



Post-infection irritable bowel syndrome in the tropical and subtropical regions: *Vibrio cholerae* is a new cause of this well-known condition

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Following acute infective gastroenteritis, 4% to 37% of patients develop new-onset persistent bowel symptoms fulfilling the Rome criteria for irritable bowel syndrome (IBS), an entity recently named as post-infection IBS (PI-IBS) [1]. The acute episode of gastroenteritis should fulfill at least two of the four criteria, namely diarrhea, fever, vomiting, and positive stool culture. PI-IBS develops mostly following acute infectious diarrhea due to multiple etiological agents, more so due to invasive pathogens (Table 1). However, a few studies did report the occurrence of PI-IBS following infection due to less invasive pathogens causing diarrhea such as viruses [19, 22]. Hence, the conventional belief that the PI-IBS occurs predominantly following acute gastroenteritis due to invasive pathogen is no more tenable. Moreover, two recent studies, one from Bangladesh and another one published in this issue of the *Journal* from India, showed the occurrence of PI-IBS following infection with *Vibrio cholerae* (*V. cholerae*), conventionally known as a noninvasive pathogen, challenging this belief further [27, 28].

In a recent study from Bangladesh by Rahman and Ghoshal, of 345 patients with acute gastroenteritis admitted to the International Centre for Diarrheal Disease Research, Dhaka, 16.5% developed PI-IBS as compared with 2.6% controls during 12-month follow up; similarly, 7.4% of them developed functional dyspepsia (FD) in contrast to 0.6% controls during the same follow up duration. Of 245 patients undergoing etiological workup for gastroenteritis, 39% had cholera. Frequency of PI-IBS was

similar among patients with cholera and those with other infection causing acute gastroenteritis. In the Indian study published in this issue of the *Journal*, of 136 patients with acute gastroenteritis admitted to SCB Medical College, Cuttack, India, 25.7% developed PI-IBS at 6 months. Of 11 patients who had cholera, 4 developed PI-IBS, further proving the role of *V. cholerae* in the development of this condition.

Considering the high global burden of cholera, the clinical, epidemiological, societal, and economic implications of the finding that cholera can lead to PI-IBS cannot be overestimated. There are 1.3 billion people at risk of cholera in the 69 cholera-endemic countries [29, 30]. It is estimated that each year, there are 1.3 to 4.0 million cases of cholera [29]. The actual number may be higher due to under-reporting [29, 31]. Of 69 countries in the world considered as cholera-endemic, India, Nigeria, China, Ethiopia, and Bangladesh are the countries in which the largest number of people are at risk for developing cholera. The finding that PI-IBS can develop following cholera has important economic implication for these nations not only due to the potential risk of mortality due to cholera, its morbidity, health care burden, and loss of work productivity of the inhabitants but also because of fear among travelers from the developed countries of developing chronic gastrointestinal (GI) diseases such as PI-IBS and PI-FD, potentially resulting in national economic loss. More studies are needed on these issues.

Mechanisms of cholera-induced PI-IBS

The rapidly developing non-inflammatory diarrhea, which is typical characteristics of cholera, is widely known to be caused by cholera toxin (CT). However, *V. cholerae* strain devoid of genes for CT can cause mild to moderate diarrhea suggesting additional mechanisms for diarrhea in cholera [32–34]. Apart from CT, there are three more toxins (zonula

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Table 1 Incidence, etiology of acute gastroenteritis, and risk factors for post-infection irritable bowel syndrome in cases and controls

