Molecular Mimicry as a Mechanism of Autoimmune Disease

Matthew F. Cusick · Jane E. Libbey · Robert S. Fujinami

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Abstract A variety of mechanisms have been suggested as the means by which infections can initiate and/or exacerbate autoimmune diseases. One mechanism is molecular mimicry, where a foreign antigen shares sequence or structural similarities with self-antigens. Molecular mimicry has typically been characterized on an antibody or T cell level. However, structural relatedness between pathogen and self does not account for T cell activation in a number of autoimmune diseases. A proposed mechanism that could have been misinterpreted for molecular mimicry is the expression of dual T cell receptors (TCR) on a single T cell. These T cells have dual reactivity to both foreign and selfantigens leaving the host vulnerable to foreign insults capable of triggering an autoimmune response. In this review, we briefly discuss what is known about molecular mimicry followed by a discussion of the current understanding of dual TCRs. Finally, we discuss three mechanisms, including molecular mimicry, dual TCRs, and chimeric TCRs, by which dual reactivity of the T cell may play a role in autoimmune diseases.

Keywords Molecular mimicry · Autoimmune diseases · Dual T cell receptor · Virus infection · Immunopathology

Chronic autoimmune diseases are the by-product of the immune system recognizing self-antigens as foreign, which can lead to inflammation and destruction of specific tissues and organs (immunopathology) [1]. The impact of these diseases is global and heterogeneous with over 100 million

M. F. Cusick · J. E. Libbey · R. S. Fujinami (🖂)
Department of Pathology, University of Utah,
30 North 1900 East, 3R330 SOM,
Salt Lake City, UT 84132, USA
e-mail: robert.fujinami@hsc.utah.edu



people afflicted with more than 80 different autoimmune diseases [2]. While the etiology of autoimmune diseases is not fully elucidated, the causes are likely based on a combination of hereditary and environmental factors [3]. Although host genetic background contributes to the induction of an immune response to self, epidemiological and molecular evidence implicates infectious agents (viral and bacterial) as the principal environmental insults responsible for the induction of autoimmune diseases (reviewed in [4-6]). Prolonged proinflammatory responses to infections have been associated with the initiation and exacerbation of autoimmune diseases (reviewed in [4, 7, 8]). Inflammation is facilitated by proinflammatory cytokines such as type I interferon (IFN), interleukin (IL)-1β, IL-12, IFN- γ , IL-17, and tumor necrosis factor (TNF)- α (reviewed in [7, 9, 10]). However, these proinflammatory cytokines are critical for clearance of pathogens, suggesting that environmental factors are able to divert the immune response towards immunopathogenesis. Although a number of immune cells are responsible for secreting proinflammatory cytokines, the primary cell types implicated in a vast majority of autoimmune disorders are autoreactive B and T cells, or antibody recognition of self [11]. Although a number of viruses and bacteria have been linked to the initiation of certain autoimmune diseases, identifying a particular virus or bacteria that is solely responsible for the induction of an autoimmune response is rare. This occurrence is due to the potential for multiple infections being involved in priming the immune system and other infections triggering disease, which could explain why no one viral infection has been conclusively linked to the development of immune-mediated autoimmune diseases [7]. However, there are a variety of examples of bacterial infections initiating and exacerbating autoimmune diseases. Streptococcus pyogenes is a gram-positive bacterium which causes group A streptococcal infection that is responsible for a number of diseases. The complications associated with S. pyogenes are rheumatic fever and glomerulonephritis. The infection causes the production of cross-reactive antibodies in response to the bacteria. Antibodies recognize the M protein (virulence factor) and the N-acetyl-β-Dglucosamine (GLcNAc) of S. pvogenes and cross-react with myosin leading to heart damage (reviewed in [8, 12, 13]). Further evidence of molecular mimicry due to the production of cross-reactive antibody includes infection with gram-negative bacteria, such as Klebsiella pneumoniae and Campylobacter jejuni. Infection with K. pneumonia or C. jejuni leads to the production of cross-reactive antibodies able to recognize the self-antigens histocompatibility leukocyte antigen (HLA)-B27 and gangliosides, which induce ankylosing spondylitis and Guillain-Barré syndrome, respectively (reviewed in [8, 14]). Examples of human autoimmune diseases with possible links with molecular mimicry are presented in Table 1.

The immune system has a number of mechanisms that are able to detect foreign pathogens by utilizing the major histocompatibility complex (MHC). This locus encodes the HLA genes and a variety of immune response (Ir) genes, thereby shaping the immune system that protects against pathogens. There are two main types of HLA antigens, HLA class I and class II. The function of HLA class I molecules is to present viral peptides at the surface of an infected cell to a T cell receptor (TCR) on a CD8⁺ T cell. The activation of these CD8+ T cells leads to the killing of the virally infected cell. This role of HLA class I, the identification of cells that are infected, explains why all nucleated cells have the capacity to express these MHC molecules. HLA class II molecules, in comparison, are expressed almost exclusively on the surface of dendritic cells, B lymphocytes, macrophages, endothelial cells, and activated T cells. Functionally, the HLA class II molecules present peptides to the TCR on CD4⁺ helper T cells. The engagement of the TCR by the peptide-MHC complex is necessary for the activation of CD4⁺ and CD8⁺ T cells, thereby leading to an effective adaptive immune response against an invading pathogen [15]. CD4⁺ T cells are central mediators of the adaptive immune response including cytokine secretion and cellular and humoral defenses against a pathogen. The HLA locus is extremely polymorphic leading to a heterogeneous population ensuring propagation of a species against novel pathogens. Unfortunately, this genetic heterogeneity adds to the complexity of identifying HLA genes implicated in autoimmune diseases.

In addition to its role in protection against pathogens, a second critical role of the MHC and Ir genes is to safeguard against self-reactivity by restriction of the immune response to self. In this regard, the immune system has developmental checkpoints for the maturation of a T cell. As a naïve T

cell expressing a pre-TCR migrates from the bone marrow to the thymus, rearrangement of α and β TCR genes occurs and T cells that have either too high avidity or lack of recognition of self-antigens are selected against and subsequently programmed for cell death. This selection mechanism for generating mature $\alpha\beta$ TCRs is named central tolerance. Further, peripheral mechanisms of tolerance are able to suppress autoreactive T cells through certain subsets of cells including regulatory T cells (Tregs) that are able to inhibit self-reactive immune cells in the periphery.

Unfortunately, there are a variety of mechanisms including molecular mimicry, bystander activation, exposure of cryptic antigens, and superantigens by which pathogens can aid in the expression of an autoimmune disease [16-21]. Inflammation induced by exposure to a foreign antigen can lead to autoimmune diseases from cross-reactive epitopes (molecular mimicry). These epitopes are segments of foreign antigens which, when presented to either T or B cells in the context of the MHC, can activate CD4⁺ or CD8⁺ T cells. The induction of the immune response and subsequent proinflammatory cytokine release is critical for clearance of a virus or bacteria. However, a sustained proinflammatory response against specific host tissues can occur when there is sequence or structural homology between foreign antigens and selfantigens, termed molecular mimicry [18]. Although this concept has been associated with autoimmunity, there are instances where mimicry (cross-reactivity) provides protection for the host, termed heterologous immunity [22]. Cross-reactivity or mimicry between various strains of viruses or bacteria could help explain how protective immunity arises in certain individuals even in the absence of prior exposure to an emerging pathogen. This example of sequence homology in which molecular mimicry between viruses leads to protective immunity is in contrast to a pathogen mimicking host epitopes (reviewed in [11]).

