REVIEW

The essential role of TAZ in normal tissue homeostasis

Mi Gyeong Jeong¹ · Hyo Kyeong Kim¹ · Eun Sook Hwang¹

Received: 15 January 2021 / Accepted: 14 March 2021 / Published online: 26 March 2021 © The Author(s) 2021

Abstract Transcriptional coactivator with PDZ-binding motif (TAZ) has been extensively characterized in organ development, tissue regeneration, and tumor progression. In particular, TAZ functions as a Hippo mediator that regulates organ size, tumor growth and migration. It is highly expressed in various types of human cancer, and has been reported to be associated with tumor metastasis and poor outcomes in cancer patients, suggesting that TAZ is an oncogenic regulator. Yes-associated protein (YAP) has 60% similarity in amino acid sequence to TAZ and plays redundant roles with TAZ in the regulation of cell proliferation and migration of cancer cells. Therefore, TAZ and YAP, which are encoded by paralogous genes, are referred to as TAZ/ YAP and are suggested to be functionally equivalent. Despite its similarity to YAP, TAZ can be clearly distinguished from YAP based on its genetic, structural, and functional aspects. In addition, targeting superabundant TAZ can be a promising therapeutic strategy for cancer treatment; however, persistent TAZ inactivation may cause failure of tissue homeostatic control. This review focuses primarily on TAZ, not YAP, discusses its structural features and physiological functions in the regulation of tissue homeostasis, and provides new insights into the drug development targeting TAZ to control reproductive and musculoskeletal disorders.

Keywords Hippo pathway · TAZ · Tissue homeostasis · Transcriptional control · YAP

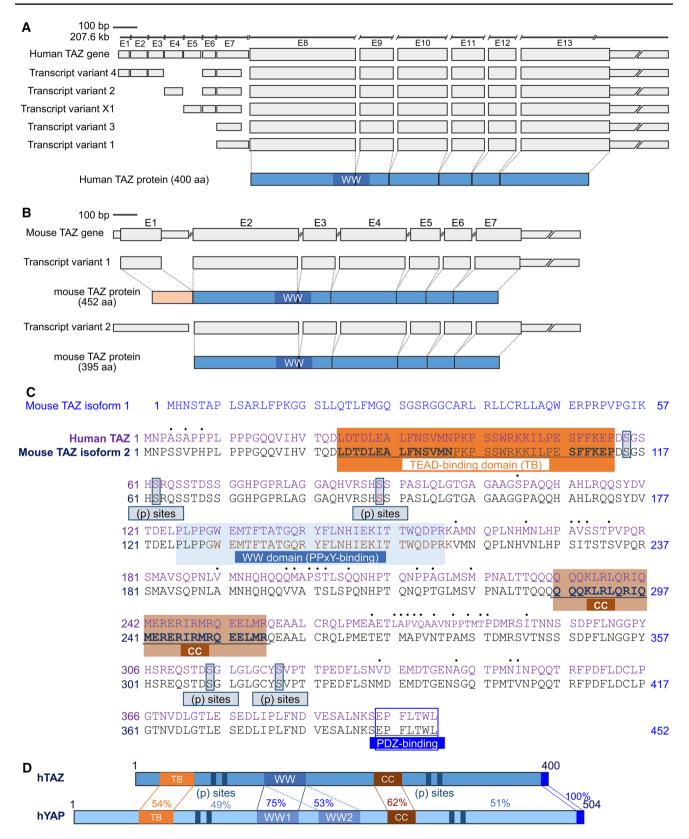
Eun Sook Hwang eshwang@ewha.ac.kr

¹ College of Pharmacy and Graduate School of Pharmaceutical Sciences, Ewha Womans University, C206 Science building, 52 Ewhayeodae-Gil, Seodaemun-Gu, Seoul 03760, Korea

Introduction

Transcriptional coactivator with PDZ-binding motif (TAZ) has been identified as a phosphoprotein that interacts with 14-3-3 protein and its activity is controlled by nuclearcytosol localization in a phosphorylation-dependent manner (Kanai et al. 2000). TAZ shares structural and functional similarities with yes-associated protein (YAP) (Macias et al. 1996; Piccolo et al. 2014). TAZ and YAP are phosphorylated and trapped in the cytoplasm via interaction with 14-3-3 or undergo proteasomal degradation through β-TrCP-mediated ubiquitination in response to Hippo signaling (Tian et al. 2007; Liu et al. 2010). The upstream mammalian Ste20like kinases 1 and 2 (MST1/2) trigger the phosphorylation and activation of large tumor suppressor kinases 1 and 2 (LATS1/2) in the Hippo signaling cascade, which in turn phosphorylate TAZ/YAP and control their cytoplasmic retention and protein stability (Pan 2010; Yu and Guan 2013; Piccolo et al. 2014). When the Hippo signaling pathway is inactive, TAZ/YAP are dephosphorylated and localized in the nucleus. TAZ and YAP are associated with many transcription factors that control various physiological cellular events, including cell proliferation, differentiation, migration, apoptosis, and senescence, and thereby result in transcriptional activation of target genes (Varelas et al. 2008; Di Palma et al. 2009; Jeong et al. 2010, 2017; Wang et al. 2016; Kim et al. 2019, 2020). In particular, TAZ and YAP interact with transcription factors, in particular, transcriptional enhanced associate domain (TEAD) family and enhance the transcription of multiple targets involved in tumorigenesis, thereby affecting the self-renewal of stem cells, tumor progression, metastasis, and drug resistance (Chan et al. 2009; Zanconato et al. 2015, 2016; Lin et al. 2017). Nuclear localization and activation of TAZ and YAP are also regulated by mechanical signals, such as shear stress, cell shape, and





extracellular rigidity (Dupont et al. 2011; Dupont 2016; Pocaterra et al. 2020). Although TAZ and YAP are paralogs with functional redundancy and tumor-promoting effects

(Kim 2019), it is clear that TAZ and YAP play distinctively essential roles in normal tissue development and homeostasis (Yu et al. 2015; Reggiani et al. 2020). We clarify the **∢Fig. 1** Structural features of TAZ. **a** Genome structure of human TAZ. Five different transcript variants of TAZ (NM 015472.6, NM 001168278.3, NM_001168280.3, NM_001348362.2, and XM_017006122.1) are produced from human chromosome 3 (GRCh38.p13) and are encoded as the same human TAZ comprising of 400 amino acids. b Mouse chromosome 3 (GRCm39) comprises of TAZ exons and introns. Two different transcript variants (NM_00168281.1 and NM_00168281.1) are generated and encoded as two isoforms of TAZ; isoform 1, 452 aa and isoform 2, 395 aa. c Amino acid alignment between human and mouse TAZ. Mouse TAZ isoform 1 contains additional 57 aa at the N-terminus and has no similarity with human TAZ. d Schematic structure of human and mouse TAZ proteins. The percent identity for a given sequence is presented. TB, TEAD-binding domain; WW, W-containing domain; CC, coiledcoil domain; and (p) sites, phosphorylation sites

genetic and protein structural features of TAZ and further highlight its biological functions in normal tissue differentiation from stem cells and in the regulation of tissue homeostasis.

