

Acute disseminated encephalomyelitis progressing to multiple sclerosis: Are infectious triggers involved?

Daniel S. Smyk · Anaïs K. Alexander ·
Mary Walker · Martin Walker

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Abstract Acute disseminated encephalomyelitis (ADEM) and multiple sclerosis (MS) are demyelinating disorders affecting the central nervous system. An auto-immune aetiology has been proposed for both. ADEM principally affects adolescents following acute infection by a variety of pathogens and has also been reported to occur following vaccination. ADEM typically resolves following medical treatment, whereas MS follows a more relapsing and remitting course. The pathogenesis of MS remains unclear, but it is thought that a combination of infectious and non-infectious environmental factors and host genetics act synergistically to cause disease. A variety of viruses, including Epstein Barr virus, cytomegalovirus, herpes simplex virus and varicella zoster virus, have been implicated as possible infectious triggers. The similar clinical and pathological presentation of ADEM and MS presents a

diagnostic challenge for distinguishing ADEM from a first episode of MS. Some cases of ADEM progress to MS for reasons that are not currently clear. This review examines the evidence for infectious agents as triggers for ADEM progressing to MS and suggests potential methods that may facilitate identification of infectious agents that may be responsible for the pathogenesis of ADEM to MS.

Keywords Autoimmunity · Autoimmune disease · Demyelinating disease · Environment · Infection · Multiple sclerosis

Abbreviations

ADEM	Acute disseminated encephalomyelitis
CIS	Clinically isolated syndrome
CMV	Cytomegalovirus
CSF	Cerebrospinal fluid
EBV	Epstein–Barr virus
HHV6	Human herpes virus 6
HSV	Herpes simplex virus
MOG	Myelin oligodendrocyte glycoprotein
MS	Multiple sclerosis
URTI	Upper respiratory tract infection
VZV	Varicella zoster virus

D. S. Smyk (✉)
Institute of Liver Studies, Division of Transplantation
Immunology and Mucosal Biology, King's College London
School of Medicine at King's College Hospital, Denmark Hill
Campus, London SE5 9RS, UK
e-mail: daniel.s.smyk@doctors.org.uk

A. K. Alexander
Department of Psychiatry, Greater Manchester West Mental
Health NHS Foundation Trust, Bury New Road, Prestwich,
Manchester M25 3BL, UK

M. Walker
Department of Medicine, Maidstone and Tunbridge Wells
Hospital NHS Trust, Hermitage Lane, Maidstone ME16 9QQ,
UK

M. Walker
Department of Infectious Disease Epidemiology, Faculty of
Medicine, School of Public Health, Imperial College London,
Norfolk Place, London W2 1PG, UK

Introduction

Acute disseminated encephalomyelitis (ADEM) is an autoimmune, demyelinating disease affecting the central nervous system (CNS) [1–3]. Children are more commonly affected than adults. The disease is often triggered by bacterial or viral infections, and rarely vaccinations [4]. Many cases are reportedly preceded by bacterial and viral upper respiratory tract infections (URTI). Like many

autoimmune diseases, it is believed that a combination of genetic predisposition and environmental factors (including infections) acts synergistically to induce autoimmunity. In multiple sclerosis (MS), bacterial and viral agents have also been implicated as triggers of autoimmunity, disease flares and disease progression and are also implicated as potential causes of concomitant autoimmune disease [5, 6]. It is well recognised that multiple autoimmune conditions often co-exist in a patient, which raises the question of common triggers between these conditions.

ADEM is characterised by multiple inflammatory lesions in the brain and spinal cord, predominantly in the white matter. These lesions are morphologically indistinguishable from those seen in MS [7]. ADEM often presents as a monophasic condition with multiple neurological signs and symptoms, as well as encephalopathy [1–3]. Magnetic resonance imaging (MRI) of patients suffering ADEM shows subcortical white matter lesions and lesions in the deep grey matter of the thalami and basal ganglia [8]. The diagnosis of ADEM is based on clinical and radiological findings, with differential diagnoses including MS and the clinically isolated syndrome (CIS), transverse myelitis and neuromyelitis optica [8]. Bacterial and viral meningitis and encephalitis must also be excluded. Relapsing cases and the appearance of new lesions are suggestive of multiphasic ADEM or MS. It is often difficult to differentiate between ADEM and MS, with some cases of ADEM progressing to MS without explanation [8, 9].

This review examines the possible roles of infectious agents in inducing the progression of ADEM to MS through mechanisms such as loss of self-tolerance to specific myelin peptides via molecular mimicry and cross reactivity, in genetically and immunologically susceptible individuals. We also focus on possible methods for detecting and characterising the many infectious agents that may be involved during the protracted disease progression.

Autoantibodies: a clue to molecular mimicry and cross reactivity?