Brief History of Molecular Mimicry

Over 30 years ago, molecular mimicry by either a virus [18] or bacteria [23] was hypothesized to initiate and exacerbate an autoimmune response through sequence or structural similarities with self-antigens. Currently, molecular mimicry is the prevailing hypothesis as to how viral antigens initiate and maintain autoimmune responses which lead to specific tissue damage [18]. Initial work by Fujinami, Oldstone, and colleagues identified mouse antibodies to measles virus and herpes simplex virus (HSV-1) obtained from antibody-secreting B cell clones [18]. These antibodies were reactive to both intermediate filaments of normal cells and the proteins of measles virus and HSV-1,



Table 1 Examples of human autoimmune diseases with possible molecular mimicry as a mechanism

| Human diseases | Target | T cells/Ab | Human antigen mimicked | Organism | Reference(s) |
|--|--|------------------|---|--|---------------------------------------|
| Spondyloarthropathies (SpAs), ankylosing spondylitis, psoriatic arthritis, reactive arthritis and undifferentiated SpA | Lumbar spine and sacroiliac joints | Abs | HLA-B27 | Klebsiella pneumoniae, Shigella, Chlamydia trachomatis, and other gram-negative bacteria | [71–73] |
| Antiphospholipid syndrome | Fetal loss and thromboembolic | Abs | β2-glycoprotein I | Bacteria, viruses, yeast, and tetanus toxin | [74] |
| Autoimmune chronic gastritis (AIG) (gastric atrophy, hypochloridria and pernicious anemia) | Stomach epithelium cells or parietal cell canaliculi | T cell/Abs | H ⁺ , K ⁺ -ATPase, parietal cell canaliculi Helicobacter pylori | Helicobacter pylori | [75] |
| Cogan's syndrome | Eye and ear | Abs | SSA/Ro; (DEP-1/CD148); connexin 26 | Reovirus III major core protein lambda 1 | [92] |
| Autoimmune thrombocytopenic purpura | Platelet | Abs | Platelet; platelet-associated immunoglobulin G (PAIgG) | Helicobacter pylori | [77] |
| Behçet's disease | Eyes, skin, oral cavity, joints, genital system, CNS and blood vessels | T cell | HSP60, HSP65, HSP70, alpha-tropomyosin, S-antigens | Mycobacterial HSP, Plasmodium falciparum | [78–82] |
| Cardiomyopathy (myocarditis) | Heart | T cell/Abs | Cardiac myosin | Coxsackie virus, group A streptococci, chlamydia, or <i>Trypanosoma cruzi</i> | [83] |
| Celiac sprue (celiac disease) | Small intestine | T cell | Transglutaminase | Gliadin (gluten), perinatal infections, adenovirus 12, hepatitis C virus (HCV) | [84, 85] |
| Chagas disease | Heart | T cell | Cardiac myosin | Trypanosoma cruzi B13 protein | [86, 87] |
| Chronic inflammatory demyelinating polyneuropathy | Schwann cells | Abs | Monosialoganglioside GM2 | Melanoma, Campylobacter jejuni | [88, 89] |
| Crohn's disease | Gastrointestinal tract | T cell | Unknown | Gram-positive bacterial peptidoglycans | [60] |
| Dermatomyositis (juvenile) | Skin and muscle | T cell | Skeletal myosin | Streptococcus pyogenes M5 protein | [91] |
| Essential mixed cryoglobulinemia | B cell | Abs | IgG-Fc | HCV | [92] |
| Guillain-Barré syndrome | Gangliosides and peripheral nerve | Abs | Peripheral nerve | Campylobacter jejuni | [63] |
| Insulin dependent diabetes (type I) | Pancreas | T cell | Islet antigens (GAD 65, proinsulin carboxypeptidase H) | Coxsackie B virus, rubella, rotavirus, herpes, rhinovirus, hantavirus, flavivirus and retrovirus | [94–96], (reviewed in [97]); [98–100] |
| Systemic lupus erythematosus | Systemic | Abs | 60 Kda Ro | Epstein-Barr virus (EBV nuclear antigen-1) | [101] |
| Multiple sclerosis | Myelin | T cell | Myelin basic protein | EBV, measles and HHV-6 | [11, 35, 102] |
| Primary biliary cirrhosis | Liver (intrahepatic bile ducts) | Abs/B and T cell | PDE2, GP210, human pyruvate dehydrogenase complex-E2 (PDC-E2), HLA-DR | Gram-negative bacterium, Escherichia coli, Helicobacter pylori, Pseudomonas aeruginosa, cytomegalovirus, and Haemophilus influenza | [103–107] |
| Psoriasis | Skin | T cell | Epidermal keratins | Streptococcus pyogenes (streptococcal M protein) | [108] |
| Rheumatic fever | Неап | Abs/F cell | Cardiac myosin | M protein (major virulence factor of group A streptococci) and streptococcus carbohydrate epitope GlcNAc | [12, 109–111] |
| Rasmussen's encephalitis | CNS | Abs | Antiglutamate receptor (GLUR3) | Microorganisms | [112, 113] |
| Acute disseminating encephalomyelitis | CNS | T cell | Myelin basic protein | Measles virus, rabies vaccine, HHV-6, coronavirus, influenza virus hemagglutinin, EBV, Semliki Forest virus | [114, 115], reviewed in [116] |



Reference(s) 122, 123] 125, 126] 128, 129] [124] [121] 127 [120]Human cytomegalovirus (hCMV) Herpes simplex virus type 1 gpD Herpes simplex virus-type Goup A streptococcus hCMV UL94 protein Borrelia burgdorferi Helicobacter pylori hCMV (pUL57) Coxsackie virus Organism Viruses Acetylcholine receptor, neurofilaments calmodulin-dependent protein (CaM) 5-Antigen, interphotoreceptor binding HLA-DR, CD13 (aminopeptidase N) **Human leukocyte function-associated** NAG-2 (tetraspan novel antigen-2) 3-Tubulin, GlcNAc, calcium/ **Human** antigen mimicked Gastric mucosa antigens antigen-1 (hLFA-1) (Lewis antigens) protein (IRBP) Corneal tissue 3060 kD F cell/Abs cells/Ab T cell T cell Abs Solid organ transplant Eye and pineal gland Neurons and β cells Endothelial cells Gastric mucosa in the brain Systemic Joints Brain Herpes stromal keratitis [able 1 (continued) Stiff-person syndrome Graft vs host disease Autoimmune uveitis Sjögren's syndrome Sydenham's chorea Myasthenia gravis Peptic/gastric ulcer Human diseases Lyme arthritis Scleroderma

thereby demonstrating a relatedness between host and viral antigens [18]. Further work by Fujinami and Oldstone used myelin basic protein (MBP), a nerve sheath protein containing an encephalitogenic T cell epitope in rabbits. The hepatitis B virus polymerase (HBVP) protein was found through computer analysis to share six consecutive amino acids with the encephalitogenic MBP epitope [16], and when rabbits were sensitized with either MBP or HBV peptides, the rabbit's tissue serum reacted against MBP. Further, rabbits sensitized with the HBVP peptide developed central nervous system (CNS) pathology similar to rabbits sensitized with whole MBP protein or the MBP peptide [16]. Importantly, the rabbits sensitized with HBVP did not contract hepatitis but still developed encephalomyelitis and presented with a similar pathology as MBPsensitized mice. These experiments were the first experimental demonstration of molecular mimicry, whereby a microbial peptide with similar amino acid sequences to the self-peptide was able to activate autoreactive T cells and subsequently cause specific tissue damage.

