

### SHORT COMMUNICATION

# Evaluation of Nanoporous Gold with Controlled Surface Structures for Laser Desorption Ionization (LDI) Analysis: Surface Area Versus LDI Signal Intensity

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#### **Abstract**

The structural effect of a nanoporous gold (NPG) surface on the signal intensities of laser desorption ionization-mass spectrometry (LDI-MS) were investigated using NPG surfaces with controlled structures. The relationship between surface area and LDI efficiency was compared and evaluated. Comparisons between bare flat gold and NPG surfaces show that nanostructures increased LDI efficiency. We also found that the LDI signal decreased with increasing depth of nanoporous layers, thus increasing the surface area. This result agrees with a previous report (Shin J. A. et al., *J. Am. Soc. Mass Spectrom.* 2010, 21, 989) in which the LDI efficiency of small molecules decreased for ZnO wires with longer lengths. This observation was explained by the penetration and deposition of samples into locations inaccessible to photons because of structural screening. The LDI-MS analysis of oils with NPG surfaces (but without matrix) showed the same trend whereby the NPG with about a 200 nm depth of porous area showed the highest sensitivity. This study clearly shows that the active surface area for solution chemistry can differ from LDI-MS and that NPGs can function as a substrate for LDI oil analysis.

**Key words:** Mass spectrometry, Nanoporous gold, Laser desorption ionization, Petroleum, Surface area

#### Introduction

L aser desorption ionization (LDI) with organic matrix, typically referred to as MALDI is widely used for

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analysis of biomolecules and polymers. However, this approach is limited for the analysis of a mixture (such as petroleum) since peaks originating from the matrix can be abundant in lower m/z ranges (e.g., m/z < 500) and may overlap with peaks from petroleum. Therefore, developing a method that can be used in conjunction with LDI to improve the ionization efficiency and limit matrix interference would be desirable.

Particles or surfaces with nanostructures have been combined with LDI to improve ionization efficiency and

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reduce matrix interference [1-13]. Although many studies have employed nanostructures to improve LDI efficiency and selectivity, relatively few reports have examined how the nanostructure is related to LDI efficiency [9]. In this study, LDI efficiency obtained from four surfaces with different gold (Au) nanostructures were compared, and the dependence of LDI efficiency on the surface areas was evaluated. Nanoporous Au (NPG) structures adopted in this study as LDI substrates have attracted considerable attention due to their unique properties and possible applications in catalysis [14]. The NPG surface area was controlled by the deposition charge during the electrochemical formation of NPG structures [15]. During the process, discrepancies between surface area and LDI efficiency were investigated. The NPG structures with the best LDI efficiencies were selected and applied for oil analysis.

# **Experimental**

## Preparation of NPG Surfaces

NPG surfaces were prepared by electrodeposition of Ag/Au alloy layers followed by selective dissolution of silver (Ag) in concentrated nitric acid [15]. Detailed information on the preparation of NPGs can be found in previous reports [16]. Briefly, the deposition charge of 0.01, 0.02, 0.04, 0.06, and 0.08 C was applied to control the thickness of the Ag/Au alloy layers at a constant potential of -0.95 V (versus Ag/ AgCl, 3 MKCl). For convenience, each substrate prepared with different deposition charge is referred to as NPG #1 (0.01 C), #2 (0.02 C), #3 (0.04 C), #4 (0.06 C), and #5 (0.08 C), respectively, for the rest of the manuscript. The alloy layers on Au/silicon (Si) wafers were transferred to concentrated nitric acid to dissolve Ag. Au nanoplate substrates were electrodeposited on Au/Si wafers from a solution containing 15 mM KAu(CN)<sub>2</sub> and 0.25 MNa<sub>2</sub>CO<sub>3</sub> at -1.1 V with a deposition charge of 0.04C [16]. Scanning

electron microscopy (SEM) characterization was performed using a LEO 1530 Field Emission SEM (Carl Zeiss, Jena, Germany). For cross-sectional SEM images, the backside of the electrodeposited Au/Si wafer was scraped with a diamond pencil and split into two parts.