The pathogenesis of ADEM has not been fully elucidated, nor have its autoantigenic determinants. Implicated antigens include myelin basic protein, proteolipid protein and myelin oligodendrocyte glycoprotein (MOG) [8, 10]. T-cell reactivity to myelin basic protein is tenfold greater in ADEM patients compared to patients with encephalitis or healthy control patients [1]. Several studies have reported some ADEM patients with high titers of anti-MOG antibodies [11–16]. Di Pauli et al. [12] found that 44 % of ADEM patients had high titer anti-MOG IgG and noted that anti-MOG titers decreased with clinical improvement

in a longitudinal analysis of 266 patients. Similar results were reported by Brilot et al. [11] who found anti-MOG IgG in 40 % of ADEM/CIS patients. Lalive et al. [13] found that anti-MOG antibodies were highly reactive in children with ADEM, and that they were also present in MS. Probstel et al. [16] followed 25 children with anti-MOG-positive ADEM over 5 years. They found that anti-MOG antibodies declined rapidly in 16 monophasic ADEM cases and in one CIS case. Interestingly, anti-MOG antibodies persisted and even increased in six of eight cases who later developed paediatric MS [16]. Adult and paediatric controls were negative for anti-MOG. Six percent of adult MS patients were also found to have low titers of anti-MOG antibodies [16]. Probstel et al. [16] proposed that anti-MOG may therefore be present at the onset of inflammation in early MS, but may decline over time. A study by Menge et al. [17] found that correctly folded antibodies against MOG are found in paediatric MS and ADEM, but not in adult MS. The authors suggest that these differences may be related to molecular mimicry and cross reactivity [17], findings that raise the question of whether common infectious agents are shared between ADEM and MS. It is possible that specific infectious agents cause MS and/or ADEM in isolation and that certain infections lead to a form of ADEM which progresses to MS in a minority of cases.

Infectious triggers in multiple sclerosis

MS is an autoimmune disease characterised by chronic inflammation, nerve demyelination and gliosis in the CNS [18, 19]. Periods of relapse and remission are common and typify the most common form of the disease [18, 19]. Inflammatory lesions with areas of demyelination in the CNS characterise the gross pathology of MS [20]. The lesions are composed of mononuclear cell infiltrates (largely T and B lymphocytes, plasma cells, macrophages and microglia) in the perivascular spaces that develop into plaques [18, 21]. The peripheral regions of the plaques stain positive for IgG [18, 21]. Furthermore, 90 % of MS patients show intrathecal IgG synthesis in cerebrospinal fluid (CSF) [18]. The risk for developing MS is typically spread over a long period of time [22–24], although several studies indicate that children as young as 11 may be susceptible to the disease [25]. Although MS is primarily diagnosed in young adults, paediatric MS has been recognised [26].

Multiple viruses have been implicated in the pathogenesis of MS and disease flares [18, 19, 27–32]. Viruses can damage infected cells directly or trigger autoimmune reactions that cause demyelination. It is believed that autoimmunity develops due to bystander activation,

epitope spreading, molecular mimicry and cross reactivity between viral and self-peptides such as those in myelin [33–36]. Although multiple viruses have been implicated with varying degrees of evidence (reviewed in [37]), Epstein–Barr virus (EBV) and human herpes virus (HHV) (specifically HHV6) are most strongly implicated [18, 19, 29, 30, 37–40].

The relapse-remittance pattern of chronic HHV infection is similar to the clinical pattern of MS [19], and HHV pools have been found in brain tissue [41, 42] and the CSF of MS patients [43, 44]. Infection with HHV6 in childhood is associated with an increased risk of developing MS [40]. It has also been demonstrated that HHV6 is capable of infecting most glial cell-types [45]. Moreover, increased HHV6 DNA has been found in MS plaques, and monoclonal antibodies were able to detect HHV6 antigen in brain tissue from MS plaques but not control brain tissue [38, 39]. Molecular mimicry has been indicated, as sequence homology has been found between myelin basic protein and HHV6 encoded U24, and cross-reactive T cells responding to both protein types are found to be increased in MS patients [46].

EBV has been strongly associated with autoimmunity and specifically with MS. EBV has been shown to be highly prevalent in MS patients [47, 48], and a higher prevalence in paediatric MS cases compared to paediatric controls [49]. Although EBV is ubiquitous, the incidence of MS is higher among individuals with histories of symptomatic infectious mononucleosis [50]. Also, initial infection of B lymphocytes with EBV could account for the presence of increased oligoclonal bands from proliferating cell clones [51]. EBV-IgG has been noted in a cohort of MS patients up to 20 years prior to the onset of MS [52], and MS patients have been reported to have increased CD4+ T cells against EBV nuclear antigens [53]. Interestingly, myelin antigens were found to be cross reactive with EBV nuclear antigens and specific CD4+ T cells in cell studies [54, 55]. It has also been proposed that EBV may infect and immortalise autoreactive B cells, although no definitive conclusions have been drawn on whether EBV-infected B cells are present in brain tissue of MS patients [56–60]. A recent study by Pender et al. [61] reports defective cytotoxic CD8+ T-cell control of EBV which may contribute to the development of MS, as autoreactive B cells infected with EBV would accumulate in the CNS [61]. Interestingly, in the same study, MS patients were treated with in vitro expanded autologous EBV-specific CD8+ T cells against viral proteins, with subjective and objective clinical improvement in symptoms [61]. Several other infectious agents have been implicated in the pathogenesis of MS, but these have a weaker evidence base. A small number of these have also been implicated in ADEM, including EBV, coronavirus,

cytomegalovirus (CMV), measles virus, and varicella zoster virus (VZV).