Relationship Between Molecular Mimicry and Autoimmune Diseases

Immune cells of the adaptive immune response are specifically activated, but the hallmark of autoimmunity is the dysregulation of the immune system, especially T and B cells recognizing self-antigens as foreign. The ability of T cells to evade central (thymic selection) and peripheral (Tregs) mechanisms of tolerance is evident by the large number of T cell-mediated human autoimmune diseases, such as type-1 diabetes, systemic lupus erythematosus, rheumatoid arthritis, and multiple sclerosis (MS) [24-28]. Molecular mimicry has been implicated in the pathogenesis of many of these autoimmune diseases including MS, spondyloarthropathies, Graves' disease, and diabetes mellitus [16, 29, 30]. In the case of MS, it has been hypothesized that certain viruses, such as Epstein-Barr virus (EBV), share sequence homology with antigenic structures in the CNS [31].

Activation of an autoimmune response could be enhanced by a variety of other, albeit, non-mutually exclusive non-specific mechanisms including bystander activation and superantigens. The difference between other non-specific mechanisms that initiate autoimmunity and molecular mimicry is that microbial mimics specifically direct the immune response towards a tissue and/or organ. Originally, T cell recognition was postulated to be highly specific and cross-reactivity was thought to be a rare phenomenon. However, the structural requirements for peptide binding by MHC class II molecules that are presented to T cells were found to be based on amino



acid properties, and amino acids sharing similar chemical features were able to bind at the same MHC peptide binding groove, thereby demonstrating that binding motifs were degenerate with only a small sequence needed for TCR recognition [32-34]. An illustration of TCR degeneracy was shown by Wucherpfennig and Strominger [35] using chemically related synthetic peptides mimicking the MBP(85-99) epitope that were incubated with human MBP(85-99)-specific T cell clones which were then tested for reactivity. Of the T cell clones that responded to the synthetic peptides, only eight of the 129 synthetic peptides were recognized by the T cell clones, and only one of the synthetic peptides that induced a response was clearly similar to MBP(85-99) [35, 36]. Therefore, these studies clearly demonstrate TCRs binding to a spectrum of specific peptides that is based upon structural relatedness, termed poly-specificity [37]. This flexibility exhibited by TCR binding and the existence of pathogens that share sequence or structural similarities with self-antigens could be one reason why investigators have been unable to conclusively associate a specific virus with autoimmune diseases, such as MS (reviewed in [4]).

Linear sequence matches in amino acid motifs is not the only criteria for mimicry [32]. It has been hypothesized that self-reactive immune cells are primed by molecular mimicry and bystander activation, thereby sensitizing the immune cells and leading to a "fertile field" but no apparent disease. Subsequent environmental insults could induce these sensitized autoreactive cells to cause an autoimmune disease. Work from our laboratory demonstrated that recombinant viruses having molecular mimicry with self-CNS antigens were unable to initiate an autoimmune disease individually [38]. However, infected mice that were subsequently challenged, after viral clearance, with a non-specific immunologic insult developed disease [38]. Further, subsequent experiments showed that conventional inflammatory responses to specific pathogens were able to induce disease in animals primed with a molecular mimic to a CNS antigen [39]. Therefore, not only is the priming of the immune system necessary for an autoimmune disease but the milieu to which the primed immune cells are exposed is an important factor in initiating an autoimmune disease. Animal models of various autoimmune diseases have explored the role of molecular mimicry as a contributing factor (Table 2).

The use of transgenic (tg) mice expressing virus proteins as transgenes in specific organs has been an important model for providing evidence for molecular mimicry. The expression of lymphocytic choriomeningitis virus (LCMV) viral antigens in pancreatic islet cells and the subsequent cross of this tg mouse with a TCR-tg mouse specific for LCMV glycoprotein resulted in an animal that only developed autoimmune disease if virally infected [40, 41]. These results demonstrated that "self'-reactive T cells are

present in the periphery and the immune cells appear to remain quiescent until an appropriate signal (viral infection) triggers the T cells to respond.

Dual TCR and How This Impacts Our Interpretation of Molecular Mimicry

There are a variety of non-mutually exclusive factors that lead to a fully activated T cell, such as the quantity of peptide-MHC presented on the surface of antigenpresenting cells and TCR avidity. The interaction between the peptide-MHC and TCR is critical for the initiation of an adaptive immune response and clearance of a pathogen [15]. In order for T cells to reach maturity, the T cell goes through a number of developmental checkpoints leading to somatic recombination of various gene segments. The TCR α - and β -chains are generated by V-D-J recombination, which leads to $\alpha\beta$ TCRs expressed on the surface of T cells [42, 43]. Although it was believed that T cell signaling was mediated by a single antigen receptor, recent evidence demonstrates that T cells are capable of expressing functional dual V\alpha TCRs at a frequency of approximately 30% in humans and 15% in mice; however, an accurate number of dual specific TCRs is lacking due to the limited availability of anti-V\alpha monoclonal antibodies (mAbs) [44– 46]. Interestingly, in contrast to the high frequency of dual expressing Va T cells, only 1% of humans and 5-7% of mice express two β-chains due to allelic exclusion mechanisms, but the frequencies of dual VB TCRs have been found to be higher with age and in TCR-tg mice [47-49]. Expression of multiple TCR Vαs on the surface of a T cell is the result of simultaneous rearrangement of both TCRα loci during thymocyte development [50–52]. Further, TCR V β -chains preferentially bind to certain V α chains leading to differential expression of chimeric TCRs on the surface of T cells [51, 53, 54].

Due to the heterogeneity of TCRs normally expressed in the periphery of humans and mice, TCR-tg mice have been used to track and determine the fate of T cells expressing dual TCRs. The use of TCR-tg mice has led to the identification of a potential role for dual TCRs in a variety of conditions including graft-versus-host disease, human immunodeficiency virus infection, inflammatory bowel disease, T cell leukemia, T cell lymphoma, and MS [55–61].