#### Mass Spectrometry Analysis

Detailed instrumental parameters for the LDI-time-of-flight (TOF) MS experiment are provided in the Supporting Information. Verapamil hydrochloride (Aldrich, St. Louis, MO, USA) solution of 3  $\mu$ g/ $\mu$ L was prepared in 50 % acetonitrile/water (vol/vol) and diluted 6-fold with 0.1 % TFA in water containing 30 % acetonitrile for LDI analyses. The sample solution (0.5  $\mu$ L) was placed on targets for MS analyses. Oil samples extracted using hydrous pyrolysis and the in situ simulator from rocks collected in the Green River Formation were studied [17]. Difference between the hydrous pyrolysis and the in situ simulator methods was explained in the supporting information. Oil samples were diluted to 1 mg/mL in 100 % toluene, and 0.5  $\mu$ L of oil solution was spotted on the targets.

# Results and Discussion

Electrochemical Characterization of NPG Surfaces with Different Surface Structures

Figure 1 shows the SEM images of some of Au target substrates with different surface morphologies used in this study. Figure 1a–c show that the NPG substrates with different deposition charges retain regular nanoporous Au surface structures in which the size of ligaments and pores are around 30 nm and 20 nm, respectively. The surface area of these NPG substrates were electrochemically estimated by integrating the charges consumed for reduction of the surface oxide monolayer from cyclic voltammograms of NPG substrates obtained from 0.1 M H<sub>2</sub>SO<sub>4</sub> [18]. A plot showing

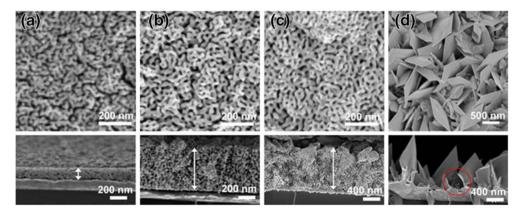


Figure 1. SEM images of Au target substrates with different surface morphologies. The deposition charges for the preparation of Ag/Au layers for NPG preparation were (a) 0.01C, (b) 0.04C, and (c) 0.08C. (d) Au nanoplate substrate. Top and bottom rows correspond to the surface and cross-sectional SEM images, respectively. Arrows represent the overall thickness of the electrodeposited nanostructure layers

**Table 1.** Average Peak Intensities and Standard Deviations of Verapamil Observed from Five Measurements by Use of NPG Targets

NPG target no.	Averaged signal-to-noise ratio	Standard deviation
1	590	25
2	380	130
3	130	46
4	94	34
5	130	42

relationship between the relative surface area (RSA) of NPG surfaces normalized with respect to the flat Au surface and deposition charges is provided in (Figure 1S, Supporting Information). RSA of the NPG substrates with higher deposition charges increased, as reported previously [15]. Figure 1d shows the structure of the Au nanoplate substrates consisting of plate-type protrusions with sharp edges. The RSA of the Au nanoplate substrate was 5.0.

To examine the three-dimensional structures, cross-sectional SEM images of the substrates were obtained (bottom of Figure 1). The cross-sectional images show that depths of the porous layer for NPG #1, #3, and #5 increase with higher deposition charges. For example, the depth of the porous layer for NPG #1 was<200 nm, but the depth of NPG #5 was >800 nm. The structure of the Au nanoplate consisted of plate-type protrusions with sharp edges, which were confirmed by the cross-sectional image.

# Evaluation and Comparison of NPG Surfaces for LDI-TOF-MS

Equal amounts of verapamil were loaded onto each NPG surface and analyzed by LDI-MS. Examples of LDI-MS spectra of verapamil loaded on different NPG surfaces is presented in Figure 2S (Supporting Information). Ion abundance measured spectra are summarized in Table 1. No signal was observed from the bare Au surface (data not shown) but signals were observed from various NPG surfaces. The fact that no signal was observed from bare Au, but signals could be observed with NPG surfaces, strongly suggests that the nanoporous structure enhances LDI efficiency.