Viruses common in ADEM and MS

Most cases of ADEM are preceded by infection, yet often the infection has cleared before neurological signs and symptoms develop. Tissues samples from ADEM lesions are rarely obtained due to the good prognosis of the condition [7]. Several bacteria and viruses have been identified in ADEM patients.

The role of EBV has been well studied in MS and has also been implicated in ADEM, and in some cases of ADEM progressing to MS. Banwell et al. [26] examined the clinical features and viral serologies of 137 paediatric MS patients and 96 controls. A first MS attack that resembled ADEM was reported in 16 % of patients. Those patients tended to be younger with polyfocal or monofocal presentations [26]. Seropositivity for EBV was found in 86 % of cases compared to 64 % of controls, irrespective of geographical location suggesting that EBV may play a role in paediatric MS [26]. Fujimoto et al. [62] identified 10 cases of CNS syndromes following infectious mononucleosis in patients between 1984 and 2002. Two of these cases were ADEM with both having EBV detected in the CSF by PCR [62]. ADEM was observed in two of these patients. Both had EBV titers detected in the CSF by PCR. The remaining cases of ADEM linked with EBV are largely comprised of individual case reports and small studies [63–81].

Coronavirus is capable of infecting neural cells and has been implicated in MS [82, 83] and ADEM [84], although the evidence for this is weak. Yeh et al. note that coronavirus can induce a chronic demyelinating disease resembling MS in a murine model [84]. There are also few studies relating CMV to MS [85] and to ADEM [77, 86–88]. Brok et al. [86] demonstrated that the human CMV major capsid protein shares sequence similarities with the MOG-34-56. Rhesus monkeys, immunised with MOG-34-56, were found to develop neurological disease resembling human ADEM, and the mononuclear cell infiltrates in the demyelinating lesions were predominantly MOG-34-56 [86] T cells. MOG-34-56 reactive CD4+ and CD8+ T cells were also induced in monkeys immunised with human CMV major capsid protein [86].

Measles virus has been loosely linked with both MS and ADEM [32, 89–97]. Hagiwara et al. [91] report a case of ADEM following *Mycoplasma pneumonia* infection complicated with measles and comment on the potential involvement of multiple infectious agents in the pathogenesis of ADEM. VZV has been isolated in the CSF of MS patients and has also been identified during MS flares

[98–100]. A small number of case reports also note VZV infection in ADEM cases, with several reporting the presence of VZV in the CSF [101–110].

Although several viruses are associated with ADEM and MS, very little research has specifically focused on the progression from ADEM to MS. Indeed, most published accounts are case reports rather than formal clinical or epidemiological studies.

Respiratory tract infections preceding MS and ADEM

A history of URTI frequently precedes cases of ADEM and has been documented in several studies of MS patients. A prospective study of paediatric ADEM patients from January 2009 to January 2011 found that 57 % of patients had a history of URTI preceding their disease, with three cases showing infection with HSV and EBV [67]. There were no differences between cases and controls with respect to other viral infections such as CMV, parvovirus B19, VZV or HSV [26]. A case–control study of 225 MS cases and 900 controls by Marrie et al. [50] set out to determine whether URTIs are related to the onset of MS symptoms using the General Practice Research Database in the United Kingdom. Mean rates of respiratory tract infection were compared at intervals of 5 weeks, 3 months and 12 months prior to the onset of first symptoms. They found an increased frequency of URTI preceding MS onset, with significantly increased MS risk. Additionally, they demonstrated that a history of infectious mononucleosis was associated with a fivefold increased risk of developing MS.

Future prospects

The factors contributing to the development of MS, the various MS types and disease flares are unclear. Likewise, why some, particularly paediatric, cases of ADEM progress to develop MS is poorly understood. It is likely that inherent immunological and genetic factors contribute to this progression, although further research is needed [111–115]. Infectious agents are also probably involved in the progression of ADEM to MS, especially since both conditions are implicated with shared infections and that there are similarities with certain autoantibody profiles. Specific infectious agents may play a role in ADEM and MS in isolation, while others lead to progression of ADEM to MS. The lack of more implicated infectious agents may be due to a paucity of research rather than negative findings. The recent introduction of the infectome model may allow researchers to identify infectious agents involved in ADEM to MS progression, or indeed, infections that may be protective [5, 6]. The infectome model is based on

geographical, epidemiological, serological and molecular evidence of the presence and co-occurrence of infectious agents associated with autoimmunity [5, 6]. In the case of ADEM, regular monitoring and sampling of patients to detect infections preceding progression to MS could be implemented. Follow-up and sampling of MS patients may also elucidate the role of infectious triggers to disease flares.

Conflict of interest There are no conflicts of interest.