Structural features of TAZ

TAZ gene is officially named as WW domain-containing transcriptional coregulatory 1 (WWTR1) gene, but it is more extensively and commonly used than WWTR1, and is different from *tafazzin (TAZ)*. The TAZ gene, a Hippo mediator, is highly conserved among divergent species, and the human TAZ gene has orthologs in 288 organisms, including zebrafish, frog, mouse, rat, chicken, cow, dog, pig, and chimpanzee. The 207.6 kb portion of the human TAZ gene located on chromosome 3 contains 13 exons and produces four different TAZ transcript variants, which encode the same 400 amino acid (aa) protein (Fig. 1a). In mouse, 7 exons transcribe two different transcript variants encoding two TAZ isoforms composed of 452 and 395 amino acids. The mouse TAZ isoform 1 is 100% homologous to mouse TAZ isoform 2 and additionally contains 57 aa at the N-terminal, which is not found in human TAZ. The human TAZ protein shares 91% identity with the mouse TAZ in the entire amino acid sequence, and the WW domain is completely identical (Fig. 1b). The WW domain is a modular protein domain composed of 40 aa, which mediates specific interactions with proteins containing P-rich motifs such as PPxY, LPxY, phosphor-(S/T)P, and PRR motifs (Hu et al. 2004). Many transcription factors, including Runt-related transcription factors (RUNXs), peroxisome proliferator-activated receptor γ (PPAR γ), TEADs, SMADs, ErbB-4, and p73 are known to interact with the WW domain in TAZ or YAP (Strano et al. 2001; Ferrigno et al. 2002; Komuro et al. 2003; Zhao et al. 2009). Since TAZ contains a single WW domain unlike two WW domains in YAP, its interaction with TEAD is structurally different from TEAD-YAP (Kaan et al. 2017; Reggiani et al. 2020). TAZ can be distinguished from YAP by its interactions with many P-rich proteins and functions within the cells. In addition, the coiled-coil domain and PDZ-binding motif are within a larger transcriptional regulatory regions at the C-terminal domain of TAZ and serve as additional protein–protein interaction domains (Kanai et al. 2000; Hong et al. 2005).

TAZ is essential for self-renewal and survival of stem cells

Stem cells are defined as undifferentiated cells capable of producing certain specialized cells and self-renewing progeny cells, and are classified as fetal, embryonic, and adult stem cells. TAZ is required to maintain the self-renewal and pluripotency of embryonic stem cells, since its deficiency in human embryonic stem cells leads to differentiation into neuroectodermal lineage (Varelas et al. 2008). TAZ also plays key roles in the expansion, self-renewal, and maintenance of stemness of tissue-specific adult stem cells. TAZ associates with snail/slug and cooperatively controls the self-renewal and osteoblastogenesis of adult skeletal stem cells (Tang et al. 2016; Tang and Weiss 2017). Exogenous TAZ expression reprograms primary differentiated mouse cells into tissue-specific stem cell or progenitor cell state, indicating the importance of TAZ for maintaining stemness of adult stem cells (Panciera et al. 2016). In addition, dysregulated TAZ expression in stem cells causes uncontrolled self-renewal and cell expansion, resulting in cancer growth and sustained survival of cancer stem cells (Bartucci et al. 2015; Mohamed et al. 2016; Elaimy et al. 2018). TAZ is crucial for maintaining stem cell population and the potency of both embryonic and adult stem cells (Fig. 2).

TAZ modulates mesenchymal stem cell differentiation into bone, adipose, and muscle

Many transcription factors that contain P-rich motifs have been reported to interact with TAZ during the execution of many developmental programs (Hong and Yaffe 2006). TAZ associates with RUNX2 and strongly activates RUNX2driven gene transcription, resulting in enhanced osteoblast differentiation (Hong et al. 2005; Long 2011). TAZ also interacts with PPARy but markedly suppresses PPARydriven adipogenic and lipogenic gene expression, whereas TAZ deletion in mesenchymal stem cells increasingly drives adipocyte differentiation (Hong et al. 2005). TAZ is thus important for reprogramming of stem cell lineage commitment. While exogenous TAZ overexpression increased bone mineral density in vivo, TAZ deletion impaired osteogenic differentiation, but enhanced adipogenic differentiation of human adipose tissue-derived stem cells (Yang et al. 2013; Zhu et al. 2018). In addition, TAZ induced myogenic differentiation through interaction with MyoD and activation of myogenic gene transcription (Jeong et al. 2010) (Fig. 2).

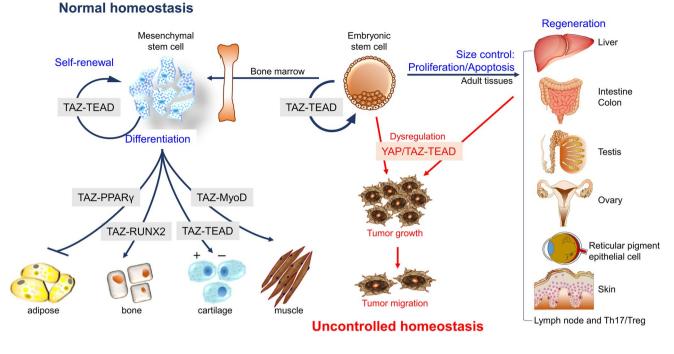


Fig. 2 Control of tissue homeostasis by TAZ. Under normal homeostasis conditions, TAZ functions in the self-renewal of embryonic and mesenchymal stem cells through TAZ-TEAD complex formation. Differentiation of mesenchymal stem cells into different lineages is controlled by the association of TAZ with several transcription factors. Development of adult tissues and injury-induced tissue regeneration requires TAZ expression, which regulates organ size by controlling both cell proliferation and apoptosis. However, uncontrolled homeostatic imbalance may lead to irreversible changes in cells and tissues and subsequently to malignant tumor development. Overexpression or hyperactivation of YAP and TAZ accelerates tumor growth and metastasis by inducing dysregulation in cell proliferation, migration, and survival of stem cells

YAP has also been reported to regulate osteo-adipogenic differentiation from human mesenchymal stem cells (Lorthongpanich et al. 2019) and myogenic differentiation (Watt et al. 2010; Chen et al. 2017). Although TAZ and YAP promote proliferation of myoblasts, TAZ, but not YAP, stimulates late myogenic differentiation, suggesting a distinctive role of TAZ from YAP in skeletal muscle stem cell function (Sun et al. 2017). TAZ is also essential for inducing cell proliferation and chondrocyte marker expression during chondrocyte differentiation; however, hyperactivation of YAP and TAZ inhibit chondrocyte maturation by repressing the transcription factor SOX9 (Deng et al. 2016; Goto et al. 2018). It remains to be clarified whether YAP and TAZ play redundant roles during chondrocyte differentiation and whether TAZ promotes or represses chondrocyte differentiation in vivo.

