ERRATUM



Erratum to: Interaction of the signaling state analog and the apoprotein form of the orange carotenoid protein with the fluorescence recovery protein

 $\label{eq:marcus} Marcus \ Moldenhauer^1 \cdot Nikolai \ N. \ Sluchanko^{2,6} \cdot Neslihan \ N. \ Tavraz^1 \cdot Cornelia \ Junghans^1 \cdot David \ Buhrke^1 \cdot Mario \ Willoweit^1 \cdot Leonardo \ Chiappisi^3 \cdot Franz-Josef \ Schmitt^1 \cdot Vladana \ Vukojević^4 \cdot Evgeny \ A. \ Shirshin^5 \cdot Vladimir \ Y. \ Ponomarev^6 \cdot Vladimir \ Z. \ Paschenko^6 \cdot Michael \ Gradzielski^3 \cdot Eugene \ G. \ Maksimov^6 \cdot Thomas \ Friedrich^1$

Published online: 19 September 2017

© Springer Science+Business Media B.V. 2017

Erratum to: Photosynth Res DOI 10.1007/s11120-017-0346-2

In Fig. 1a in the original article, the amino acid side chains were incorrectly labeled in the structure representation of the orange carotenoid protein (OCP). The corrected Fig. 1 is printed in this erratum.

The online version of the original article can be found under doi:10.1007/s11120-017-0346-2.

- ☐ Thomas Friedrich friedrich@chem.tu-berlin.de
- ¹ Institut für Chemie Sekr. PC 14, Technische Universität Berlin, Straße des 17. Juni 135, 10623 Berlin, Germany
- A.N. Bach Institute of Biochemistry, Federal Research Center "Fundamentals of Biotechnology", Russian Academy of Sciences, 33 Leninsky prospect, building 1, Moscow, Russian Federation 119071
- ³ Institut für Chemie Sekr. TC 7, Technische Universität Berlin, Straße des 17. Juni 124, 10623 Berlin, Germany
- Department of Clinical Neuroscience, Center for Molecular Medicine, Karolinska Institutet, CMM L8:01, 17176 Stockholm, Sweden
- Department of Quantum Electronics, Faculty of Physics, M.V. Lomonosov Moscow State University, Leninskie Gory, Moscow, Russian Federation 119992
- Department of Biophysics, Faculty of Biology, M.V. Lomonosov Moscow State University, Leninskie Gory, 1, p. 12, Moscow, Russian Federation 119992



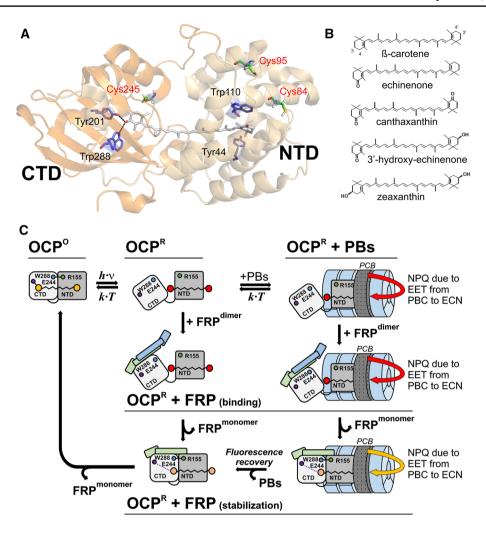


Fig. 1 Structure of the OCP protein and scheme for the interplay of OCP with PBs and FRP during NPQ. **a** OCP crystal structure (PDB entry 4XB5, (Leverenz et al. 2015)) with the canthaxanthin cofactor shown in stick representation. OCP is divided into an N-terminal and a C-terminal domain (NTD, CTD), with Trp288 and Tyr201 in the CTD involved in H-bond interactions (black dashed lines) to the 4-keto group of one of the β-rings. Tyr44 and Trp110 in the NTD are also involved in carotenoid coordination. The three cysteines in *Synechocystis* OCP are also shown (Cys84 and Cys95 in the NTD and Cys245 in the CTD). **b** Chemical structures of some carotenoids mentioned in this work. **c** Scheme of the major stages of NPQ. Upon absorption of blue-green light, the orange form of OCP (OCP^O) is photoconverted into the active red state (OCP^R) (Wilson et al. 2008). Consequently, NTD and CTD dissociate, the salt bridge between Glu244 (CTD) and Arg155 (NTD) breaks and the carotenoid trans-

locates into the NTD. NPQ activation in vivo is limited by the rate at which the OCP^R binds to PBs (Gorbunov et al. 2011; Maksimov et al. 2015a). During this dark phase of NPQ activation, OCP^R forms a stable complex with PBs, leading to PBs fluorescence quenching (Maksimov et al. 2014). Alternatively, OCP^R can spontaneously reconvert into the OCP^O form, or form a complex with FRP. The presence of FRP leads to an almost 10-fold increase of the OCP^R-OCP^O conversion rate (Boulay et al. 2010). Finally, under low-light conditions, OCP uncouples from PBs and energy flow from PBs to the photosynthetic reaction centers is restored. Of note, FRP exists in a dimeric form that becomes monomeric upon interaction with the OCP^R form (Sluchanko et al. 2017). Therefore, the interaction between OCP and FRP involves FRP dimer dissociation prior to or during binding to OCP^R

