatives of patients with childhood onset epilepsy. Neurology. 1982;32(2): 174–9.
- Hemminki K, et al. Familial risks for epilepsy among siblings based on hospitalizations in Sweden. Neuroepidemiology. 2006;27(2):67–73.
- 6. Ottman R, et al. Seizure risk in offspring of parents with generalized versus partial epilepsy. Epilepsia. 1989;30(2):157-61.
- Reid CA, Berkovic SF, Petrou S. Mechanisms of human inherited epilepsies. Prog Neurobiol. 2009;87(1):41–57.
- Steinlein OK, et al. A missense mutation in the neuronal nicotinic acetylcholine receptor alpha 4 subunit is associated with autosomal dominant nocturnal frontal lobe epilepsy. Nat Genet. 1995;11(2):201–3.
- 9. Helbig I, et al. Navigating the channels and beyond: unravelling the genetics of the epilepsies. Lancet Neurol. 2008;7(3):231–45.
- Hildebrand MS, et al. Recent advances in the molecular genetics of epilepsy. J Med Genet. 2013;50(5):271–9.

- 11. Stankiewicz P, Lupski JR. Genome architecture, rearrangements and genomic disorders. Trends Genet. 2002;18(2):74–82.
- Lupski JR, et al. DNA duplication associated with Charcot– Marie–Tooth disease type 1A. Cell. 1991;66(2):219–32.
- Magenis RE, et al. Is Angelman syndrome an alternate result of del(15)(q11q13)? Am J Med Genet. 1987;28(4):829–38.
- Butler MG, Meaney FJ, Palmer CG. Clinical and cytogenetic survey of 39 individuals with Prader-Labhart-Willi syndrome. Am J Med Genet. 1986;23(3):793–809.
- Perez Jurado LA, et al. Molecular definition of the chromosome 7 deletion in Williams syndrome and parent-of-origin effects on growth. Am J Hum Genet. 1996;59(4):781–92.
- Lee JA, Carvalho CM, Lupski JR. A DNA replication mechanism for generating nonrecurrent rearrangements associated with genomic disorders. Cell. 2007;131(7):1235–47.
- Vissers LE, et al. Mutations in a new member of the chromodomain gene family cause CHARGE syndrome. Nat Genet. 2004;36(9):955–7.
- Kleefstra T, et al. Disruption of the gene Euchromatin Histone Methyl Transferase1 (Eu-HMTase1) is associated with the 9q34 subtelomeric deletion syndrome. J Med Genet. 2005;42(4): 299–306.
- de Stahl TD, et al. Profiling of copy number variations (CNVs) in healthy individuals from three ethnic groups using a human genome 32 K BAC-clone-based array. Hum Mutat. 2008;29(3): 398–408.
- 20. Iafrate AJ, et al. Detection of large-scale variation in the human genome. Nat Genet. 2004;36(9):949–51.
- Locke DP, et al. Linkage disequilibrium and heritability of copynumber polymorphisms within duplicated regions of the human genome. Am J Hum Genet. 2006;79(2):275–90.
- 22. Pinto D, et al. Copy-number variation in control population cohorts. Hum Mol Genet. 2007;16(Spec No. 2):R168–73.
- 23. Redon R, et al. Global variation in copy number in the human genome. Nature. 2006;444(7118):444–54.
- 24. Sebat J, et al. Large-scale copy number polymorphism in the human genome. Science. 2004;305(5683):525–8.
- 25. Sharp AJ, et al. Segmental duplications and copy-number variation in the human genome. Am J Hum Genet. 2005;77(1):78–88.
- Simon-Sanchez J, et al. Genome-wide SNP assay reveals structural genomic variation, extended homozygosity and cell-line induced alterations in normal individuals. Hum Mol Genet. 2007;16(1):1–14.
- Zogopoulos G, et al. Germ-line DNA copy number variation frequencies in a large North American population. Hum Genet. 2007;122(3–4):345–53.
- Mefford HC, Batshaw ML, Hoffman EP. Genomics, intellectual disability, and autism. N Engl J Med. 2012;366(8):733–43.
- Mefford HC, Eichler EE. Duplication hotspots, rare genomic disorders, and common disease. Curr Opin Genet Dev. 2009;19(3):196–204.
- Marini C, et al. SCN1A duplications and deletions detected in Dravet syndrome: implications for molecular diagnosis. Epilepsia. 2009;50(7):1670–8.
- Mulley JC, et al. A new molecular mechanism for severe myoclonic epilepsy of infancy: exonic deletions in SCN1A. Neurology. 2006;67(6):1094–5.
- 32. •• Heinzen EL, et al. Rare deletions at 16p13.11 predispose to a diverse spectrum of sporadic epilepsy syndromes. Am J Hum Genet. 2010;86(5):pp. 707–18. This study highlights the importance of the recurrent 16p13.11 deletion and other CNVs in focal epilepsies.
- Mefford HC, et al. Genome-wide copy number variation in epilepsy: novel susceptibility loci in idiopathic generalized and focal epilepsies. PLoS Genet. 2010;6:e1000962.