TAZ promotes injury-induced liver regeneration and intestinal regeneration

The Hippo signaling pathway plays critical roles in liver size control and tumorigenesis through phosphorylation and activation of the large tumor suppressor kinases 1/2 (Lats1/2) and inhibition of YAP activation. Any defects in Hippo signaling molecules, including YAP, induce aberrant

hepatomegaly and tumorigenesis (Dong et al. 2007; Zhou et al. 2009; Lu et al. 2010; Zhao et al. 2011). Although YAP deletion fails to restore liver mass and the expression of connective tissue growth factor (CTGF), there is no clear evidence that TAZ plays a redundant role in liver regeneration (Konishi et al. 2018; Lu et al. 2018). Recently, it was clarified that liver-specific TAZ deletion delayed liver regeneration and enhanced cell death after partial hepatectomy (Kim et al. 2019). TAZ is thus believed to stimulate liver regeneration through IL-6-mediated hepatocyte proliferation and inhibition of cell death after injury. Furthermore, TAZ is essential for intestinal regeneration following gamma-irradiation, and its overexpression is associated with intestinal tumor formation (Byun et al. 2017). Intestinal TAZ function is highly correlated with important roles in the Hippo pathway and with YAP in the control of colonic epithelial regeneration after injury (Hong et al. 2016; Yui et al. 2018). Therefore, TAZ expression as well as its activation are essential factors for tissue regeneration.

TAZ expression is required for the maintenance of testicular structure and function

TAZ is indispensable for quantitative regulation and functional maintenance of embryonic and adult stem cells and contributes to cellular reprogramming and rejuvenation in many types of tissues. However, the functional role of TAZ is relatively lacking in reproductive organs and fertility. Jeong et al. (2017) first reported that TAZ was expressed in testicular cells and its deletion caused structural abnormality of the testicles and functional defects in fertility. TAZ deletion further facilitated testicular aging and apoptotic death of spermatogenic stem cells (Jeong et al. 2017). More recently, TAZ was found to interact with nuclear receptor 4A1 in Leydig cells and negatively modulate the expression of steroidogenic enzymes, resulting in decreased testosterone production by Leydig cells (Shin et al. 2020). In addition to the TAZ functions in the male reproductive system, TAZ has been shown to play crucial roles in oogenesis and fertilization in zebrafish and murine ovarian folliculogenesis (Dingare et al. 2018; Xia et al. 2019; Yi et al. 2019; Bernabe et al. 2020). Consistently, LATS1/2 and YAP in the Hippo pathway have been demonstrated to be essential for the proliferation of ovarian granulosa cells and maintaining normal follicle development in the female reproductive system (Plewes et al. 2019; Tsoi et al. 2019).

TAZ modulates normal tissue homeostasis in several tissues

TAZ is expressed in most tissues and affects their specific gene expression as a mediator of the Hippo signaling pathway. TAZ deletion impairs the optic vesicle progenitor cells to form retinal pigment epithelial cells, whereas ectopic TAZ expression enhances ectopic pigmentation in optic vesicle progenitor cells (Miesfeld et al. 2015). TAZ is necessary for eye development through determination of retinal pigment epithelial cell fate. Furthermore, TAZ modulates skin homeostasis. TAZ is expressed in the nucleus of the basal layer cells of the skin and is elevated in response to wound healing. TAZ is normally localized to the cytoplasm in the dermis, but is distributed in both the nucleus and cytoplasm at 1 day after skin damage, and its deficiency markedly delays skin wound healing (Lee et al. 2014). Skin-specific TAZ deletion slows the growth of basal layer cells, leading to hair loss and inhibition of skin regeneration (Elbediwy et al. 2016a, 2016b). The association of TAZ with TEAD promotes proliferation of skin cells but the inhibition of their interaction increases the expression of kruppel-like factor 4 (KLF4), thereby increasing keratinocyte differentiation. TEAD and KLF4 regulate the activity of each other through modulation of TAZ interaction with TEAD and other transcription factors during keratinocyte differentiation (Yuan et al. 2020). Therefore, TAZ contributes in maintaining epidermal and dermal cell populations during development and also regulates skin homeostasis during wound healing. Moreover, TAZ has been suggested to promote Th17 cell differentiation and inhibit Treg cell development in immune system (Geng et al. 2017), suggesting a potential role of TAZ in autoimmune diseases. Inhibition of dysregulated TAZ expression in arthritic patient induces Treg cell-mediated anti-inflammatory effect (Du et al. 2020). TAZ also regulates lymph node differentiation, in particular, commitment and maturation of fibroblastic reticular cells (Choi et al. 2020). The fine-tuning of TAZ expression and its activity is essential for maintaining normal tissue homeostasis and limiting cancer incidence.

Small molecules that stimulate TAZ activity are promising candidates for disease treatment

TAZ is believed to be a promising anti-cancer target because it promotes cancer cell proliferation, survival, and drug resistance. Various kinds of small molecules targeting TAZ, such as those that block the nuclear localization of TAZ, inhibit TAZ-TEAD complex formation, and suppress TAZ-TEAD target genes, are being discovered as anticancer agents. Since MST1/2 and LATS1/2 are crucial for the regulation of TAZ phosphorylation and activation, small molecules that regulate their activities in biological functions have been identified. Compound 9E1, XMU-MP-1, and neratinib inhibit MST1/2 and thus suppress the activation of LATS1/2 and subsequent YAP/TAZ-mediated cellular proliferation and tissue regeneration (Anand et al. 2009; Fan et al. 2016; Ardestani et al. 2019a). However, C19 stimulates MST-induced LATS1/2 phosphorylation and YAP/ TAZ inactivation, suggesting an anti-tumor potential (Basu et al. 2014). Dobutamine has been identified as an inhibitor of YAP-mediated gene transcription (Bao et al. 2011). Dasatinib, statins, and pazopanib are known to inhibit the nuclear localization of TAZ and decrease cell proliferation and chemoresistance of breast cancer cells (Oku et al. 2015). In addition, digitoxin, verteporfin, and flufenamic acid inhibit YAP-TEAD interaction and suppress cancer growth (Liu-Chittenden et al. 2012; Sudol et al. 2012; Pobbati et al. 2015; Pobbati and Hong 2020). Although YAP inhibition is beneficial for treating many cancers, YAP activation through the inhibition of Hippo signaling is also required for pancreatic β -cell regeneration (Ardestani et al. 2019a, 2019b). Furthermore, TAZ activators are also helpful for treating diseases, including osteoporosis, diabetes, and muscular atrophy. A few compounds have been identified to enhance the nuclear localization of TAZ. Ethacridine enhances nuclear retention of TAZ and inhibits adipogenic gene expression (Kawano et al. 2015). The TAZ modulator, TM-25659, stimulates TAZ-RUNX2 interaction and the subsequent osteogenic differentiation and suppresses adipogenic differentiation by potentiating TAZ-PPARy complex formation (Jang et al. 2012; Zhu et al. 2018). TM-25659 protects against obesity and diabetic hyperglycemia induced by high-fat diet in an animal model (Jung et al. 2015). TM-53