Study details	Country	Cause of gastroenteritis	IBS in cases (%)	IBS in controls	Risk factors for post-infection IBS
McKendrick and Read (1994) [2]	UK	<i>Salmonella</i>	12/38 (31.6)	No control	Severity of acute illness, vomiting, and weight loss
Gwee et al. (1996) [3]	UK	<i>Shigella</i> , <i>Campylobacter</i> , <i>Salmonella</i>	20/75 (26.6)	No control	Anxiety, depression, somatisation, and neurotic trait
Neal et al. (1997) [4]	UK	Bacteria	23/347 (6.6)	No control	Longer duration of diarrhea, younger age, and female sex
Gwee et al. (1999) [5]	UK	–	19/109 (17.4)	Psychological and rectal biopsy: 21 HS	Psychological factors and persistent rectal inflammation
Rodriguez and Ruigomez (1999) [6]	UK	Bacteria	14/318 (4.4)	2027/584308 (0.3%)	Not evaluated
Mearin et al. (2005) [7]	Spain	<i>Salmonella</i>	13.2%	1.5%	No risk factor identified
Ilnyckyj et al. (2003) [8]	Canada	Traveler's diarrhea	2/48 (4.2)	1/61 (1.6%)	Not evaluated
Dunlop et al. (2003) [9]	UK	<i>Campylobacter</i>	103/747 (13.8)	No control	Increased enterochromaffin cells in lamina propria and depression
Parry et al. (2003) [10]	UK	<i>Campylobacter</i> , <i>Salmonella</i>	18/108 (16.7)	4/219 (1.9%)	Not evaluated
Wang et al. (2004) [11]	China	<i>Shigella</i>	24/295 (8.1)	2/243 (0.8%)	Longer diarrhea, IL-1 β mRNA expression, and mast cell in ileum and rectosigmoid
Okhuysen et al. (2004) [12]	USA	Traveler's diarrhea	60 (6)	No control	More diarrhea, medical consultation, and stool negative for the pathogen
Ji et al. (2005) [13]	Korea	<i>Shigellosis</i>	15/101 (14.8)	6/102 (5.8%)	Diarrhea duration
Parry et al. (2005) [14]	UK	Bacteria	16/107 (15)	No control	Smoking
Kim et al. (2006) [15]	Korea	<i>Shigella</i>	13/95 (13.6)	4/105 (3.8%)	Pre-existing FBD other than IBS
Marshall et al. (2006) [16]	Canada	<i>E. coli</i> , <i>Campylobacter</i>	417/1368 (30.5)	71/701 (10.2%)	Young age, female, bloody stools, weight loss, and long diarrhea
Borgaonkar et al. (2006) [17]	Canada	Bacteria	7/191 (3.7)	No control	Fever during gastroenteritis
Stermmer et al. (2006) [18]	Israel	Traveler's diarrhea	16/118 (13.6)	7/287 (2.4%)	Female gender, abdominal pain, prolonged diarrhea, and antibiotic use
Marshall et al. (2007) [19]	Canada	Viral diarrhea	21/89 (23.6)	1/29 (3.4%)	Vomiting during gastroenteritis
Spence et al. (2007) [20]	New Zealand	<i>Campylobacter</i>	86/581 (14.8)	No control	Psychological co-morbidity and lack of rest during gastroenteritis
Hanevik et al. (2009) [21]	Norway	<i>Giardia</i>	66/82 (80.5)	No control	Not evaluated
Zanini et al. (2012) [22]	Italy	<i>Norovirus</i>	40/186 (21.5)	3/198 (1.5%)	Not evaluated
Cremon et al. (2014) [23]	Italy	<i>Salmonella enterica subsp. enterica</i> serovar <i>Typhi</i>	75/204 (36.8)	44/189 (23.3%)	Anxiety and functional dyspepsia
Persson et al. (2015) [24]	Norway	<i>Giardia</i>	224/724 (32)	96/847 (11.4%)	Not evaluated
Wadhwa et al. (2016) [25]	USA	<i>Clostridium difficile</i>	52/205 (25)	No control	Longer infection duration, current anxiety, and higher BMI
Andresen et al. (2016) [26]	Germany	Shiga-like toxin-producing <i>E. coli</i>	98/389 (25.3)	No control	Higher somatization and anxiety scores
Rahman and Ghoshal (2018) [27]	Bangladesh	<i>E. coli</i> , <i>Campylobacter</i> , <i>V. Cholerae</i> , <i>Salmonella</i> , <i>Shigella</i> , <i>Aeromonas</i>	57/345 (16.5)	9/345 (2.6%)	Dyspeptic symptoms, continuing bowel dysfunction, and weight loss
Current study (2019) [28]	India	<i>E. coli</i> , <i>Campylobacter</i> , <i>V. Cholerae</i> , <i>Salmonella</i> , <i>Shigella</i> , <i>Y. enterocolitica</i>	35/136 (25.7)	No control	Younger age, prolonged duration of diarrhea, and abdominal cramps