References

- Pohl-Koppe A, Burchett SK, Thiele EA, Hafler DA. Myelin basic protein reactive Th2 T cells are found in acute disseminated encephalomyelitis. *J Neuroimmunol*. 1998;91:19–27.
- Stuve O, Zamvil SS. Pathogenesis, diagnosis, and treatment of acute disseminated encephalomyelitis. *Curr Opin Neurol*. 1999;12:395–401.
- Tenembaum S, Chitnis T, Ness J, Hahn JS. Acute disseminated encephalomyelitis. *Neurology*. 2007;68:S23–36.
- Fenichel GM. Neurological complications of immunization. *Ann Neurol*. 1982;12:119–28.
- Bogdanos DP, Smyk DS, Invernizzi P, Rigopoulou EI, Blank M, Pouria S, et al. Infectome: a platform to trace infectious triggers of autoimmunity. *Autoimmun Rev*. 2013;12:726–40.
- Bogdanos DP, Smyk DS, Invernizzi P, Rigopoulou EI, Blank M, Sakkas L, et al. Tracing environmental markers of autoimmunity: introducing the infectome. *Immunol Res*. 2013;56:220–40.
- Guenther AD, Munoz DG. Plaque-like demyelination in acute disseminated encephalomyelitis (ADEM)—an autopsy case report. *Clin Neuropathol*. 2013;32:486–91.
- Lee YJ. Acute disseminated encephalomyelitis in children: differential diagnosis from multiple sclerosis on the basis of clinical course. *Korean J Pediatr*. 2011;54:234–40.
- Liptai Z, Ujhelyi E, Mihaly I, Rudas G, Barsi P. Acute disseminated encephalomyelitis in childhood. *Ideggyogy Sz*. 2009;62:244–54.
- Stonehouse M, Gupte G, Wassmer E, Whitehouse WP. Acute disseminated encephalomyelitis: recognition in the hands of general paediatricians. *Arch Dis Child*. 2003;88:122–4.
- Brilot F, Dale RC, Selter RC, Grummel V, Kalluri SR, Aslam M, et al. Antibodies to native myelin oligodendrocyte glycoprotein in children with inflammatory demyelinating central nervous system disease. *Ann Neurol*. 2009;66:833–42.
- Di Pauli F, Mader S, Rostasy K, Schanda K, Bajer-Kornek B, Ehling R, et al. Temporal dynamics of anti-MOG antibodies in CNS demyelinating diseases. *Clin Immunol*. 2011;138:247–54.
- Lalive PH, Hausler MG, Maurey H, Mikaeloff Y, Tardieu M, Wiendl H, et al. Highly reactive anti-myelin oligodendrocyte glycoprotein antibodies differentiate demyelinating diseases from viral encephalitis in children. *Mult Scler*. 2011;17:297–302.
- McLaughlin KA, Chitnis T, Newcombe J, Franz B, Kennedy J, McArdel S, et al. Age-dependent B cell autoimmunity to a myelin surface antigen in pediatric multiple sclerosis. *J Immunol*. 2009;183:4067–76.
- O'Connor KC, McLaughlin KA, De Jager PL, Chitnis T, Bettelli E, Xu C, et al. Self-antigen tetramers discriminate between myelin autoantibodies to native or denatured protein. *Nat Med*. 2007;13:211–7.
- Probstel AK, Dornmair K, Bittner R, Sperl P, Jenne D, Magalhaes S, et al. Antibodies to MOG are transient in childhood