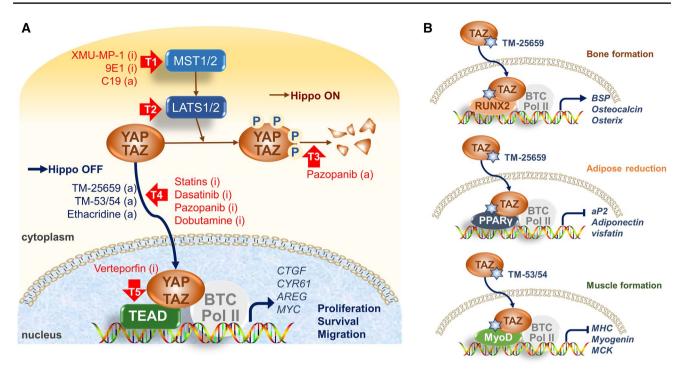


Fig. 3 Identification of small molecules targeting the Hippo-TAZ/YAP pathway. **a** The Hippo pathway can be regulated at several target points (T1 through T5). The Hippo signaling molecules, MST1/2 (T1) and LATS1/2 (T2), are directly and indirectly affected by compounds such as XMU-MP-1, 9E1, and C19, which affect the activation of TAZ and YAP. YAP and TAZ undergo proteasomal degradation (T3), which is promoted by pazopanib. Nuclear localization of TAZ and YAP (T4) is negatively regulated by statins, dasatinib, pazopanib, and dobutamine, but positively regulated by TM-25659, TM-53/54, and ethacridine. Then YAP/TAZ-TEAD association (T5) is repressed by verteporfin. Pharma-cological regulation of the Hippo-TAZ/YAP pathway affects cell proliferation, differentiation, survival, and migration. **b** Small molecules such as TM-25659 and TM-53/54 promote nuclear localization of TAZ and specifically modulate the association of TAZ with RUNX2, PPAR γ , or MyoD and subsequent cell specific gene transcription for bone, adipose, and muscle, respectively

and TM-54 increased nuclear localization of TAZ and potentiated MyoD-induced myoblast and muscle differentiation (Park et al. 2014) (Fig. 3). These findings suggest that TAZ activators have beneficial effects in the treatment of metabolic and musculoskeletal disorders.

Conclusions and future perspectives

Since TAZ contributes to the self-renewal and potency of stem cells associated with the acquisition of many cancer traits, targeting TAZ is considered to be more efficient in the control of several types of cancers. It is noteworthy that the development of anticancer drugs targeting TAZ is promising for treating certain types of cancers, including skin, breast, and lung cancer (Pobbati and Hong 2020). However, TAZ intrinsically serves as a physiological regulator for organ development, size control, and tissue repair and regeneration. TAZ positively or negatively regulates gene transcription through interaction with many transcription factors and modulates lineage commitment during homeostasis of many types of tissues, including the musculoskeletal system, adipose tissue, liver, digestive system, reproductive system, and skin. It seems that pharmacological activation of TAZ is required for restoring normal tissue homeostasis by increasing stem cell activity and cell lineage commitment. Considering the complex function of TAZ in cancer cells and normal tissues, including stem cells, while complete inhibition of TAZ may be effective in the treatment of cancer, there is a risk of disrupting normal tissues homeostasis. Therefore, pharmacological development of TAZ inhibitors for cancer treatment deserves a great deal of attention with respect to prediction of toxicity and side effects on tissue homeostasis. It is indispensable to develop selective inhibitors or activators that target specific binding sites between TAZ and its interacting partner, which may not interfere with normal tissue homeostasis and may contribute to desirable therapeutic effects (Pobbati and Rubin 2020).

Acknowledgements This work was supported by grants from the National Research Foundation (2018R1A5A2025286 and 2020R1A2C2004679) funded by the Ministry of Education, Science, and Technology.

Declarations

Conflict of interest The authors declare no conflict of interest.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

References

- Anand R, Maksimoska J, Pagano N, Wong EY, Gimotty PA, Diamond SL, Meggers E, Marmorstein R (2009) Toward the development of a potent and selective organoruthenium mammalian sterile 20 kinase inhibitor. J Med Chem 52:1602–1611. https:// doi.org/10.1021/jm8005806
- Ardestani A, Li S, Annamalai K, Lupse B, Geravandi S, Dobrowolski A, Yu S, Zhu S, Baguley TD, Surakattula M, Oetjen J, Hauberg-Lotte L, Herranz R, Awal S, Altenhofen D, Nguyen-Tran V, Joseph S, Schultz PG, Chatterjee AK, Rogers N, Tremblay MS, Shen W, Maedler K (2019a) Neratinib protects pancreatic beta cells in diabetes. Nat Commun 10:5015. https://doi.org/10.1038/ s41467-019-12880-5
- Ardestani A, Tremblay MS, Shen W, Maedler K (2019b) Neratinib is an MST1 inhibitor and restores pancreatic beta-cells in diabetes. Cell Death Discov 5:149. https://doi.org/10.1038/ s41420-019-0232-0
- Bao Y, Nakagawa K, Yang Z, Ikeda M, Withanage K, Ishigami-Yuasa M, Okuno Y, Hata S, Nishina H, Hata Y (2011) A cell-based assay to screen stimulators of the hippo pathway reveals the inhibitory effect of dobutamine on the YAP-dependent gene transcription. J Biochem 150:199–208. https://doi.org/10.1093/jb/mvr063
- Bartucci M, Dattilo R, Moriconi C, Pagliuca A, Mottolese M, Federici G, Benedetto AD, Todaro M, Stassi G, Sperati F, Amabile MI, Pilozzi E, Patrizii M, Biffoni M, Maugeri-Sacca M, Piccolo S, De Maria R (2015) TAZ is required for metastatic activity and chemoresistance of breast cancer stem cells. Oncogene 34:681–690. https://doi.org/10.1038/onc.2014.5
- Basu D, Lettan R, Damodaran K, Strellec S, Reyes-Mugica M, Rebbaa A (2014) Identification, mechanism of action, and antitumor activity of a small molecule inhibitor of hippo, TGF-beta, and Wnt signaling pathways. Mol Cancer Ther 13:1457–1467. https://doi.org/10.1158/1535-7163.MCT-13-0918
- Bernabe BP, Woodruff T, Broadbelt LJ, Shea LD (2020) Ligands, receptors, and transcription factors that mediate inter-cellular and intra-cellular communication during ovarian follicle development. Reprod Sci 27:690–703. https://doi.org/10.1007/ s43032-019-00075-8
- Byun MR, Hwang JH, Kim AR, Kim KM, Park JI, Oh HT, Hwang ES, Hong JH (2017) SRC activates TAZ for intestinal tumorigenesis and regeneration. Cancer Lett 410:32–40. https://doi. org/10.1016/j.canlet.2017.09.003
- Chan SW, Lim CJ, Loo LS, Chong YF, Huang C, Hong W (2009) TEADs mediate nuclear retention of TAZ to promote oncogenic