IBS irritable bowel syndrome, HS healthy subject, FBD functional bowel disease, BMI body mass index

occludens toxin [Zot], accessory cholera enterotoxin [Ace], hemolysin/cytolysin toxin), which are found almost in all strains of *V. cholerae*, and several more toxins (Shiga-like

toxin, ST, new cholera toxin, sodium channel inhibitor, and thermostable direct hemolysin) are found in a small proportion of patients [32, 34]. The classical mechanisms of action of CT

are cAMP-dependent ion transport activation leading to increased Cl^- secretion in crypt cells and reduced NaCl -coupled absorption in the villous cells. There are alternate or additional mechanisms by which CT stimulates intestinal secretions. These are (a) prostaglandin secretions, (b) activation of the enteric nervous system (ENS), (c) alteration in epithelial barrier function, (d) intestinal inflammatory response, and (e) immunomodulation. A number of studies suggest that prostaglandins (PGE1 and PGE2) and platelet-activating factor (PAF) are involved in the pathogenesis of intestinal secretion caused by CT [35–39]. CT stimulates the epithelial cell phospholipase to produce prostaglandins (PGs). PG production at the submucosal compartment has also been implicated in the pathogenesis of diarrhea in cholera [34]. CT causes activation of ENS. There are number of evidences that suggest the role of CT in ENS activation: (i) a variety of ganglionic or neurotransmitter blockers inhibit the CT-induced secretion [40–42]. 5-Hydroxytryptamine (5-HT) receptor antagonists markedly diminish the CT-induced secretion [43]. Vasoactive intestinal peptide (VIP) release from intestine in patients with cholera can be blocked by tetratoxin [44]. It is estimated that 60% of the effect of CT on intestinal fluid secretion may result from activation of ENS [41]. CT stimulates ENS by stimulating the 5-HT release by Enterochromaffin (EC) cells and/or by stimulating the smooth muscle cell activity of the small intestine. 5-HT causes intestinal secretions in several ways: (i) directly stimulates ENS leading to VIP secretion, and (ii) causes PG release that either stimulates transport function directly or activates the ENS [32]. CT has immunomodulatory effects. It is a potent oral immunogen having a variety of effects on the cells of the immune system. It stimulates interleukin (IL)-1 production and enhances antigen presentation by macrophages [45]. It promotes B cell isotype differentiation [46] and inhibits Th1 cells [47]. CT can alter the intestinal epithelial barrier function [32]. Altered intestinal epithelial barrier function may also result from zot toxin [48]. CT induces a modest inflammatory response. Duodenal biopsy of cholera patients demonstrated modest inflammation [49]. It stimulates the production of IL-6, which then activates the generation of PGs and leukotrienes that stimulate intestinal secretion. High levels of lactoferrin have been found in stool samples of healthy volunteers infected with *V. cholerae* from which toxin genes have been deleted or mutated [50]. Many of the above-mentioned mechanisms have been implicated in the pathogenesis of PI-IBS. However, the mechanism of cholera in acute stage has been studied extensively; pathophysiological aspects of its long-term consequences such as the development of PI-IBS need to be studied further.

Micro-organic basis of PI-IBS

Although studies on PI-IBS provide us probably the best evidence for a micro-organic basis of this hitherto enigmatic

disorder, the pathophysiological mechanisms of PI-IBS have not been studied much. Current evidence suggests that PI-IBS is complex and is probably a multifactorial disease involving visceral hypersensitivity, gut dysmotility, microbial dysbiosis, altered intestinal permeability, immune dysregulation, alteration in neuroendocrine pathways, and genetic factors [1, 51, 52].

Gut microbiota dysbiosis is associated with acute gastroenteritis, its recovery, and development of PI-IBS [53, 54]. A few studies suggest that susceptibility to GI infection can be increased due to an imbalanced gut microbiota [54, 55]. This may result from a complex interaction between the microbiota and host immune system, which reduces the response to GI infections [56]. Gut microbiota analysis identified potential etiological factors in acute gastroenteritis [57]. In a prospective study, the abattoir workers developing *Campylobacter jejuni* infection had increased levels of Bacteroides species prior to infection compared with those who did not develop an infection. The uninfected person had significantly higher levels of uncultured Clostridiales [58]. In short-term, altered gut microbiota was found after acute GI infection [59, 60]. Travelers who develop infectious diarrhea have low levels of Bacteroides [61].

PI-IBS patients have the inability to restore the microbial ecosystem after an attack of acute gastroenteritis [1]. The significant difference in gut microbiota profile of PI-IBS patients compared with healthy control has been found [62, 63]. A recent study demonstrated significant difference in gut microbiota profile between the healthy subjects and the patients with PI-IBS (increase in Bacteroides species and decrease in number of uncultured Clostridiales) [62].