The expression of dual TCRs by the same T cell has been proposed to be a potential mechanism for autoimmune disease. Normally, high avidity self-reactive T cells are thymically depleted, but it has been hypothesized that the expression of a self-TCR on a T cell is lower when presented in the context of a second TCR, thereby providing a cover for high avidity self-TCRs from both central and peripheral tolerance. Blichfeldt et al. [62]



Table 2 Examples of murine models of autoimmune diseases where molecular mimicry is proposed as a mechanism

| Human autoimmune disease | Mouse strain | Initiating agent(s) | Reference(s) |
|-------------------------------------|--|--|----------------|
| Behçet's disease | ICR mice | HSV type 1 (F strain) inoculation in ear lobe | [131] |
| Myocarditis | BALB/c | Mouse cytomegalovirus (MCMV) | [132] |
| Insulin-dependent diabetes (type I) | Tg mice expressing LCMV protein in pancreas | Pichinde virus infection of mice | [133] |
| Guillain-Barré syndrome | BALB/c | lipooligosaccharide of Brucella melitensis | [134] |
| Autoimmune hepatitis type 2 | FVB | infection with recombinant adenovirus encoding human cytochome CYP2D6 | [135, 136] |
| Herpes stromal keratitis | C.AL-20 | HSV-1 | [120, 137] |
| Autoimmune uveitis | C3H/HeN | Salmonella typhimurium | [138, 139] |
| Sjögren's syndrome | C57BL/6; [B6]+/+; Fas-deficient B6-lpr/lpr; TNFR1-deficient B6; and TNFR1-deficient lpr/lpr) | MCMV | [140] |
| Multiple sclerosis | SJL/J C57BL/6 | Theiler's murine encephalomyelitis virus Semliki Forest virus infection | [141] [142] |

demonstrated that dual tg-TCRs, which have lower expression of each TCR on the surface of a T cell, needed higher concentrations of peptide, presented by MHC, to induce a similar T cell proliferative response compared to a single receptor T cell.

A potential role of dual TCRs in autoimmunity is in the rescue of autoreactive T cells from thymic selection. For example, the double tg mouse for autoimmune diabetes, in which the mice express a TCR specific for peptide 111–119 of hemagglutinin (HA) (TCR-HA) under the control of the rat insulin promoter and develop spontaneous diabetes and insulitis [63], were used to determine how T cells could escape tolerance mechanisms even if the antigen was ubiquitously expressed [64]. Low expressing TCR-HA coexpressing T cells were more effective at transferring diabetes than TCR-HA high dual TCRs, suggesting that the surface level expression of a dual TCR can be modulated by a second TCR expressed on the same T cell, thus "escape" of autoreactive T cells could be the first step in an autoimmune disease.

The "trigger" of an autoimmune disease could be linked to environmental insults, such as viruses. A T cell co-expressing TCRs specific for a self-antigen and a foreign antigen could potentially allow for autoreactive T cells to be activated if the host is exposed to that foreign antigen. The activation of a subset of T cells could than lead to tolerance being broken and the initiation of an autoimmune disease if these T cells experienced a particular organ or tissue that expressed the self-antigen for the other TCR expressed at the surface of the T cell. In support of a role for dual TCRs in autoimmune diseases, work performed in our laboratory characterized autoreactive CD8⁺ T cells isolated from the spleens of Theiler's murine encephalomyelitis virus (TMEV)-infected SJL/J mice [65]. In vitro assays testing CD8⁺ T

cell killing activity found a population of CD8⁺ T cells that killed uninfected syngeneic cells [65]. Adoptively transferring these TMEV-specific autoreactive CD8⁺ T cells into non-infected SJL/J mice caused CNS pathology [65]. Further support for the importance of the mechanism by which viral infection could induce an autoimmune disease through dual TCR-expressing T cells was performed by Ji et al. [61] using MBP(79-87) TCR-tg mice [66]. Cytometric phenotyping, in vitro CD8⁺ T cell killing assays, and adoptive transfer experiments were used to track the expansion and killing capacity of Vα8Vβ8 MBP (79-87)-specific TCR and Vα8Vβ6-vaccinia virusspecific TCR. Infection of these tg mice with vaccinia virus induced autoimmune disease, thus demonstrating a virus triggering an autoimmune disease through dual TCR expressing T cells [61]. Although several tg TCR β-chains have been described on peripheral T cell [61, 67–70], there is no evidence that co-expression of dual TCRs leads to autoimmunity without the use of TCR-tg mice. As described above, current work in our laboratory has characterized TMEV-specific autoreactive CD8⁺ T cell clones derived from a wild-type animal, and these autoreactive TMEV-specific T cell clones express dual TCRs (manuscript in preparation). Importantly, we were able to induce CNS pathology in naïve SJL/J mice by adoptively transferring the TMEV-specific clones. Although further work is needed in order to identify the self-antigen that activates these CD8⁺ T cells, to our knowledge these results are the first demonstration of an autoimmune disease initiated by a dual expressing TCR characterized in the virus' natural host.

Taken together, three possible mechanisms could explain how the dual reactivity of the TCR may play a role in autoimmune diseases (manuscript in preparation). The first mechanism is molecular mimicry, whereby the induction of



an autoimmune response to self is due to a single TCR recognizing both a virus and a self-antigen. The second mechanism is the expression of dual TCRs on a single T cell, where one TCR is able to recognize a microbial antigen and the other TCR recognizes self. The third mechanism involves a T cell expressing chimeric TCRs generated from either a single $V\alpha$ combining with two different $V\beta$ s or a single $V\beta$ combining with two different $V\alpha$ s, resulting in a T cell with the potential of expressing two different chimeric TCRs specific for a self-antigen and a foreign antigen.

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References

- Abou-Raya A, Abou-Raya S (2006) Inflammation: a pivotal link between autoimmune diseases and atherosclerosis. Autoimmun Rev 5:331–337
- Selgrade MK, Cooper GS, Germolec DR, Heindel JJ (1999) Linking environmental agents and autoimmune disease: an agenda for future research. Environ Health Perspect 107(Suppl 5):811–813
- von Herrath MG, Fujinami RS, Whitton JL (2003) Microorganisms and autoimmunity: making the barren field fertile? Nat Rev Microbiol 1:151–157
- Libbey JE, Fujinami RS (2010) Potential triggers of MS. Results Probl Cell Differ 51:21–42
- Fujinami RS (2001) Viruses and autoimmune disease—two sides of the same coin? Trends Microbiol 9:377–381
- Ascherio A, Munger KL (2007) Environmental risk factors for multiple sclerosis. Part II: noninfectious factors. Ann Neurol 61:504–513
- McCoy L, Tsunoda I, Fujinami RS (2006) Multiple sclerosis and virus induced immune responses: autoimmunity can be primed by molecular mimicry and augmented by bystander activation. Autoimmunity 39:9–19
- Sfriso P, Ghirardello A, Botsios C et al (2010) Infections and autoimmunity: the multifaceted relationship. J Leukoc Biol 87:385–395
- Trinchieri G (2010) Type I interferon: friend or foe? J Exp Med 207:2053–2062
- Iwakura Y, Ishigame H, Saijo S, Nakae S (2011) Functional specialization of interleukin-17 family members. Immunity 34:149–162
- Libbey JE, McCoy LL, Fujinami RS (2007) Molecular mimicry in multiple sclerosis. Int Rev Neurobiol 79:127–147
- Whitton JL, Feuer R (2004) Myocarditis, microbes and autoimmunity. Autoimmunity 37:375–386
- 13. Libbey JE, Fujinami RS (2010) Role for antibodies in altering behavior and movement. Autism Res 3:147–152
- Shahrizaila N, Yuki N (2011) Guillain–Barré syndrome animal model: the first proof of molecular mimicry in human autoimmune disorder. J Biomed Biotechnol 2011:829129
- Ahmed R (1992) Immunological memory against viruses. Semin Immunol 4:105–109
- Fujinami RS, Oldstone MBA (1985) Amino acid homology between the encephalitogenic site of myelin basic protein and virus: Mechanism for autoimmunity. Science 230:1043–1045