transformation. J Biol Chem 284:14347-14358. https://doi.org/ 10.1074/jbc.M901568200

- Chen TH, Chen CY, Wen HC, Chang CC, Wang HD, Chuu CP, Chang CH (2017) YAP promotes myogenic differentiation via the MEK5-ERK5 pathway. FASEB J 31:2963–2972. https://doi. org/10.1096/fj.201601090R
- Choi SY, Bae H, Jeong SH, Park I, Cho H, Hong SP, Lee DH, Lee CK, Park JS, Suh SH, Choi J, Yang MJ, Jang JY, Onder L, Moon JH, Jeong HS, Adams RH, Kim JM, Ludewig B, Song JH, Lim DS, Koh GY (2020) YAP/TAZ direct commitment and maturation of lymph node fibroblastic reticular cells. Nat Commun 11:519. https://doi.org/10.1038/s41467-020-14293-1
- Deng Y, Wu A, Li P, Li G, Qin L, Song H, Mak KK (2016) Yap1 regulates multiple steps of chondrocyte differentiation during skeletal development and bone repair. Cell Rep 14:2224–2237. https://doi.org/10.1016/j.celrep.2016.02.021
- Di Palma T, D'Andrea B, Liguori GL, Liguoro A, de Cristofaro T, Del Prete D, Pappalardo A, Mascia A, Zannini M (2009) TAZ is a coactivator for Pax8 and TTF-1, two transcription factors involved in thyroid differentiation. Exp Cell Res 315:162–175. https://doi.org/10.1016/j.yexcr.2008.10.016
- Dingare C, Niedzwetzki A, Klemmt PA, Godbersen S, Fuentes R, Mullins MC, Lecaudey V (2018) The Hippo pathway effector Taz is required for cell morphogenesis and fertilization in zebrafish. Development 145:dev167023. https://doi.org/10. 1242/dev.167023
- Dong J, Feldmann G, Huang J, Wu S, Zhang N, Comerford SA, Gayyed MF, Anders RA, Maitra A, Pan D (2007) Elucidation of a universal size-control mechanism in Drosophila and mammals. Cell 130:1120–1133. https://doi.org/10.1016/j.cell.2007. 07.019
- Du YY, Chen ZX, Liu MY, Liu QP, Lin CS, Chu CQ, Xu Q (2020) Leonurine regulates Treg/Th17 balance to attenuate rheumatoid arthritis through inhibition of TAZ expression. Front Immunol 11:556526. https://doi.org/10.3389/fimmu.2020.556526
- Dupont S (2016) Role of YAP/TAZ in cell-matrix adhesion-mediated signalling and mechanotransduction. Exp Cell Res 343:42–53. https://doi.org/10.1016/j.yexcr.2015.10.034
- Dupont S, Morsut L, Aragona M, Enzo E, Giulitti S, Cordenonsi M, Zanconato F, Le Digabel J, Forcato M, Bicciato S, Elvassore N, Piccolo S (2011) Role of YAP/TAZ in mechanotransduction. Nature 474:179–183. https://doi.org/10.1038/nature10137
- Elaimy AL, Guru S, Chang C, Ou J, Amante JJ, Zhu LJ, Goel HL, Mercurio AM (2018) VEGF-neuropilin-2 signaling promotes stem-like traits in breast cancer cells by TAZ-mediated repression of the Rac GAP beta2-chimaerin. Sci Signal 11:eaao6897. https://doi.org/10.1126/scisignal.aao6897
- Elbediwy A, Vincent-Mistiaen ZI, Spencer-Dene B, Stone RK, Boeing S, Wculek SK, Cordero J, Tan EH, Ridgway R, Brunton VG, Sahai E, Gerhardt H, Behrens A, Malanchi I, Sansom OJ, Thompson BJ (2016a) Integrin signalling regulates YAP and TAZ to control skin homeostasis. Development 143:1674–1687. https://doi.org/10.1242/dev.133728
- Elbediwy A, Vincent-Mistiaen ZI, Thompson BJ (2016b) YAP and TAZ in epithelial stem cells: a sensor for cell polarity, mechanical forces and tissue damage. Bioessays 38:644–653. https://doi. org/10.1002/bies.201600037
- Fan F, He Z, Kong LL, Chen Q, Yuan Q, Zhang S, Ye J, Liu H, Sun X, Geng J, Yuan L, Hong L, Xiao C, Zhang W, Sun X, Li Y, Wang P, Huang L, Wu X, Ji Z, Wu Q, Xia NS, Gray NS, Chen L, Yun CH, Deng X, Zhou D (2016) Pharmacological targeting of kinases MST1 and MST2 augments tissue repair and regeneration. Sci Transl Med 8:352ra108. https://doi.org/10.1126/scitranslmed.aaf2304
- Ferrigno O, Lallemand F, Verrecchia F, L'Hoste S, Camonis J, Atfi A, Mauviel A (2002) Yes-associated protein (YAP65) interacts

with Smad7 and potentiates its inhibitory activity against TGFbeta/Smad signaling. Oncogene 21:4879–4884. https://doi.org/ 10.1038/sj.onc.1205623