- acute disseminated encephalomyelitis. *Neurology*. 2011;77:580–8.
17. Menge T, Kieseier BC, Nessler S, Hemmer B, Hartung HP, Stuve O. Acute disseminated encephalomyelitis: an acute hit against the brain. *Curr Opin Neurol*. 2007;20:247–54.
 18. Gildden DH. Infectious causes of multiple sclerosis. *Lancet Neurol*. 2005;4:195–202.
 19. Kakalacheva K, Lunemann JD. Environmental triggers of multiple sclerosis. *FEBS Lett*. 2011;585:3724–9.
 20. Noseworthy JH, Lucchinetti C, Rodriguez M, Weinshenker BG. Multiple sclerosis. *N Engl J Med*. 2000;343:938–52.
 21. Frohman EM, Racke MK, Raine CS. Multiple sclerosis—the plaque and its pathogenesis. *N Engl J Med*. 2006;354:942–55.
 22. Hammond SR, English DR, McLeod JG. The age-range of risk of developing multiple sclerosis: evidence from a migrant population in Australia. *Brain*. 2000;123(Pt 5):968–74.
 23. Kurtzke JF, Delasnerie-Laupretre N, Wallin MT. Multiple sclerosis in North African migrants to France. *Acta Neurol Scand*. 1998;98:302–9.
 24. Kurtzke JF, Hyllested K. Multiple sclerosis in the Faroe Islands: I. Clinical and epidemiological features. *Ann Neurol*. 1979;5:6–21.
 25. Kurtzke JF. Epidemiologic evidence for multiple sclerosis as an infection. *Clin Microbiol Rev*. 1993;6:382–427.
 26. Banwell B, Krupp L, Kennedy J, Tellier R, Tenenbaum S, Ness J, et al. Clinical features and viral serologies in children with multiple sclerosis: a multinational observational study. *Lancet Neurol*. 2007;6:773–81.
 27. Fleming JO. Helminths and multiple sclerosis: will old friends give us new treatments for MS? *J Neuroimmunol*. 2011;233:3–5.
 28. Gaisford W, Cooke A. Can infections protect against autoimmunity? *Curr Opin Rheumatol*. 2009;21:391–6.
 29. Giovannoni G, Cutter GR, Lunemann J, Martin R, Munz C, Sriram S, et al. Infectious causes of multiple sclerosis. *Lancet Neurol*. 2006;5:887–94.
 30. Giovannoni G, Ebers G. Multiple sclerosis: the environment and causation. *Curr Opin Neurol*. 2007;20:261–8.
 31. Sibley WA, Bamford CR, Clark K. Clinical viral infections and multiple sclerosis. *Lancet*. 1985;1:1313–5.
 32. Jacobson S, Flerlage ML, McFarland HF. Impaired measles virus-specific cytotoxic T cell responses in multiple sclerosis. *J Exp Med*. 1985;162:839–50.
 33. Wisniewski HM, Bloom BR. Primary demyelination as a non-specific consequence of a cell-mediated immune reaction. *J Exp Med*. 1975;141:346–59.
 34. Brosnan CF, Selmaj K, Raine CS. Hypothesis: a role for tumor necrosis factor in immune-mediated demyelination and its relevance to multiple sclerosis. *J Neuroimmunol*. 1988;18:87–94.
 35. Wucherpfennig KW, Strominger JL. Molecular mimicry in T cell-mediated autoimmunity: viral peptides activate human T cell clones specific for myelin basic protein. *Cell*. 1995;80:695–705.
 36. Lehmann PV, Forsthuber T, Miller A, Sercarz EE. Spreading of T-cell autoimmunity to cryptic determinants of an autoantigen. *Nature*. 1992;358:155–7.
 37. Cermelli C, Jacobson S. Viruses and multiple sclerosis. *Viral Immunol*. 2000;13:255–67.
 38. Caserta MT, Hall CB, Schnabel K, McIntyre K, Long C, Costanzo M, et al. Neuroinvasion and persistence of human herpesvirus 6 in children. *J Infect Dis*. 1994;170:1586–9.
 39. Challoner PB, Smith KT, Parker JD, MacLeod DL, Coulter SN, Rose TM, et al. Plaque-associated expression of human herpesvirus 6 in multiple sclerosis. *Proc Natl Acad Sci USA*. 1995;92:7440–4.
 40. Kurtzke JF, Hyllested K, Heltberg A. Multiple sclerosis in the Faroe Islands: transmission across four epidemics. *Acta Neurol Scand*. 1995;91:321–5.