In a recent prospective household survey in Bangladesh, the authors found that abnormal gut microbial communities are host factors for susceptibility to *V. cholerae* infection. Susceptibility to cholera was associated with a decreased level of Bacteroides [64]. A study involving time-series metagenomic analysis of fecal samples collected during the acute diarrhea and recovery phases of cholera in a cohort of Bangladeshi adults found that recovery was characterized by a pattern of accumulation of bacterial taxa that showed similarities to the pattern of assembly/maturation of the gut microbiota in healthy Bangladeshi children [65]. When an artificial community composed of human gut bacterial species that directly correlated with recovery from cholera in adults and were indicative of normal microbiota in healthy Bangladeshi children was introduced into gnotobiotic mice, *Ruminococcus obeum* (*R. obeum*) exhibited a consistent increase in its relative abundance upon *V. cholerae* infection of the mice. Follow up analyses, including mono- and co-colonization studies, established that *R. obeum* restricts *V. cholerae* colonization [65]. The study demonstrated that during diarrheal or recovery phase (up to 2 months), increase in the relative abundance of species of the genera Bacteroides, Prevotella, Ruminococcus/Blautia, and Faecalibacterium strongly correlated with a shift

in community structure towards a healthy adult configuration [65]. It is well-known that IBS in general and PI-IBS, in particular, are associated with gut microbial dysbiosis, which is also associated with susceptibility and occurrence of cholera suggesting a link between PI-IBS and cholera.

There is a complex interaction between gut microbiota and host gene expression and immune response, which are bidirectional [1, 62, 63]. Current evidence suggests that gut microbial composition affects the susceptibility to GI infection by invasive bacteria such as *Campylobacter jejuni* [58] as well as by *V. cholerae* [64]. Dysbiosis has been found after acute gastroenteritis by invasive bacteria as well as by *V. cholerae* at least in short-term [65]. However, a number of studies reported the occurrence of PI-IBS after acute gastroenteritis caused by invasive bacteria. Some of the recent studies demonstrated that PI-IBS occurred after cholera and its frequency was comparable to other invasive bacteria [27, 28]. Although the gut microbiota alteration has been studied in PI-IBS after acute gastroenteritis caused by invasive bacteria [62, 63], currently, such alteration is unknown among PI-IBS patients resulting from cholera.

Incidence of PI-IBS and its relationship with the hygiene hypothesis

The frequency of acute gastroenteritis in the tropics including India and Bangladesh is high [66]. On the other hand, in most of the studies, the prevalence of IBS among the general adult population in India and Bangladesh has been found to be between 4% and 8% [67], which is lower compared with the global prevalence of 11.2% [68]. The reason for the low prevalence of IBS in spite of the high prevalence of acute gastroenteritis in these countries remains enigmatic. The hygiene hypothesis was put forward to explain this enigma, which states immune tolerance develops if an individual is exposed repeatedly to the microorganism in early life. Though subsequent infection in later life induces intestinal immune response quickly and effectively resulting in its clearance, the immune activation is switched off rapidly due to a strong T-regulatory function [69]. However, the two current studies from Bangladesh and India on PI-IBS suggest that the frequency of IBS after acute gastroenteritis was 16.5% and 26%, respectively [27, 28]. These frequencies are somewhat comparable or higher than those reported in the other studies from the temperate countries (Table 1). Do these findings from these two countries contradict the hygiene hypothesis? To reconcile these findings in the context of the hygiene hypothesis, it is important to review the risk factors for the development of PI-IBS [70].

Risk factors for PI-IBS

Several risk factors for the development of PI-IBS following an attack of acute gastroenteritis have been identified (Table 1). These include demographic parameters (e.g. young age, female gender, and smoking), gastroenteritis-related variables (e.g. long duration diarrhea of more than 1 week, abdominal pain, blood in the stool, and antibiotic treatment), and psychological factors (anxiety, depression, somatization, neuroticism, adverse life events in preceding year, hypochondriasis, extroversion, negative illness beliefs, and history of stress and sleep disturbance) [70] (Fig. 1).