- Geng J, Yu S, Zhao H, Sun X, Li X, Wang P, Xiong X, Hong L, Xie C, Gao J, Shi Y, Peng J, Johnson RL, Xiao N, Lu L, Han J, Zhou D, Chen L (2017) The transcriptional coactivator TAZ regulates reciprocal differentiation of TH17 cells and Treg cells. Nat Immunol 18:800–812. https://doi.org/10.1038/ni.3748
- Goto H, Nishio M, To Y, Oishi T, Miyachi Y, Maehama T, Nishina H, Akiyama H, Mak TW, Makii Y, Saito T, Yasoda A, Tsumaki N, Suzuki A (2018) Loss of Mob1a/b in mice results in chondrodysplasia due to YAP1/TAZ-TEAD-dependent repression of SOX9. Development 145:dev159244. https://doi.org/10.1242/ dev.159244
- Hong JH, Yaffe MB (2006) TAZ: a beta-catenin-like molecule that regulates mesenchymal stem cell differentiation. Cell Cycle 5:176–179. https://doi.org/10.4161/cc.5.2.2362
- Hong JH, Hwang ES, McManus MT, Amsterdam A, Tian Y, Kalmukova R, Mueller E, Benjamin T, Spiegelman BM, Sharp PA, Hopkins N, Yaffe MB (2005) TAZ, a transcriptional modulator of mesenchymal stem cell differentiation. Science 309:1074– 1078. https://doi.org/10.1126/science.1110955
- Hong AW, Meng Z, Guan KL (2016) The Hippo pathway in intestinal regeneration and disease. Nat Rev Gastroenterol Hepatol 13:324–337. https://doi.org/10.1038/nrgastro.2016.59
- Hu H, Columbus J, Zhang Y, Wu D, Lian L, Yang S, Goodwin J, Luczak C, Carter M, Chen L, James M, Davis R, Sudol M, Rodwell J, Herrero JJ (2004) A map of WW domain family interactions. Proteomics 4:643–655. https://doi.org/10.1002/ pmic.200300632
- Jang EJ, Jeong H, Kang JO, Kim NJ, Kim MS, Choi SH, Yoo SE, Hong JH, Bae MA, Hwang ES (2012) TM-25659 enhances osteogenic differentiation and suppresses adipogenic differentiation by modulating the transcriptional co-activator TAZ. Br J Pharmacol 165:1584–1594. https://doi.org/10.1111/j.1476-5381.2011.01664.x
- Jeong H, Bae S, An SY, Byun MR, Hwang JH, Yaffe MB, Hong JH, Hwang ES (2010) TAZ as a novel enhancer of MyoD-mediated myogenic differentiation. FASEB J 24:3310–3320. https://doi. org/10.1096/fj.09-151324
- Jeong MG, Song H, Shin JH, Jeong H, Kim HK, Hwang ES (2017) Transcriptional coactivator with PDZ-binding motif is required to sustain testicular function on aging. Aging Cell 16:1035– 1042. https://doi.org/10.1111/acel.12631
- Jung JG, Yi SA, Choi SE, Kang Y, Kim TH, Jeon JY, Bae MA, Ahn JH, Jeong H, Hwang ES, Lee KW (2015) TM-25659-induced activation of FGF21 level decreases insulin resistance and inflammation in skeletal muscle via GCN2 pathways. Mol Cells 38:1037–1043. https://doi.org/10.14348/molcells.2015.0100
- Kaan HYK, Chan SW, Tan SKJ, Guo F, Lim CJ, Hong W, Song H (2017) Crystal structure of TAZ-TEAD complex reveals a distinct interaction mode from that of YAP-TEAD complex. Sci Rep 7:2035. https://doi.org/10.1038/s41598-017-02219-9
- Kanai F, Marignani PA, Sarbassova D, Yagi R, Hall RA, Donowitz M, Hisaminato A, Fujiwara T, Ito Y, Cantley LC, Yaffe MB (2000) TAZ: a novel transcriptional co-activator regulated by interactions with 14-3-3 and PDZ domain proteins. EMBO J 19:6778–6791. https://doi.org/10.1093/emboj/19.24.6778
- Kawano S, Maruyama J, Nagashima S, Inami K, Qiu W, Iwasa H, Nakagawa K, Ishigami-Yuasa M, Kagechika H, Nishina H, Hata Y (2015) A cell-based screening for TAZ activators identifies ethacridine, a widely used antiseptic and abortifacient, as a compound that promotes dephosphorylation of TAZ and inhibits adipogenesis in C3H10T1/2 cells. J Biochem 158:413–423. https:// doi.org/10.1093/jb/mvv051

- Kim (2019) Therapeutic targeting of lipid synthesis metabolism for selective elimination of cancer stem cells. Arch Pharm Res 42:25– 39. https://doi.org/10.1007/s12272-018-1098-z
- Kim AR, Park JI, Oh HT, Kim KM, Hwang JH, Jeong MG, Kim EH, Hwang ES, Hong JH (2019) TAZ stimulates liver regeneration through interleukin-6-induced hepatocyte proliferation and inhibition of cell death after liver injury. FASEB J 33:5914–5923. https://doi.org/10.1096/fj.201801256RR
- Kim KM, Oh HT, Yoo GD, Hwang JH, Oh A, Hwang ES, Hong JH (2020) Transcriptional coactivator with PDZ-binding motif stimulates epidermal regeneration via induction of amphiregulin expression after ultraviolet damage. Biochem Biophys Res Commun 524:242–248. https://doi.org/10.1016/j.bbrc.2020.01.079
- Komuro A, Nagai M, Navin NE, Sudol M (2003) WW domain-containing protein YAP associates with ErbB-4 and acts as a co-transcriptional activator for the carboxyl-terminal fragment of ErbB-4 that translocates to the nucleus. J Biol Chem 278:33334–33341. https://doi.org/10.1074/jbc.M305597200
- Konishi T, Schuster RM, Lentsch AB (2018) Proliferation of hepatic stellate cells, mediated by YAP and TAZ, contributes to liver repair and regeneration after liver ischemia/reperfusion injury. Am J Physiol Gastrointest Liver Physiol 314:G471–G482. https:// doi.org/10.1152/ajpgi.00153.2017
- Lee MJ, Byun MR, Furutani-Seiki M, Hong JH, Jung HS (2014) YAP and TAZ regulate skin wound healing. J Invest Dermatol 134:518– 525. https://doi.org/10.1038/jid.2013.339
- Lin KC, Park HW, Guan KL (2017) Regulation of the hippo pathway transcription factor TEAD. Trends Biochem Sci 42:862–872. https://doi.org/10.1016/j.tibs.2017.09.003
- Liu CY, Zha ZY, Zhou X, Zhang H, Huang W, Zhao D, Li T, Chan SW, Lim CJ, Hong W, Zhao S, Xiong Y, Lei QY, Guan KL (2010) The hippo tumor pathway promotes TAZ degradation by phosphorylating a phosphodegron and recruiting the SCF{beta}-TrCP E3 ligase. J Biol Chem 285:37159–37169. https://doi.org/10.1074/ jbc.M110.152942
- Liu-Chittenden Y, Huang B, Shim JS, Chen Q, Lee SJ, Anders RA, Liu JO, Pan D (2012) Genetic and pharmacological disruption of the TEAD-YAP complex suppresses the oncogenic activity of YAP. Genes Dev 26:1300–1305. https://doi.org/10.1101/gad. 192856.112
- Long F (2011) Building strong bones: molecular regulation of the osteoblast lineage. Nat Rev Mol Cell Biol 13:27–38. https://doi.org/10.1038/nrm3254
- Lorthongpanich C, Thumanu K, Tangkiettrakul K, Jiamvoraphong N, Laowtammathron C, Damkham N, Y UP, Issaragrisil S (2019) YAP as a key regulator of adipo-osteogenic differentiation in human MSCs. Stem Cell Res Ther 10:402. https://doi.org/10. 1186/s13287-019-1494-4
- Lu L, Li Y, Kim SM, Bossuyt W, Liu P, Qiu Q, Wang Y, Halder G, Finegold MJ, Lee JS, Johnson RL (2010) Hippo signaling is a potent in vivo growth and tumor suppressor pathway in the mammalian liver. Proc Natl Acad Sci U S A 107:1437–1442. https:// doi.org/10.1073/pnas.0911427107
- Lu L, Finegold MJ, Johnson RL (2018) Hippo pathway coactivators Yap and Taz are required to coordinate mammalian liver regeneration. Exp Mol Med 50:e423. https://doi.org/10.1038/emm.2017. 205
- Macias MJ, Hyvonen M, Baraldi E, Schultz J, Sudol M, Saraste M, Oschkinat H (1996) Structure of the WW domain of a kinaseassociated protein complexed with a proline-rich peptide. Nature 382:646–649. https://doi.org/10.1038/382646a0
- Miesfeld JB, Gestri G, Clark BS, Flinn MA, Poole RJ, Bader JR, Besharse JC, Wilson SW, Link BA (2015) Yap and Taz regulate retinal pigment epithelial cell fate. Development 142:3021– 3032. https://doi.org/10.1242/dev.119008