 41. Albright AV, Lavi E, Black JB, Goldberg S, O'Connor MJ, Gonzalez-Scarano F. The effect of human herpesvirus-6 (HHV-6) on cultured human neural cells: oligodendrocytes and microglia. *J Neurovirol*. 1998;4:486–94.
 42. Chan PK, Ng HK, Cheng AF. Detection of human herpesviruses 6 and 7 genomic sequences in brain tumours. *J Clin Pathol*. 1999;52:620–3.
 43. Kim JS, Lee KS, Park JH, Kim MY, Shin WS. Detection of human herpesvirus 6 variant A in peripheral blood mononuclear cells from multiple sclerosis patients. *Eur Neurol*. 2000;43:170–3.
 44. Opsahl ML, Kennedy PG. Early and late HHV-6 gene transcripts in multiple sclerosis lesions and normal appearing white matter. *Brain*. 2005;128:516–27.
 45. Lusso P, Markham PD, Tschachler E, di Marzo Veronese F, Salahuddin SZ, Ablashi DV, et al. In vitro cellular tropism of human B-lymphotropic virus (human herpesvirus-6). *J Exp Med*. 1988;167:1659–70.
 46. Mirandola P, Stefan A, Brambilla E, Campadelli-Fiume G, Grimaldi LM. Absence of human herpesvirus 6 and 7 from spinal fluid and serum of multiple sclerosis patients. *Neurology*. 1999;53:1367–8.
 47. Goodin DS. The causal cascade to multiple sclerosis: a model for MS pathogenesis. *PLoS ONE*. 2009;4:e4565.
 48. Sumaya CV, Myers LW, Ellison GW, Ench Y. Increased prevalence and titer of Epstein–Barr virus antibodies in patients with multiple sclerosis. *Ann Neurol*. 1985;17:371–7.
 49. Alotaibi S, Kennedy J, Tellier R, Stephens D, Banwell B. Epstein–Barr virus in pediatric multiple sclerosis. *JAMA*. 2004;291:1875–9.
 50. Marrie RA, Wolfson C, Sturkenboom MC, Gout O, Heinzlef O, Roulet E, et al. Multiple sclerosis and antecedent infections: a case-control study. *Neurology*. 2000;54:2307–10.
 51. Rand KH, Houck H, Denslow ND, Heilman KM. Epstein–Barr virus nuclear antigen-1 (EBNA-1) associated oligoclonal bands in patients with multiple sclerosis. *J Neurol Sci*. 2000;173:32–9.
 52. DeLorenze GN, Munger KL, Lennette ET, Orentreich N, Vogelmann JH, Ascherio A. Epstein–Barr virus and multiple sclerosis: evidence of association from a prospective study with long-term follow-up. *Arch Neurol*. 2006;63:839–44.
 53. Lunemann JD, Edwards N, Muraro PA, Hayashi S, Cohen JI, Munz C, et al. Increased frequency and broadened specificity of latent EBV nuclear antigen-1-specific T cells in multiple sclerosis. *Brain*. 2006;129:1493–506.
 54. Lunemann JD, Jelcic I, Roberts S, Lutterotti A, Tackenberg B, Martin R, et al. EBNA1-specific T cells from patients with multiple sclerosis cross react with myelin antigens and co-produce IFN-gamma and IL-2. *J Exp Med*. 2008;205:1763–73.
 55. Bray PF, Luka J, Bray PF, Culp KW, Schlight JP. Antibodies against Epstein–Barr nuclear antigen (EBNA) in multiple sclerosis CSF, and two pentapeptide sequence identities between EBNA and myelin basic protein. *Neurology*. 1992;42:1798–804.
 56. Peferoen LA, Lamers F, Lodder LN, Gerritsen WH, Huitinga I, Melief J, et al. Epstein Barr virus is not a characteristic feature in the central nervous system in established multiple sclerosis. *Brain*. 2010;133:e137.
 57. Serafini B, Rosicarelli B, Franciotta D, Magliozzi R, Reynolds R, Cinque P, et al. Dysregulated Epstein–Barr virus infection in the multiple sclerosis brain. *J Exp Med*. 2007;204:2899–912.
 58. Thorley-Lawson DA. Epstein–Barr virus: exploiting the immune system. *Nat Rev Immunol*. 2001;1:75–82.
 59. Willis SN, Stadelmann C, Rodig SJ, Caron T, Gattenloehner S, Mallozzi SS, et al. Epstein–Barr virus infection is not a characteristic feature of multiple sclerosis brain. *Brain*. 2009;132:3318–28.
 60. Sanders VJ, Felisan S, Waddell A, Tourtellotte WW. Detection of herpesviridae in postmortem multiple sclerosis brain tissue