- 17. Oldstone MBA (1987) Molecular mimicry and autoimmune disease. Cell 50:819–820
- Fujinami RS, Oldstone MBA, Wroblewska Z, Frankel ME, Koprowski H (1983) Molecular mimicry in virus infection: crossreaction of measles virus phosphoprotein or of herpes simplex virus protein with human intermediate filaments. Proc Natl Acad Sci USA 80:2346–2350
- McRae BL, Vanderlugt CL, Dal Canto MC, Miller SD (1995) Functional evidence for epitope spreading in the relapsing pathology of experimental autoimmune encephalomyelitis. J Exp Med 182:75–85
- Vanderlugt CL, Begolka WS, Neville KL et al (1998) The functional significance of epitope spreading and its regulation by co-stimulatory molecules. Immunol Rev 164:63–72
- Scherer MT, Ignatowicz L, Winslow GM, Kappler JW, Marrack P (1993) Superantigens: Bacterial and viral proteins that manipulate the immune system. Annu Rev Cell Biol 9:101–128
- 22. Chen HD, Fraire AE, Joris I, Brehm MA, Welsh RM, Selin LK (2001) Memory CD8⁺ T cells in heterologous antiviral immunity and immunopathology in the lung. Nat Immunol 2:1067–1076
- Zabriskie JB, Freimer EH (1966) An immunological relationship between the group A streptococcus and mammalian muscle. J Exp Med 124:661–678
- 24. Gregersen PK, Silver J, Winchester RJ (1987) The shared epitope hypothesis. An approach to understanding the molecular genetics of susceptibility to rheumatoid arthritis. Arthritis Rheum 30:1205–1213
- Burns J, Rosenzweig A, Zweiman B, Lisak RP (1983) Isolation of myelin basic protein-reactive T-cell lines from normal human blood. Cell Immunol 81:435–440
- Mathis D, Vence L, Benoist C (2001) β-Cell death during progression to diabetes. Nature 414:792–798
- Tisch R, McDevitt H (1996) Insulin-dependent diabetes mellitus. Cell 85:291–297
- Bertsias GK, Salmon JE, Boumpas DT (2010) Therapeutic opportunities in systemic lupus erythematosus: state of the art and prospects for the new decade. Ann Rheum Dis 69:1603–1611
- Ebringer A, Baines M, Ptaszynska T (1985) Spondyloarthritis, uveitis, HLA-B27 and Klebsiella. Immunol Rev 86:101–116
- Quaratino S, Thorpe CJ, Travers PJ, Londei M (1995) Similar antigenic surfaces, rather than sequence homology, dictate Tcell epitope molecular mimicry. Proc Natl Acad Sci USA 92:10398–10402
- 31. Wandinger K-P, Jabs W, Siekhaus A et al (2000) Association between clinical disease activity and Epstein-Barr virus reactivation in MS. Neurology 55:178-184
- Sinigaglia F, Hammer J (1994) Defining rules for the peptide— MHC class II interaction. Curr Opin Immunol 6:52–56
- 33. Wucherpfennig KW, Sette A, Southwood S et al (1994) Structural requirements for binding of an immunodominant myelin basic protein peptide to DR2 isotypes and for its recognition by human T cell clones. J Exp Med 179:279–290
- 34. Reay PA, Kantor RM, Davis MM (1994) Use of global amino acid replacements to define the requirements for MHC binding and T cell recognition of moth cytochrome c (93–103). J Immunol 152:3946–3957
- Wucherpfennig KW, Strominger JL (1995) Molecular mimicry in T cell-mediated autoimmunity: viral peptides activate human T cell clones specific for myelin basic protein. Cell 80:695–705
- Hausmann S, Martin M, Gauthier L, Wucherpfennig KW (1999) Structural features of autoreactive TCR that determine the degree of degeneracy in peptide recognition. J Immunol 162:338–344
- Wucherpfennig KW, Allen PM, Celada F et al (2007) Polyspecificity of T cell and B cell receptor recognition. Semin Immunol 19:216–224



- Theil DJ, Tsunoda I, Rodriguez F, Whitton JL, Fujinami RS (2001) Viruses can silently prime for and trigger central nervous system autoimmune disease. J Neurovirol 7:220–227
- Tsunoda I, Libbey JE, Fujinami RS (2007) Sequential polymicrobial infections lead to CNS inflammatory disease: possible involvement of bystander activation in heterologous immunity. J Neuroimmunol 188:22–33
- Ohashi PS, Oehen S, Buerki K et al (1991) Ablation of "tolerance" and induction of diabetes by virus infection in viral antigen transgenic mice. Cell 65:305–317
- Oldstone MBA, Nerenberg M, Southern P, Price J, Lewicki H (1991) Virus infection triggers insulin-dependent diabetes mellitus in a transgenic model: role of anti-self (virus) immune response. Cell 65:319–331
- Petrie HT, Livak F, Schatz DG, Strasser A, Crispe IN, Shortman K (1993) Multiple rearrangements in T cell receptor α chain genes maximize the production of useful thymocytes. J Exp Med 178:615–622
- Pang SS, Berry R, Chen Z et al (2010) The structural basis for autonomous dimerization of the pre-T-cell antigen receptor. Nature 467:844–848
- 44. Casanova J-L, Romero P, Widmann C, Kourilsky P, Maryanski JL (1991) T cell receptor genes in a series of class I major histocompatibility complex-restricted cytotoxic T lymphocyte clones specific for a *Plasmodium berghei* nonapeptide: implications for T cell allelic exclusion and antigen-specific repertoire. J Exp Med 174:1371–1383
- Padovan E, Casorati G, Dellabona P, Meyer S, Brockhaus M, Lanzavecchia A (1993) Expression of two T cell receptor α chains: dual receptor T cells. Science 262:422–424
- 46. Corthay A, Nandakumar KS, Holmdahl R (2001) Evaluation of the percentage of peripheral T cells with two different T cell receptor α-chains and of their potential role in autoimmunity. J Autoimmun 16:423–429
- Davodeau F, Peyrat M-A, Romagné F et al (1995) Dual T cell receptor β chain expression on human T lymphocytes. J Exp Med 181:1391–1398
- 48. Padovan E, Giachino C, Cella M, Valitutti S, Acuto O, Lanzavecchia A (1995) Normal T lymphocytes can express two different T cell receptor β chains: implications for the mechanism of allelic exclusion. J Exp Med 181:1587–1591
- Munthe LA, Blichfeldt E, Sollien A, Dembic Z, Bogen B (1996)
 T cells with two Tcrβ chains and reactivity to both MHC/ idiotypic peptide and superantigen. Cell Immunol 170:283–290
- Alam SM, Crispe IN, Gascoigne NR (1995) Allelic exclusion of mouse T cell receptor α chains occurs at the time of thymocyte TCR up-regulation. Immunity 3:449–458
- Marolleau J-P, Fondell JD, Malissen M et al (1988) The joining of germ-line Vα to Jα genes replaces the preexisting Vα-Jα complexes in a T cell receptor α, β positive T cell line. Cell 55:291–300
- 52. Malissen M, Trucy J, Letourneur F et al (1988) A T cell clone expresses two T cell receptor α genes but uses one αβ heterodimer for allorecognition and self MHC-restricted antigen recognition. Cell 55:49–59
- Saito T, Sussman JL, Ashwell JD, Germain RN (1989) Marked differences in the efficiency of expression of distinct αβ T cell receptor heterodimers. J Immunol 143:3379–3384
- Vacchio MS, Granger L, Kanagawa O et al (1993) T cell receptor Vα-Vβ combinatorial selection in the expressed T cell repertoire. J Immunol 151:1322–1327
- Morris GP, Allen PM (2009) Cutting edge: highly alloreactive dual TCR T cells play a dominant role in graft-versus-host disease. J Immunol 182:6639–6643
- 56. Taupin J-L, Halary F, Dechanet J et al (1999) An enlarged subpopulation of T lymphocytes bearing two distinct $\gamma\delta$ TCR in an HIV-positive patient. Int Immunol 11:545–552