- Mohamed A, Sun C, De Mello V, Selfe J, Missiaglia E, Shipley J, Murray GI, Zammit PS, Wackerhage H (2016) The Hippo effector TAZ (WWTR1) transforms myoblasts and TAZ abundance is associated with reduced survival in embryonal rhabdomyosarcoma. J Pathol 240:3–14. https://doi.org/10.1002/path.4745
- Oku Y, Nishiya N, Shito T, Yamamoto R, Yamamoto Y, Oyama C, Uehara Y (2015) Small molecules inhibiting the nuclear localization of YAP/TAZ for chemotherapeutics and chemosensitizers against breast cancers. FEBS Open Bio 5:542–549. https://doi. org/10.1016/j.fob.2015.06.007
- Pan D (2010) The hippo signaling pathway in development and cancer. Dev Cell 19:491–505. https://doi.org/10.1016/j.devcel. 2010.09.011
- Panciera T, Azzolin L, Fujimura A, Di Biagio D, Frasson C, Bresolin S, Soligo S, Basso G, Bicciato S, Rosato A, Cordenonsi M, Piccolo S (2016) Induction of expandable tissue-specific stem/ progenitor cells through transient expression of YAP/TAZ. Cell Stem Cell 19:725–737. https://doi.org/10.1016/j.stem.2016.08. 009
- Park GH, Jeong H, Jeong MG, Jang EJ, Bae MA, Lee YL, Kim NJ, Hong JH, Hwang ES (2014) Novel TAZ modulators enhance myogenic differentiation and muscle regeneration. Br J Pharmacol 171:4051–4061. https://doi.org/10.1111/bph.12755
- Piccolo S, Dupont S, Cordenonsi M (2014) The biology of YAP/ TAZ: hippo signaling and beyond. Physiol Rev 94:1287–1312. https://doi.org/10.1152/physrev.00005.2014
- Plewes MR, Hou X, Zhang P, Liang A, Hua G, Wood JR, Cupp AS, Lv X, Wang C, Davis JS (2019) Yes-associated protein 1 is required for proliferation and function of bovine granulosa cells in vitrodagger. Biol Reprod 101:1001–1017. https://doi.org/10. 1093/biolre/ioz139
- Pobbati AV, Hong W (2020) A combat with the YAP/TAZ-TEAD oncoproteins for cancer therapy. Theranostics 10:3622–3635. https://doi.org/10.7150/thno.40889
- Pobbati AV, Rubin BP (2020) Protein-Protein Interaction Disruptors of the YAP/TAZ-TEAD Transcriptional Complex. Molecules 25:6001. https://doi.org/10.3390/molecules25246001
- Pobbati AV, Han X, Hung AW, Weiguang S, Huda N, Chen GY, Kang C, Chia CS, Luo X, Hong W, Poulsen A (2015) Targeting the Central Pocket in Human Transcription Factor TEAD as a Potential Cancer Therapeutic Strategy. Structure 23:2076–2086. https://doi.org/10.1016/j.str.2015.09.009
- Pocaterra A, Romani P, Dupont S (2020) YAP/TAZ functions and their regulation at a glance. J Cell Sci 133:230425. https://doi. org/10.1242/jcs.230425
- Reggiani F, Gobbi G, Ciarrocchi A, Sancisi V (2020) YAP and TAZ are not identical twins. Trends Biochem Sci 46:154–168. https:// doi.org/10.1016/j.tibs.2020.08.012
- Shin JH, Lee G, Jeong MG, Kim HK, Won HY, Choi Y, Lee JH, Nam M, Choi CS, Hwang GS, Hwang ES (2020) Transcriptional coactivator with PDZ-binding motif suppresses the expression of steroidogenic enzymes by nuclear receptor 4 A1 in Leydig cells. FASEB J 34:5332–5347. https://doi.org/10.1096/fj.20190 0695RRRR
- Strano S, Munarriz E, Rossi M, Castagnoli L, Shaul Y, Sacchi A, Oren M, Sudol M, Cesareni G, Blandino G (2001) Physical interaction with yes-associated protein enhances p73 transcriptional activity. J Biol Chem 276:15164–15173. https://doi.org/ 10.1074/jbc.M010484200
- Sudol M, Shields DC, Farooq A (2012) Structures of YAP protein domains reveal promising targets for development of new cancer drugs. Semin Cell Dev Biol 23:827–833. https://doi.org/10. 1016/j.semcdb.2012.05.002
- Sun C, De Mello V, Mohamed A, Ortuste Quiroga HP, Garcia-Munoz A, Al Bloshi A, Tremblay AM, von Kriegsheim A, Collie-Duguid E, Vargesson N, Matallanas D, Wackerhage H, Zammit