- and controls by polymerase chain reaction. *J Neurovirol.* 1996;2:249–58.
61. Pender MP, Csurhes PA, Smith C, Beagley L, Hooper KD, Raj M, et al. Epstein–Barr virus-specific adoptive immunotherapy for progressive multiple sclerosis. *Mult Scler.* 2014. doi:10.1177/1352458514521888.
 62. Fujimoto H, Asaoka K, Imaizumi T, Ayabe M, Shoji H, Kaji M. Epstein–Barr virus infections of the central nervous system. *Intern Med.* 2003;42:33–40.
 63. Apilanez Urquiola M, Sarasua Miranda A, Perez Ruiz E, Nogues Perez A, Garcia Santiago J. Acute disseminated encephalomyelitis secondary to Epstein–Barr virus. *An Pediatr (Barc).* 2003;58:282–3.
 64. Bahadori HR, Williams VC, Turner RP, Rumboldt Z, Reigart JR, Fowler SL, et al. Acute disseminated encephalomyelitis following infectious mononucleosis. *J Child Neurol.* 2007;22:324–8.
 65. Baron J, Herrero-Velazquez S, Ruiz-Pinero M, Pedraza MI, Rojo-Rello S, Guerrero-Peral AL. Encephalitis due to the Epstein–Barr virus: a description of a clinical case and review of the literature. *Rev Neurol.* 2013;57:451–4.
 66. Caucheteux N, Maarouf A, Daelman L, Toupance O, Lavaud S, Tourbah A. Acute disseminated encephalomyelitis in two renal transplant patients: is there a role for Epstein–Barr virus reactivation? *Mult Scler.* 2013;19:1222–5.
 67. Elhassanien AF, Aziz HA. Acute demyelinating encephalomyelitis: clinical characteristics and outcome. *J Pediatr Neurosci.* 2013;8:26–30.
 68. Erol I, Ozkale Y, Alkan O, Alehan F. Acute disseminated encephalomyelitis in children and adolescents: a single center experience. *Pediatr Neurol.* 2013;49:266–73.
 69. Gomez Sanchez E, Mateos Beato F, Sanchez Diaz JI, Simon de las Heras R, Ballester Diaz Y. Acute disseminated encephalomyelitis: experience of a tertiary hospital in Spain. *An Pediatr (Barc).* 2005;63:203–11.
 70. Grillo E, da Silva RJ, Barbato Filho JH. Epstein–Barr virus acute encephalomyelitis in a 13-year-old boy. *Eur J Paediatr Neurol.* 2008;12:417–20.
 71. Hoshino T, Hatsumi N, Takada S, Sakura T, Sakurai A, Miyawaki S. Acute disseminated encephalomyelitis during treatment for idiopathic thrombocytopenic purpura. *Rinsho Ketsueki.* 2008;49:505–9.
 72. Igarashi K, Kajino M, Shirai M, Oki J, Seki K. A case of acute disseminated encephalomyelitis associated with Epstein–Barr virus infection. *No To Hattatsu.* 2011;43:59–61.
 73. Ipsen P. CT-verified intracranial calcifications and contrast enhancement in acute disseminated encephalomyelitis: a case report. *Pediatr Radiol.* 1998;28:591–3.
 74. Kim SC, Jang HJ, Han DJ. Acute disseminated encephalomyelitis after renal transplantation in patients with positive Epstein–Barr virus antibody. *Transplant Proc.* 1998;30:3139.
 75. Mohsen H, Abu Zeinab GF, Elstouhy AH, Mohamed K. Acute disseminated encephalomyelitis following infectious mononucleosis in a toddler. *BMJ Case Rep.* 2013. doi:10.1136/bcr-2013-010048.
 76. Murthy SN, Faden HS, Cohen ME, Bakshi R. Acute disseminated encephalomyelitis in children. *Pediatrics.* 2002;110:e21.
 77. Revel-Vilk S, Hurvitz H, Klar A, Virozov Y, Korn-Lubetzki I. Recurrent acute disseminated encephalomyelitis associated with acute cytomegalovirus and Epstein–Barr virus infection. *J Child Neurol.* 2000;15:421–4.
 78. Selter RC, Brilof F, Grummel V, Kraus V, Cepok S, Dale RC, et al. Antibody responses to EBV and native MOG in pediatric inflammatory demyelinating CNS diseases. *Neurology.* 2010;74:1711–5.
 79. Shoji H, Kusuhara T, Honda Y, Hino H, Kojima K, Abe T, et al. Relapsing acute disseminated encephalomyelitis associated with chronic Epstein–Barr virus infection: MRI findings. *Neuroradiology.* 1992;34:340–2.
 80. Tolly TL, Wells RG, Sty JR. MR features of fleeting CNS lesions associated with Epstein–Barr virus infection. *J Comput Assist Tomogr.* 1989;13:665–8.
 81. Ueda M, Tateishi T, Shigetou H, Yamasaki R, Ohyagi Y, Kira J. A case of acute disseminated encephalomyelitis associated with Epstein–Barr virus reactivation during infliximab therapy. *Rinsho Shinkeigaku.* 2010;50:461–6.
 82. Murray RS, Brown B, Brian D, Cabirac GF. Detection of coronavirus RNA and antigen in multiple sclerosis brain. *Ann Neurol.* 1992;31:525–33.
 83. Stewart JN, Mounir S, Talbot PJ. Human coronavirus gene expression in the brains of multiple sclerosis patients. *Virology.* 1992;191:502–5.
 84. Yeh EA, Collins A, Cohen ME, Duffner PK, Faden H. Detection of coronavirus in the central nervous system of a child with acute disseminated encephalomyelitis. *Pediatrics.* 2004;113:e73–6.
 85. Wroblewska Z, Gilden D, Devlin M, Huang ES, Rorke LB, Hamada T, et al. Cytomegalovirus isolation from a chimpanzee with acute demyelinating disease after inoculation of multiple sclerosis brain cells. *Infect Immun.* 1979;25:1008–15.
 86. Brok HP, Boven L, van Meurs M, Kerlero de Rosbo N, Celebi-Paul L, Kap YS, et al. The human CMV-UL86 peptide 981-1003 shares a crossreactive T-cell epitope with the encephalitogenic MOG peptide 34-56, but lacks the capacity to induce EAE in rhesus monkeys. *J Neuroimmunol.* 2007;182:135–52.
 87. Kanzaki A, Yabuki S. Acute disseminated encephalomyelitis (ADEM) associated with cytomegalovirus infection—a case report. *Rinsho Shinkeigaku.* 1994;34:511–3.
 88. Zaguri R, Shelef I, Ifergan G, Almog Y. Fatal acute disseminated encephalomyelitis associated with cytomegalovirus infection. *BMJ Case Rep.* 2009. doi:10.1136/bcr.07.2008.0443.
 89. Haase AT, Ventura P, Gibbs CJ Jr, Tourtellotte WW. Measles virus nucleotide sequences: detection by hybridization in situ. *Science.* 1981;212:672–5.
 90. Chowdhary J, Ashraf SM, Khajuria K. Measles with acute disseminated encephalomyelitis (ADEM). *Indian Pediatr.* 2009;46:72–4.
 91. Hagiwara H, Sakamoto S, Katsumata T, Katayama Y. Acute disseminated encephalomyelitis developed after *Mycoplasma pneumoniae* infection complicating subclinical measles infection. *Intern Med.* 2009;48:479–83.
 92. Lee WT, Wang PJ, Liu HM, Young C, Tseng CL, Chang YC, et al. Acute disseminated encephalomyelitis in children: clinical, neuroimaging and neurophysiologic studies. *Zhonghua Minguo xiao er ke yi xue hui za zhi [Journal] Zhonghua Minguo xiao er ke yi xue hui.* 1996;37:197–203.
 93. Murphy J, Austin J. Spontaneous infection or vaccination as cause of acute disseminated encephalomyelitis. *Neuroepidemiology.* 1985;4:138–45.
 94. Nagai K, Mori T. Acute disseminated encephalomyelitis with probable measles vaccine failure. *Pediatr Neurol.* 1999;20:399–402.
 95. Nardone R, Golaszewski S, Trinka E, Tezzon F, Zuccoli G. Acute disseminated encephalomyelitis preceding measles exanthema. *J Child Neurol.* 2011;26:1590–2.
 96. Sriram S, Steinman L. Postinfectious and postvaccinial encephalomyelitis. *Neurol Clin.* 1984;2:341–53.
 97. Yokoyama T, Sakurai M, Aota Y, Wakabayashi Y, Ohyashiki K. An adult case of acute disseminated encephalomyelitis accompanied with measles infection. *Intern Med.* 2005;44:1204–5.
 98. Mancuso R, Delbue S, Borghi E, Pagani E, Calvo MG, Caputo D, et al. Increased prevalence of varicella zoster virus DNA in cerebrospinal fluid from patients with multiple sclerosis. *J Med Virol.* 2007;79:192–9.