- Söderström K, Bucht A, Halapi E, Grönberg A, Magnusson I, Kiessling R (1996) Increased frequency of abnormal γδ T cells in blood of patients with inflammatory bowel diseases. J Immunol 156:2331–2339
- 58. Hinz T, Marx S, Nerl C, Kabelitz D (1996) Clonal expansion of $\gamma\delta$ T cells expressing two distinct T-cell receptors. Br J Haematol 94:62–64
- 59. Boehrer S, Hinz T, Schui D et al (2001) T-large granular lymphocyte leukaemia with natural killer cell-like cytotoxicity and expression of two different α -and β -T-cell receptor chains. Br J Haematol 112:201–203
- 60. Weidmann E, Hinz T, Klein S et al (2000) Cytotoxic hepatosplenic $\gamma\delta$ T-cell lymphoma following acute myeloid leukemia bearing two distinct γ chains of the T-cell receptor. Biologic and clinical features. Haematologica 85:1024–1031
- Ji Q, Perchellet A, Goverman JM (2010) Viral infection triggers central nervous system autoimmunity via activation of CD8⁺ T cells expressing dual TCRs. Nat Immunol 11:628–634
- 62. Blichfeldt E, Munthe LA, Rotnes JS, Bogen B (1996) Dual T cell receptor T cells have a decreased sensitivity to physiological ligands due to reduced density of each T cell receptor. Eur J Immunol 26:2876–2884
- 63. Sarukhan A, Lanoue A, Franzke A, Brousse N, Buer J, von Boehmer H (1998) Changes in function of antigen-specific lymphocytes correlating with progression towards diabetes in a transgenic model. EMBO J 17:71–80
- 64. Sarukhan A, Garcia C, Lanoue A, von Boehmer H (1998) Allelic inclusion of T cell receptor α genes poses an autoimmune hazard due to low-level expression of autospecific receptors. Immunity 8:563–570
- 65. Tsunoda I, Kuang L-Q, Kobayashi-Warren M, Fujinami RS (2005) Central nervous system pathology caused by autoreactive CD8⁺ T cell clones following virus infection. J Virol 79:14640– 14646
- Perchellet A, Stromnes I, Pang JM, Goverman J (2004) CD8⁺ T cells maintain tolerance to myelin basic protein by 'epitope theft' Nat Immunol 5:606–614
- 67. Borgulya P, Kishi H, Uematsu Y, von Boehmer H (1992) Exclusion and inclusion of α and β T cell receptor alleles. Cell 69:529–537
- Balomenos D, Balderas RS, Mulvany KP, Kaye J, Kono DH, Theofilopoulos AN (1995) Incomplete T cell receptor Vβ allelic exclusion and dual Vβ-expressing cells. J Immunol 155:3308–3312
- Hurst SD, Sitterding SM, Ji S, Barrett TA (1997) Functional differentiation of T cells in the intestine of T cell receptor transgenic mice. Proc Natl Acad Sci USA 94:3920–3925
- 70. Heath WR, Miller JFAP (1993) Expression of two α chains on the surface of T cells in T cell receptor transgenic mice. J Exp Med 178:1807–1811
- Stagg AJ, Breban M, Hammer RE, Knight SC, Taurog JD (1995)
 Defective dendritic cell (DC) function in a HLA-B27 transgenic rat model of spondyloarthropathy (SpA). Adv Exp Med Biol 378:557–559
- Ebringer R, Cawdell D, Ebringer A (1979) Klebsiella pneumoniae and acute anterior uveitis in ankylosing spondylitis. Br Med J 1:383
- 73. Schwimmbeck PL, Yu DTY, Oldstone MBA (1987) Autoantibodies to HLA B27 in the sera of HLA B27 patients with ankylosing spondylitis and Reiter's syndrome. Molecular mimicry with *Klebsiella pneumoniae* as potential mechanism of autoimmune disease. J Exp Med 166:173–181
- Shoenfeld Y, Blank M, Cervera R, Font J, Raschi E, Meroni PL (2006) Infectious origin of the antiphospholipid syndrome. Ann Rheum Dis 65:2–6
- 75. Amedei A, Bergman MP, Appelmelk BJ et al (2003) Molecular mimicry between *Helicobacter pylori* antigens and H+, K+-