A risk score was developed and validated by Thabane et al. [71]. The model used demographic factors, psychological stress, and clinical characteristics of acute gastroenteritis that included gender, diarrheal illness lasting more than 7 days, peak frequency of seven or more loose stools per day, and weight loss of at least 10 lbs. This risk score has been validated in the Indian study on PI-IBS published in this issue of the *Journal* [28]. In the original study, three discrete categories were developed for the level of risks; low (score < 42), intermediate (43–68), and high (> 69). In the derivation set, 10%, 35%, and 60% of subjects developed PI-IBS from the low, intermediate, and high-risk score group, with no differences from the validation cohort [71]. In the Indian study, with a cutoff value of > 50, the sensitivity and specificity of the risk score were 91.4% and 84.2%, respectively, to predict PI-IBS development at 6 months. These findings suggest that this risk score, which is easy to use, may be clinically useful to guide the therapy and counseling of patients with acute gastroenteritis.

The above-mentioned risk factors for PI-IBS help to reconcile the apparently comparable or greater frequency of PI-IBS in the tropical rather than the temperate world that seems to challenge the hygiene hypothesis (Table 1). Since all the patients in the Indian and the Bangladeshi studies were admitted to the hospitals, expectedly had severe gastroenteritis, and a large proportion received antibiotics, there is a possibility of over-estimation of the frequency of PI-IBS in these series. More studies, therefore, are needed including consecutive patients with acute gastroenteritis attending outpatient clinics without specific risk factors for PI-IBS to know its true frequency after an attack of acute gastroenteritis in tropical countries.

Tropical sprue vs. PI-IBS

Tropical sprue (TS) has been reported from southern India almost seven decades ago [72]. It has also been reported from Vietnam, Burma, and eastern India (present-day Bangladesh) [73, 74] and recently from the USA [75]. Several studies on malabsorption have demonstrated that TS is still the leading cause of sporadic malabsorption in India [76–78]. Epidemic TS or post-infectious malabsorption (PI-MAS) has