- Bartnik M, et al. Application of array comparative genomic hybridization in 102 patients with epilepsy and additional neurodevelopmental disorders. Am J Med Genet B Neuropsychiatr Genet. 2012;159B(7):760–71.
- 35. Mefford HC, et al. Rare copy number variants are an important cause of epileptic encephalopathies. Ann Neurol. 2011;70(6):pp. 974–85. This study identifies rare pathogenic CNVs in severe, early-onset epilepsy syndromes. Most CNVs in this class of epilepsy are non-recurrent.
- Miller DT, et al. Consensus statement: chromosomal microarray is a first-tier clinical diagnostic test for individuals with developmental disabilities or congenital anomalies. Am J Hum Genet. 2010;86(5):749–64.
- Depienne C, et al. Sporadic infantile epileptic encephalopathy caused by mutations in PCDH19 resembles Dravet syndrome but mainly affects females. PLoS Genet. 2009;5(2):e1000381.
- Reutlinger C, et al. Deletions in 16p13 including GRIN2A in patients with intellectual disability, various dysmorphic features, and seizure disorders of the rolandic region. Epilepsia. 2010;51(9):1870–3.
- 39. Lesca G, et al. GRIN2A mutations in acquired epileptic aphasia and related childhood focal epilepsies and encephalopathies with speech and language dysfunction. Nat Genet. 2013;45(9): 1061–6.
- 40. Carvill GL, et al. GRIN2A mutations cause epilepsy-aphasia spectrum disorders. Nat Genet. 2013;45(9):1073–6.
- 41. Lemke JR, et al. Mutations in GRIN2A cause idiopathic focal epilepsy with rolandic spikes. Nat Genet. 2013;45(9):pp. 1067–72. Based on previous studies that find CNVs involving GRIN2A, three groups independently sequenced the gene. Overall, mutations in GRIN2A are responsible for 5–20% of patients with epilepsy-aphasia disorders.
- 42. Mullen SA, et al. Copy number variants are frequent in genetic generalized epilepsy with intellectual disability. Neurology. 2013;81(17):p. 1507–14. This study highlights the increased diagnostic yield of CNV testing in patients with epilepsy with intellectual disability—so called 'dual disabilities'.
- Carvill GL, et al. Targeted resequencing in epileptic encephalopathies identifies de novo mutations in CHD2 and SYNGAP1. Nat Genet. 2013;45(7):825–30.
- 44. Sharp AJ, et al. Discovery of previously unidentified genomic disorders from the duplication architecture of the human genome. Nat Genet. 2006;38(9):1038–42.
- 45. •• Helbig I, et al. 15q13.3 microdeletions increase risk of idiopathic generalized epilepsy. Nat Genet. 2009;41(2):pp. 160–2. *This is one of the first studies to identify a recurrent CNV as an important risk factor for epilepsy.*
- 46. Sharp AJ, et al. A recurrent 15q13.3 microdeletion syndrome associated with mental retardation and seizures. Nat Genet. 2008;40(3):322–8.
- Dibbens LM, et al. Familial and sporadic 15q13.3 microdeletions in idiopathic generalized epilepsy: precedent for disorders with complex inheritance. Hum Mol Genet. 2009;18(19):3626–31.
- Kirov A, et al. 15q13.3 microdeletions in a prospectively recruited cohort of patients with idiopathic generalized epilepsy in Bulgaria. Epilepsy Res. 2013;104(3):241–5.
- 49. de Kovel CG, et al. Recurrent microdeletions at 15q11.2 and 16p13.11 predispose to idiopathic generalized epilepsies. Brain. 2010;133(Pt 1):23–32.
- International Schizophrenia Consortium. Rare chromosomal deletions and duplications increase risk of schizophrenia. Nature. 2008;455(7210):237–41.
- 51. •• Stefansson H, et al. Large recurrent microdeletions associated with schizophrenia. Nature. 2008;455(7210):pp. 232–6. *These two studies (refs 50-51) were the first to identify recurrent CNVs*

as important risk factors for schizophrenia. Each associated CNV also confers risk for other neurodevelopmental disorders.

- 52. Pagnamenta AT, et al. A 15q13.3 microdeletion segregating with autism. Eur J Hum Genet. 2009;17(5):687–92.
- van Bon BW, et al. Further delineation of the 15q13 microdeletion and duplication syndromes: a clinical spectrum varying from non-pathogenic to a severe outcome. J Med Genet. 2009;46(8): 511–23.
- 54. Hannes FD, et al. Recurrent reciprocal deletions and duplications of 16p13.11: the deletion is a risk factor for MR/MCA while the duplication may be a rare benign variant. J Med Genet. 2009;46(4):223–32.
- Kirov G, et al. Support for the involvement of large cnvs in the pathogenesis of schizophrenia. Hum Mol Genet. 2009;18(8): 1497–503.

- Need AC, et al. A genome-wide investigation of SNPs and CNVs in schizophrenia. PLoS Genet. 2009;5(2):e1000373.
- Ullmann R, et al. Array CGH identifies reciprocal 16p13.1 duplications and deletions that predispose to autism and/or mental retardation. Hum Mutat. 2007;28(7):674–82.
- Muhle H, et al. Absence seizures with intellectual disability as a phenotype of the 15q13.3 microdeletion syndrome. Epilepsia. 2011;52(12):e194–8.
- 59. Coppola A, et al. Different electroclinical picture of generalized epilepsy in two families with 15q13.3 microdeletion. Epilepsia. 2013;54(5):e69–73.
- Mefford HC, Mulley JC. Genetically complex epilepsies, copy number variants and syndrome constellations. Genome Med. 2010;2(10):71.