- adenosine triphosphatase in human gastric autoimmunity. J Exp Med 198:1147-1156
- Lunardi C, Bason C, Leandri M et al (2002) Autoantibodies to inner ear and endothelial antigens in Cogan's syndrome. Lancet 360:915–921
- 77. Takahashi T, Yujiri T, Shinohara K et al (2004) Molecular mimicry by *Helicobacter pylori* CagA protein may be involved in the pathogenesis of *H. pylori*-associated chronic idiopathic thrombocytopenic purpura. Br J Haematol 124:91–96
- Direskeneli H, Hasan A, Shinnick T et al (1996) Recognition of B-cell epitopes of the 65 kDa HSP in Behçet's disease. Scand J Immunol 43:464–471
- de Smet MD, Ramadan A (2001) Circulating antibodies to inducible heat shock protein 70 in patients with uveitis. Ocul Immunol Inflamm 9:85–92
- 80. Suzuki Y, Hoshi K, Matsuda T, Mizushima Y (1992) Increased peripheral blood $\gamma \delta^+$ T cells and natural killer cells in Behçet's disease. J Rheumatol 19:588–592
- Mahesh SP, Li Z, Buggage R et al (2005) Alpha tropomyosin as a self-antigen in patients with Behçet's disease. Clin Exp Immunol 140:368–375
- 82. de Smet MD, Dayan M (2000) Prospective determination of T-cell responses to S-antigen in Behçet's disease patients and controls. Invest Ophthalmol Vis Sci 41:3480–3484
- 83. Cunningham MW (2004) T cell mimicry in inflammatory heart disease. Mol Immunol 40:1121–1127
- Dieterich W, Ehnis T, Bauer M et al (1997) Identification of tissue transglutaminase as the autoantigen of celiac disease. Nat Med 3:797–801
- Plot L, Amital H (2009) Infectious associations of Celiac disease. Autoimmun Rev 8:316–319
- 86. Iwai LK, Juliano MA, Juliano L, Kalil J, Cunha-Neto E (2005) T-cell molecular mimicry in Chagas disease: identification and partial structural analysis of multiple cross-reactive epitopes between *Trypanosoma cruzi* B13 and cardiac myosin heavy chain. J Autoimmun 24:111–117
- 87. Cunha-Neto E, Bilate AM, Hyland KV, Fonseca SG, Kalil J, Engman DM (2006) Induction of cardiac autoimmunity in Chagas heart disease: a case for molecular mimicry. Autoimmunity 39:41–54
- Weiss MD, Luciano CA, Semino-Mora C, Dalakas MC, Quarles RH (1998) Molecular mimicry in chronic inflammatory demyelinating polyneuropathy and melanoma. Neurology 51:1738–1741
- Köller H, Kieseier BC, Jander S, Hartung H-P (2005) Chronic inflammatory demyelinating polyneuropathy. N Engl J Med 352:1343–1356
- 90. Klasen IS, Melief MJ, van Halteren AGS et al (1994) The presence of peptidoglycan–polysaccharide complexes in the bowel wall and the cellular responses to these complexes in Crohn's disease. Clin Immunol Immunopathol 71:303–308
- 91. Massa M, Costouros N, Mazzoli F et al (2002) Self epitopes shared between human skeletal myosin and *Streptococcus pyogenes* M5 protein are targets of immune responses in active juvenile dermatomyositis. Arthritis Rheum 46:3015–3025
- De Re V, Sansonno D, Simula MP et al (2006) HCV-NS3 and IgG-Fc crossreactive IgM in patients with type II mixed cryoglobulinemia and B-cell clonal proliferations. Leukemia 20:1145–1154
- Yuki N, Tagawa Y, Handa S (1996) Autoantibodies to peripheral nerve glycosphingolipids SPG, SLPG, and SGPG in Guillain-Barré syndrome and chronic inflammatory demyelinating polyneuropathy. J Neuroimmunol 70:1–6
- Atkinson MA, Bowman MA, Campbell L, Darrow BL, Kaufman DL, Maclaren NK (1994) Cellular immunity to a determinant common to glutamate decarboxylase and coxsackie virus in insulin-dependent diabetes. J Clin Invest 94:2125–2129

- Kaufman DL, Erlander MG, Clare-Salzler M, Atkinson MA, Maclaren NK, Tobin AJ (1992) Autoimmunity to two forms of glutamate decarboxylase in insulin-dependent diabetes mellitus. J Clin Invest 89:283–292
- 96. Ou D, Mitchell LA, Metzger DL, Gillam S, Tingle AJ (2000) Cross-reactive rubella virus and glutamic acid decarboxylase (65 and 67) protein determinants recognised by T cells of patients with type I diabetes mellitus. Diabetologia 43:750–762
- 97. van der Werf N, Kroese FGM, Rozing J, Hillebrands J-L (2007) Viral infections as potential triggers of type 1 diabetes. Diabetes Metab Res Rev 23:169–183
- Baum H, Brusic V, Choudhuri K, Cunningham P, Vergani D, Peakman M (1995) MHC molecular mimicry in diabetes. Nat Med 1:388
- Jones DB, Armstrong NW (1995) Coxsackie virus and diabetes revisited. Nat Med 1:284
- 100. Endl J, Otto H, Jung G et al (1997) Identification of naturally processed T cell epitopes from glutamic acid decarboxylase presented in the context of HLA-DR alleles by T lymphocytes of recent onset IDDM patients. J Clin Invest 99:2405–2415
- 101. McClain MT, Heinlen LD, Dennis GJ, Roebuck J, Harley JB, James JA (2005) Early events in lupus humoral autoimmunity suggest initiation through molecular mimicry. Nat Med 11:85–89
- 102. Fujinami RS, von Herrath MG, Christen U, Whitton JL (2006) Molecular mimicry, bystander activation, or viral persistence: infections and autoimmune disease. Clin Microbiol Rev 19:80–94
- 103. Baum H (1995) Mitochondrial antigens, molecular mimicry and autoimmune disease. Biochim Biophys Acta 1271:111–121
- 104. Shimoda S, Nakamura M, Ishibashi H et al (2003) Molecular mimicry of mitochondrial and nuclear autoantigens in primary biliary cirrhosis. Gastroenterology 124:1915–1925
- 105. Kita H, Matsumura S, He X-S et al (2002) Analysis of TCR antagonism and molecular mimicry of an HLA-A*0201restricted CTL epitope in primary biliary cirrhosis. Hepatology 36:918–926
- Selmi C, Bowlus CL, Gershwin ME, Coppel RL (2011) Primary biliary cirrhosis. Lancet 377:1600–1609
- 107. Bogdanos D-P, Baum H, Grasso A et al (2004) Microbial mimics are major targets of crossreactivity with human pyruvate dehydrogenase in primary biliary cirrhosis. J Hepatol 40:31–39
- 108. Johnston A, Gudjonsson JE, Sigmundsdottir H, Love TJ, Valdimarsson H (2004) Peripheral blood T cell responses to keratin peptides that share sequences with streptococcal M proteins are largely restricted to skin-homing CD8⁺ T cells. Clin Exp Immunol 138:83–93
- 109. Brandt ER, Yarwood PJ, McMillan DJ et al (2001) Antibody levels to the class I and II epitopes of the M protein and myosin are related to group A streptococcal exposure in endemic populations. Int Immunol 13:1335–1343
- Cunningham MW, McCormack JM, Talaber LR et al (1988)
 Human monoclonal antibodies reactive with antigens of the group A Streptococcus and human heart. J Immunol 141:2760–2766
- 111. Adderson EE, Shikhman AR, Ward KE, Cunningham MW (1998) Molecular analysis of polyreactive monoclonal antibodies from rheumatic carditis: human anti-N-acetylglucosamine/anti-myosin antibody V region genes. J Immunol 161:2020–2031
- 112. Rogers SW, Andrews PI, Gahring LC et al (1994) Autoantibodies to glutamate receptor GluR3 in Rasmussen's encephalitis. Science 265:648–651
- 113. McNamara JO, Patel M, He XP, Janumpalli S, Whitney KD (1996) Glutamate receptor autoimmunity in Rasmussen's encephalitis. Cold Spring Harb Symp Quant Biol 61:327–332
- 114. Hemachudha T, Griffin DE, Giffels JJ, Johnson RT, Moser AB, Phanuphak P (1987) Myelin basic protein as an encephalitogen in encephalomyelitis and polyneuritis following rabies vaccination. N Engl J Med 316:369–374