99. Ordonez G, Pineda B, Garcia-Navarrete R, Sotelo J. Brief presence of varicella-zoster viral DNA in mononuclear cells during relapses of multiple sclerosis. *Arch Neurol*. 2004;61:529–32.
100. Sotelo J, Martinez-Palomo A, Ordonez G, Pineda B. Varicella-zoster virus in cerebrospinal fluid at relapses of multiple sclerosis. *Ann Neurol*. 2008;63:303–11.
101. Corti M, Trione N, Villafane MF, Risso D, Yampolsky C, Mamanna L. Acute meningoencephalomyelitis due to varicella-zoster virus in an AIDS patient: report of a case and review of the literature. *Rev Soc Bras Med Trop*. 2011;44:784–6.
102. Curcoy Barcenilla AI, Pons Odena M, Vernet Bori A. Acute disseminated encephalomyelitis secondary to varicella. *An Esp Pediatr*. 2002;56:68–9.
103. Gilden DH. Varicella zoster virus vasculopathy and disseminated encephalomyelitis. *J Neurol Sci*. 2002;195:99–101.
104. Gucuyener K, Kula S, Serdaroglu A, Tah ET. Acute disseminated encephalomyelitis exacerbated by varicella. *Acta Paediatr Jpn*. 1997;39:619–23.
105. Lademann M, Gabelin P, Lafrenz M, Wernitz C, Ehmke H, Schmitz H, et al. Acute disseminated encephalomyelitis following *Plasmodium falciparum* malaria caused by varicella zoster virus reactivation. *Am J Trop Med Hyg*. 2005;72:478–80.
106. Mariotti P, Colosimo C, Frisullo G, Caggiula M, Della Marca GD, Valentini P, et al. Relapsing demyelinating disease after chicken pox in a child. *Neurology*. 2006;66:1953–4.
107. Murthy JM, Yangala R, Meena AK, Jaganmohan Reddy J. Acute disseminated encephalomyelitis: clinical and MRI study from South India. *J Neurol Sci*. 1999;165:133–8.
108. Pahud BA, Glaser CA, Dekker CL, Arvin AM, Schmid DS. Varicella zoster disease of the central nervous system: epidemiological, clinical, and laboratory features 10 years after the introduction of the varicella vaccine. *J Infect Dis*. 2011;203:316–23.
109. Samile N, Hassan T. Acute disseminated encephalomyelitis in children. A descriptive study in Tehran, Iran. *Saud Med J*. 2007;28:396–9.
110. Sawanyawisuth K, Phuttharak W, Tiamkao S, Boonpila A. MRI findings in acute disseminated encephalomyelitis following varicella infection in an adult. *J Clin NeuroSci*. 2007;14:1230–3.
111. Lowther DE, Chong DL, Ascough S, Ettore A, Ingram RJ, Boyton RJ, et al. Th1 not Th17 cells drive spontaneous MS-like disease despite a functional regulatory T cell response. *Acta Neuropathol*. 2013;126:501–15.
112. Martinez NE, Karlsson F, Sato F, Kawai E, Omura S, Minagar A, et al. Protective and detrimental roles for regulatory T cells in a viral model for multiple sclerosis. *Brain Pathol*. 2014. doi:10.1111/bpa.12119.
113. Mocanu V, Oboroceanu T, Zugun-Eloae F. Current status in vitamin d and regulatory T cells—immunological implications. *Rev Med Chir Soc Med Nat Iasi*. 2013;117:965–73.
114. Sakuishi K, Miyake S, Yamamura T. Role of NK cells and invariant NKT cells in multiple sclerosis. *Results Probl Cell Differ*. 2010;51:127–47.
115. Yamamura T, Sakuishi K, Illes Z, Miyake S. Understanding the behavior of invariant NKT cells in autoimmune diseases. *J Neuroimmunol*. 2007;191:8–15.