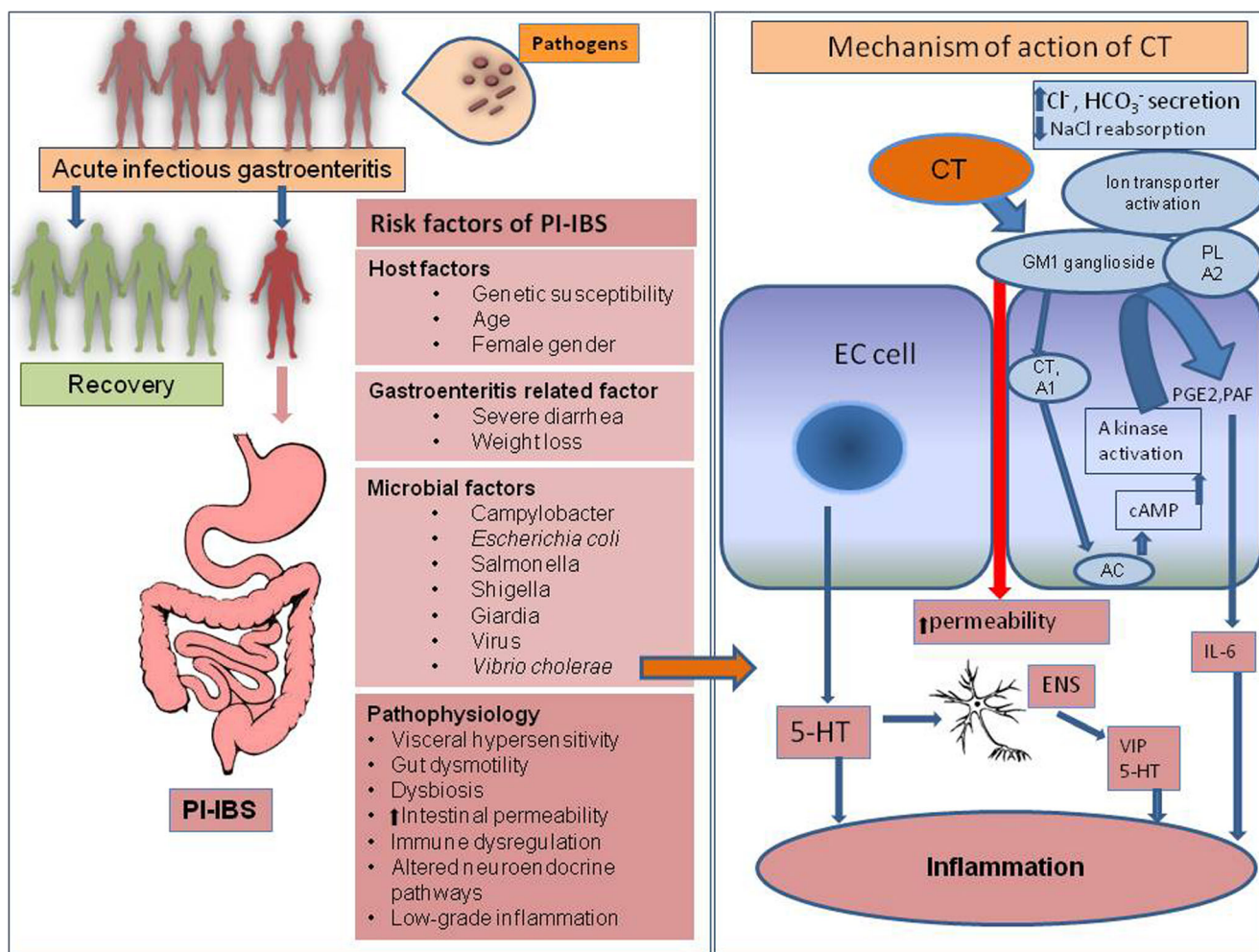


Fig. 1 Risk factors and pathophysiology of post-infection irritable bowel syndrome (PI-IBS) and mechanism of action of cholera toxin. *CT* cholera toxin, *AC* adenylate cyclase, *PL-A2* phospholipase A2, *PGE2* prostaglandin E2, *PAF* platelet activating factor, *5-HT* 5-hydroxytryptamine, *ENS*

enteric nervous system, *VIP* vasoactive intestinal polypeptide, *EC cell* enterochromaffin cell, *IL-6* interleukin-6, *cAMP* cyclic adenosine monophosphate

epidemiologic, clinical, and pathophysiological similarities with PI-IBS [51, 73, 74]. Both the disorders occur after an attack of acute gastroenteritis. The predominant clinical features are chronic diarrhea in both the disorders. Abnormal intestinal permeability, small intestinal bacterial overgrowth, abnormal gut motility, and neurohormonal dysfunction have been found both in PI-MAS and PI-IBS [51, 73, 74]. Since PI-IBS is a symptom-based diagnosis by current Rome criteria [1], it is possible that some patients with PI-MAS without overt malnutrition are diagnosed as PI-IBS by symptom-based criteria. However, none of the previous studies on PI-IBS excluded the PI-MAS by tests of mucosal malabsorption except one recent study from Bangladesh. In this study, the authors found that about 9% of the patients fulfilling the Rome III criteria of PI-IBS had at least two abnormal mucosal absorption tests, (i) D-xylose hydrogen breath test, (ii) fecal fat microscopy using Sudan III stain, and (iii) histopathological examination of duodenal biopsies obtained endoscopically,

suggesting these patients had PI-MAS or tropical sprue. In this study, 22% of the 23 subjects who were investigated for malabsorption had at least one test abnormal [27]. This study suggests that the patients who develop continued bowel symptoms after acute gastroenteritis need to be investigated for mucosal malabsorption even though they might have fulfilled the Rome criteria for IBS; this highlights the importance of further study on this issue on larger number of patients.

Summary and future direction

Two recent studies from India and Bangladesh, including one published in this issue of the *Journal*, have demonstrated that infection with *V. cholerae* is a risk factor for the development PI-IBS. The frequency of PI-IBS following acute gastroenteritis in these two countries is comparable or higher than that in the other studies published previously. However, further

prospective case-control studies are needed to know the risk of PI-IBS in less severe forms of acute gastroenteritis caused by different pathogens including *V. cholerae* in tropical countries. Similarly, further validation of the risk score is needed in less severe forms of acute gastroenteritis. Although a number of mechanistic studies including gut microbiota have been conducted in acute cholera, how this infection leads to long-term consequences including PI-IBS requires further studies. About 9% of the patients who are diagnosed as PI-IBS actually had PI-MAS in the Bangladeshi study, and this issue has not been investigated in the Indian study. More studies are needed on this issue. Clinical and economic consequences of acute gastroenteritis among travelers from temperate to tropical countries are also important issues that require further study.

Compliance with ethical standards

Conflict of interest UCG, and MMR declare that they have no conflict of interest.

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