- 115. Johnson RT, Griffin DE, Hirsch RL et al (1984) Measles encephalomyelitis—clinical and immunologic studies. N Engl J Med 310:137–141
- Tenembaum S, Chitnis T, Ness J, Hahn JS (2007) Acute disseminated encephalomyelitis. Neurology 68(Suppl 2):S23–S36
- 117. Schwimmbeck PL, Dyrberg T, Drachman DB, Oldstone MBA (1989) Molecular mimicry and myasthenia gravis. An autoantigenic site of the acetylcholine receptor α-subunit that has biologic activity and reacts immunochemically with herpes simplex virus. J Clin Invest 84:1174–1180
- 118. Fujinami RS, Nelson JA, Walker L, Oldstone MBA (1988) Sequence homology and immunologic cross-reactivity of human cytomegalovirus with HLA-DR β chain: a means for graft rejection and immunosuppression. J Virol 62:100–105
- Nauclér CS, Larsson S, Möller E (1996) A novel mechanism for virus-induced autoimmunity in humans. Immunol Rev 152:175–192
- 120. Zhao Z-S, Granucci F, Yeh L, Schaffer PA, Cantor H (1998) Molecular mimicry by herpes simplex virus-type 1: autoimmune disease after viral infection. Science 279:1344–1347
- 121. Gross DM, Forsthuber T, Tary-Lehmann M et al (1998) Identification of LFA-1 as a candidate autoantigen in treatmentresistant Lyme arthritis. Science 281:703-706
- 122. Kirvan CA, Swedo SE, Heuser JS, Cunningham MW (2003) Mimicry and autoantibody-mediated neuronal cell signaling in Sydenham chorea. Nat Med 9:914–920
- 123. Kirvan CA, Cox CJ, Swedo SE, Cunningham MW (2007) Tubulin is a neuronal target of autoantibodies in Sydenham's chorea. J Immunol 178:7412–7421
- 124. Thurau SR, Diedrichs-Möhring M, Fricke H, Arbogast S, Wildner G (1997) Molecular mimicry as a therapeutic approach for an autoimmune disease: oral treatment of uveitis-patients with an MHC-peptide crossreactive with autoantigen—first results. Immunol Lett 57:193–201
- 125. Lunardi C, Bason C, Navone R et al (2000) Systemic sclerosis immunoglobulin G autoantibodies bind the human cytomegalovirus late protein UL94 and induce apoptosis in human endothelial cells. Nat Med 6:1183–1186
- 126. Lunardi C, Dolcino M, Peterlana D et al (2006) Antibodies against human cytomegalovirus in the pathogenesis of systemic sclerosis: a gene array approach. PLoS Med 3:e2
- 127. Stathopoulou EA, Routsias JG, Stea EA, Moutsopoulos HM, Tzioufas AG (2005) Cross-reaction between antibodies to the major epitope of Ro60 kD autoantigen and a homologous peptide of Coxsackie virus 2B protein. Clin Exp Immunol 141:148–154
- 128. Hiemstra HS, Schloot NC, van Veelen PA et al (2001) Cytomegalovirus in autoimmunity: T cell crossreactivity to viral antigen and autoantigen glutamic acid decarboxylase. Proc Natl Acad Sci USA 98:3988–3991
- Ali F, Rowley M, Jayakrishnan B, Teuber S, Gershwin ME, Mackay IR (2011) Stiff-person syndrome (SPS) and anti-GAD-

- related CNS degenerations: protean additions to the autoimmune central neuropathies. J Autoimmun 37:79–87
- 130. Appelmelk BJ, Simoons-Smit I, Negrini R et al (1996) Potential role of molecular mimicry between *Helicobacter pylori* lipopolysaccharide and host Lewis blood group antigens in autoimmunity. Infect Immun 64:2031–2040
- 131. Sohn S, Lee ES, Bang D, Lee S (1998) Behcet's disease-like symptoms induced by the Herpes simplex virus in ICR mice. Eur J Dermatol 8:21–23
- 132. Lawson CM, O'Donoghue HL, Reed WD (1992) Mouse cytomegalovirus infection induces antibodies which cross-react with virus and cardiac myosin: A model for the study of molecular mimicry in the pathogenesis of viral myocarditis. Immunology 75:513–519
- 133. Christen U, Edelmann KH, McGavern DB et al (2004) A viral epitope that mimics a self antigen can accelerate but not initiate autoimmune diabetes. J Clin Invest 114:1290–1298
- 134. Watanabe K, Kim S, Nishiguchi M, Suzuki H, Watarai M (2005) Brucella melitensis infection associated with Guillain-Barré syndrome through molecular mimicry of host structures. FEMS Immunol Med Microbiol 45:121–127
- 135. Hintermann E, Holdener M, Bayer M et al (2011) Epitope spreading of the anti-CYP2D6 antibody response in patients with autoimmune hepatitis and in the CYP2D6 mouse model. J Autoimmun 37:242–253
- 136. Holdener M, Hintermann E, Bayer M et al (2008) Breaking tolerance to the natural human liver autoantigen cytochrome P450 2D6 by virus infection. J Exp Med 205:1409–1422
- 137. Avery AC, Zhao Z-S, Rodriguez A et al (1995) Resistance to herpes stromal keratitis conferred by an IgG2a-derived peptide. Nature 376:431–434
- 138. de Smet MD, Chan CC (2001) Regulation of ocular inflammation—what experimental and human studies have taught us. Prog Retin Eye Res 20:761–797
- 139. Shen DF, Chang MA, Matteson DM, Buggage R, Kozhich AT, Chan CC (2000) Biphasic ocular inflammatory response to endotoxin-induced uveitis in the mouse. Arch Ophthalmol 118:521–527
- 140. Fleck M, Kern ER, Zhou T, Lang B, Mountz JD (1998) Murine cytomegalovirus induces a Sjögren's syndrome-like disease in C57Bl/6-lpr/lpr mice. Arthritis Rheum 41:2175–2184
- 141. Yamada M, Zurbriggen A, Fujinami RS (1990) Monoclonal antibody to Theiler's murine encephalomyelitis virus defines a determinant on myelin and oligodendrocytes, and augments demyelination in experimental allergic encephalomyelitis. J Exp Med 171:1893–1907
- 142. Mokhtarian F, Zhang Z, Shi Y, Gonzales E, Sobel RA (1999) Molecular mimicry between a viral peptide and a myelin oligodendrocyte glycoprotein peptide induces autoimmune demyelinating disease in mice. J Neuroimmunol 95:43–54