The relative surface areas of the NPG calculated from cyclic voltammograms were 10–60 times higher than that of the flat Au surface. An increased surface area can enhance LDI signals [1, 2, 7, 8, 10], but comparisons among NPG surfaces showed differences between surface area and LDI efficiency. Although the RSA is larger by the order of NPG #5>#4>#3>#2>#1 (refer to Figure 1S, Supporting Information), the LDI signal from NPG #1 was larger than those from NPG #2 and #3. No significant difference was observed from NPG #3, #4, and #5. Thus, the increased surface area does not necessarily correlate with LDI efficiency.

Oils samples were loaded on the NPG surfaces without matrix and the obtained LDI-TOF-MS spectra were compared and presented in the supporting information (refer to Figure S3, Supporting Information). The LDI-MS analysis of oils showed the same trend where the NPG #1 was the

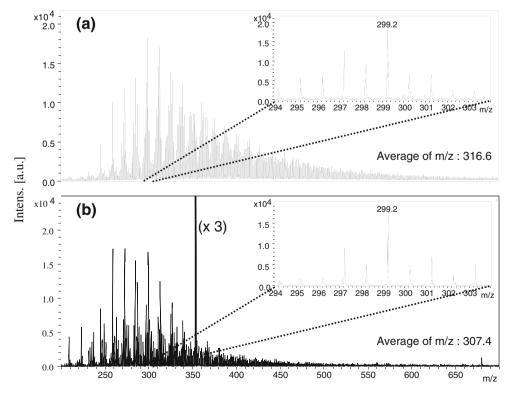


Figure 2. LDI-MS spectra and their expanded view of oil samples each prepared by (a) the hydrous pyrolysis and (b) in situ simulator methods

most sensitive. NPG #1 was selected and used for further analyses of oil samples. Oil samples extracted from oil shale rocks by two different methods were loaded onto the NPG plates and analyzed. The obtained spectra are presented in Figure 2. The big peak near m/z 350 in Figure 2b is originated from crude oil and not from the NPG plates because the peak was found only with the oil shale sample. The average molecular weights can be calculated from the spectra in terms of the magnitude-weighted average as

$$\label{eq:Molecular weight} \text{Molecular weight}_{average} = \frac{\sum_{i} I_{i} \times (molecular \, weight)_{i}}{\sum_{i} I_{i}} \ \ (1)$$

where  $I_i$  and (molecular weight)<sub>i</sub> are the abundance and molecular weight value of peak i, respectively.

The average molecular weights calculated from the spectra obtained from the hydrous pyrolysis and in situ simulator methods are 316.7 and 307.4, respectively. This agrees with a previous study [17] in which a higher molecular weight distribution was observed for samples prepared by hydrous pyrolysis compared to using the in situ simulator method.

The lack of (or even negative) correlation between surface area and LDI efficiency can be partly explained by the deposition of samples onto areas inaccessible by laser photons. Since electrochemical reactions occur in a solution phase, areas partially covered by or located under other structures are accessible to analyte and are electrochemically active. However, this concept does not apply for the LDI experiment. The area partially covered by other structures could be shaded from laser flux and thus inaccessible to photons. For example, if the solution penetrates deeply into the porous layer in Figure 1b and c because of gravity, the laser photons may not be able to reach them. Another example can be found in the SEM image of the Au nanoplate. The area marked by the red circle in Figure 1 may not be exposed to the laser photon flux because of its structural environment. A previous study reported that signal intensity obtained by LDI gradually decreased as ZnO nanowire length increased over 250 nm [9]. They also noted that physical screening using the nanowire could be partly responsible for the observed phenomena. It should be noted that laser shots may change surface morphologies of the substrates. However, since the diameter of laser beam is very small (~100 μm), the substrates could be reused up to two or three times without losing reproducibility.

#### **Conclusions**

This study demonstrates that surface morphologies are associated with LDI efficiency. We found that surface area, especially that calculated using solution chemistry, is not correlated with LDI-MS efficiency. Combined with the results of a previous study, photon screening of a nanostructure must be optimized to maximize the efficiency of nanostructure-assisted LDI-MS analysis. Additionally, the nanostructure-assisted LDI-MS analysis may be applicable to oil analysis.

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