PS (2017) Common and distinctive functions of the hippo effectors taz and yap in skeletal muscle stem cell function. Stem Cells 35:1958–1972. https://doi.org/10.1002/stem.2652

- Tang Y, Weiss SJ (2017) Snail/Slug-YAP/TAZ complexes cooperatively regulate mesenchymal stem cell function and bone formation. Cell Cycle 16:399–405. https://doi.org/10.1080/15384101. 2017.1280643
- Tang Y, Feinberg T, Keller ET, Li XY, Weiss SJ (2016) Snail/Slug binding interactions with YAP/TAZ control skeletal stem cell selfrenewal and differentiation. Nat Cell Biol 18:917–929. https://doi. org/10.1038/ncb3394
- Tian Y, Kolb R, Hong JH, Carroll J, Li D, You J, Bronson R, Yaffe MB, Zhou J, Benjamin T (2007) TAZ promotes PC2 degradation through a SCFbeta-Trcp E3 ligase complex. Mol Cell Biol 27:6383–6395. https://doi.org/10.1128/MCB.00254-07
- Tsoi M, Morin M, Rico C, Johnson RL, Paquet M, Gevry N, Boerboom D (2019) Lats1 and Lats2 are required for ovarian granulosa cell fate maintenance. FASEB J 33:10819–10832. https://doi.org/10. 1096/fj.201900609R
- Varelas X, Sakuma R, Samavarchi-Tehrani P, Peerani R, Rao BM, Dembowy J, Yaffe MB, Zandstra PW, Wrana JL (2008) TAZ controls Smad nucleocytoplasmic shuttling and regulates human embryonic stem-cell self-renewal. Nat Cell Biol 10:837–848. https://doi.org/10.1038/ncb1748
- Wang J, Xiao Y, Hsu CW, Martinez-Traverso IM, Zhang M, Bai Y, Ishii M, Maxson RE, Olson EN, Dickinson ME, Wythe JD, Martin JF (2016) Yap and Taz play a crucial role in neural crest-derived craniofacial development. Development 143:504–515. https://doi. org/10.1242/dev.126920
- Watt KI, Judson R, Medlow P, Reid K, Kurth TB, Burniston JG, Ratkevicius A, De Bari C, Wackerhage H (2010) Yap is a novel regulator of C2C12 myogenesis. Biochem Biophys Res Commun 393:619–624. https://doi.org/10.1016/j.bbrc.2010.02.034
- Xia P, Gutl D, Zheden V, Heisenberg CP (2019) Lateral Inhibition in Cell Specification Mediated by Mechanical Signals Modulating TAZ Activity. Cell 176:1379–1392.e1314. https://doi.org/10. 1016/j.cell.2019.01.019
- Yang JY, Cho SW, An JH, Jung JY, Kim SW, Kim SY, Kim JE, Shin CS (2013) Osteoblast-targeted overexpression of TAZ increases bone mass in vivo. PLoS ONE 8:e56585. https://doi.org/10.1371/ journal.pone.0056585
- Yi X, Yu J, Ma C, Dong G, Shi W, Li H, Li L, Luo L, Sampath K, Ruan H, Huang H (2019) The effector of Hippo signaling, Taz, is required for formation of the micropyle and fertilization in zebrafish. PLoS Genet 15:e1007408. https://doi.org/10.1371/ journal.pgen.1007408
- Yu FX, Guan KL (2013) The Hippo pathway: regulators and regulations. Genes Dev 27:355–371. https://doi.org/10.1101/gad. 210773.112
- Yu FX, Zhao B, Guan KL (2015) Hippo pathway in organ size control, tissue homeostasis, and cancer. Cell 163:811–828. https://doi.org/ 10.1016/j.cell.2015.10.044
- Yuan Y, Park J, Feng A, Awasthi P, Wang Z, Chen Q, Iglesias-Bartolome R (2020) YAP1/TAZ-TEAD transcriptional networks maintain skin homeostasis by regulating cell proliferation and limiting KLF4 activity. Nat Commun 11:1472. https://doi.org/10.1038/ s41467-020-15301-0
- Yui S, Azzolin L, Maimets M, Pedersen MT, Fordham RP, Hansen SL, Larsen HL, Guiu J, Alves MRP, Rundsten CF, Johansen JV, Li Y, Madsen CD, Nakamura T, Watanabe M, Nielsen OH, Schweiger PJ, Piccolo S, Jensen KB (2018) YAP/TAZ-dependent reprogramming of colonic epithelium links ecm remodeling to tissue regeneration. Cell Stem Cell 22:35–49.e37. https://doi.org/10.1016/j. stem.2017.11.001
- Zanconato F, Forcato M, Battilana G, Azzolin L, Quaranta E, Bodega B, Rosato A, Bicciato S, Cordenonsi M, Piccolo S (2015)

Genome-wide association between YAP/TAZ/TEAD and AP-1 at enhancers drives oncogenic growth. Nat Cell Biol 17:1218–1227. https://doi.org/10.1038/ncb3216

- Zanconato F, Battilana G, Cordenonsi M, Piccolo S (2016) YAP/TAZ as therapeutic targets in cancer. Curr Opin Pharmacol 29:26–33. https://doi.org/10.1016/j.coph.2016.05.002
- Zhao B, Kim J, Ye X, Lai ZC, Guan KL (2009) Both TEAD-binding and WW domains are required for the growth stimulation and oncogenic transformation activity of yes-associated protein. Cancer Res 69:1089–1098. https://doi.org/10.1158/0008-5472. CAN-08-2997
- Zhao B, Tumaneng K, Guan KL (2011) The Hippo pathway in organ size control, tissue regeneration and stem cell self-renewal. Nat Cell Biol 13:877–883. https://doi.org/10.1038/ncb2303
- Zhou D, Conrad C, Xia F, Park JS, Payer B, Yin Y, Lauwers GY, Thasler W, Lee JT, Avruch J, Bardeesy N (2009) Mst1 and Mst2

maintain hepatocyte quiescence and suppress hepatocellular carcinoma development through inactivation of the Yap1 oncogene. Cancer Cell 16:425–438. https://doi.org/10.1016/j.ccr.2009.09. 026

Zhu Y, Wu Y, Cheng J, Wang Q, Li Z, Wang Y, Wang D, Wang H, Zhang W, Ye J, Jiang H, Wang L (2018) Pharmacological activation of TAZ enhances osteogenic differentiation and bone formation of adipose-derived stem cells. Stem Cell Res Ther 9:53. https://doi.org/10.1186/s13287-018-0799-z

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.