

# Which East Asian herbal medicines can decrease viral infections?

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**Abstract** Whilst Western research for the COVID-19 crisis focuses on vaccination, in East Asia traditional herbal prescriptions are studied for SARS-CoV2 therapy. In Japan, Maoto (Ephedrae herba 4 g, Armeniacae semen 4 g, Cinnamomi cortex 3 g, and Glycyrrhizae radix 2 g, JPXVII) is used based on clinical evidence for its effect on early phase influenza (also caused by RNA viruses) comparable to that of

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oseltamivir. The Health Ministry of Thailand has approved Andrographis paniculata (Jap. Senshinren) extracts for treatment of COVID-19. Its combination (4 g) with Maoto, Maoto-ka-senshinren, seems most promising for the treatment of viral pandemics. In China, the official guideline for COVID-19 treatment contains TCM medications with antiviral, as well as immunmodulatory and anti-inflammatory effects such as: Qing-Fei-Pai-Du-Tang (Jap. Seihai-haidokuto) contains 21 drugs; Shufeng Jiedu Jiaonang (Bupleuri radix 8 g, Forsythiae fructus 8 g, Glycyrrhizae radix 4 g, Isatidis radix 8 g, Patriniae herba 8 g, Phragmitis rhizoma 6 g, Polygoni cuspidati rhizoma 10 g, Verbenae herba 8 g); Fufang Yuxingcao Heiji (Forsythiae fructus 0.6 g, Houttuyniae herba 6 g, Isatidis radix 1.5 g, Lonicerae flos 0.6 g, Scutellariae radix 1.5 g) first gained prominence during the 2002 SARS epidemic. With no Western medicine available, the following overview discusses efficacy and mechanisms in view of viral entry and replication of different East Asian herbal remedies for COVID-19 treatment.

**Keywords** COVID-19 · Influenza virus · Andrographis paniculata · Kampo · Maoto

### Introduction

Anti-viral activity has been reported from numerous medicinal plant extracts and preparations. For

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or *Diospyros kaki*. In our own research work, we were able to demonstrate that the pharmaceutical oleoresin Labdanum of *Cistus creticus* exerts pronounced in vitro anti-dengue virus activity (Kuchta et al. 2019).

The most probable targets of anti-viral natural products are those related to the replication cycle of viruses, which depends on several steps: Their recognition of the host cell, formation of the endosome, release of virus RNA within the cytoplasm, RNA replication and translation. The new endosome can then release the virus by exocytosis. All these steps can be targeted by entry-inhibitors, fusion inhibitors, RNA polymerase inhibitors, proteaseand release inhibitors.

The commonly used Oseltamivir is a neuraminidase inhibitor, i.e. a competitive inhibitor of the viral neuraminidase enzyme. Inhibition of the enzyme prevents cleaving of the virus from the host cell, and thus prevents the spreading of the virus. However, several resistance mechanisms are known. Current studies suggest that it is not effective in the treatment of the new COVID-19 pandemic. However, East Asian herbal medicines have shown anti-viral activity in the past and in the current reports on corona viruses.

For the current overview, Kampo prescriptions commonly recommended in Japan were assessed together with related herbal medications from China, Korea, and Thailand.

Although the past year has seen a flood of papers on Chinese medicines for the treatment of COVID-19, most of the prescriptions discussed therein are either new, previously untested drug combinations and / or hardly available internationally. Thus a selection was done for such prescriptions that are either commonly available on international markets or already established in the practice of Eastern Medicine in Europe. Especially established prescriptions—and simple variations thereof—that can easily be formulated as Single Prescriptions for individual patients from decoction pieces by doctors in the West were included.

### Anti-viral activity of Andrographis paniculata

Andrographis paniculata (Burm.f.) Nees (Fig. 1) is native to tropical Southeast Asia and traditionally used in Indian Ayurvedic medicine, traditional Thai medicine and traditional Indonesian Jamu medicine (Herrmann 1996) against diarrhea, bacterial dysentery and as a bitter tonic for numerous diseases. Especially in Jamu, *A. paniculata* is also well documented as a traditional treatment for malaria (Herrmann 1996).

In Thai Traditional Medicine, *A. paniculata* (Th. ฟ้าหะลาบโจร / Fha talai jone) is one of the most commonly used herbal drugs (Inta et al. 2013) and often administered as a decoction or pill for the prevention of all health problems. It is also used for the treatment of numerous ailments such as fever, cough, sore throat, apthous ulcer, wounds, abscesses, rashes, and as a carminative, for gastritis, pain, diabetes, hypertension, jaundice, and for detoxification (Inta et al. 2013).

Later, this medicinal plant was adopted into Traditional Chinese Medicine (TCM) as an antipyretic and against bronchitis, colitis, cystitis and similar inflammatory diseases (Wagner et al. 2011). Most recently, its introduction into the Chinese pharmacopoeia has resulted in its insertion into the newest edition of the European Pharmacopoeia (PhEur 9) and, due to current efforts to integrate more herbal drugs of East Asian Medicine, into the European regulatory framework.

The efficacy and safety of A. paniculata containing preparations for prophylaxis and symptomatic treatment of respiratory infections such as the common cold, bronchitis, sinusitis pharyngotonsillitis, urinary tract infections, and acute diarrhea has been supported by clinical studies, as laid out in a recent HMPC Assessment report (EMA/HMPC/320433/ 2012). Numerous, in the context of COVID-19 especially relevant studies in patients with viral lounge infections were performed in Scandinavia, South America, and India (Hancke et al. 1995; Caceres et al. 1997, 1999; Melchior et al. 1996; Saxena et al. 2010). A meta-analysis of 33 randomized controlled trials showed that A. paniculata extracts relieve inflammatory symptoms and shortens the duration of cough, sore throat, and disease duration in comparison with standard care (Hu et al. 2017). Saxena et al. (2010) carried out a randomized double blind placebo controlled clinical evaluation of *A. paniculata* extract in patients with uncomplicated upper respiratory tract infection regarding cough, fever, sputum, nasal mucus, headache, fatigue and sleep disorders resulting in 53% of improvement compared with placebo. When 158 common cold patients took 1.2 g of dried extract of *A. paniculata* for 5 days, symptoms like sleep disorder, nasal juice and sore throat improved. Akbar (2011) examined *A. paniculata* for the prevention and treatment of the common cold used a standardized *A. paniculata* dry extract for 5 days, resulting in a significant decrease of tiredness, sleeplessness, sore throat and nasal secretion.

Ding et al. (2017) demonstrated that treatment of C57BL/6 mice infected with the mouse-adapted H1N1 strain PR8A/PR/8/34 with andrographolide [10 mg/kg], with or without influenza virus entry inhibitor CL-385319 [10 mg/kg] improved body weight, lung function and reduced inflammation. The combination group had the highest survival rate but andrographolide treatment alone improved the survival rate as well as virus loads and inflammatory cytokine expression (Ding et al. 2017). Numerous natural products have been isolated from A. paniculata several of which-such as diverse lactones, flavonoids, diterpenes, and especially andrographolides (Fig. 1)-are regarded as contributors to its documented activity against influenza viruses.

All the above indicates that A. paniculata can be used for the prevention and treatment of the common cold and even influenza virus infection. It remains to be shown if A. paniculata is also effective in SARS-COVID-19 infection. However, similar strategies such as protease inhibition and cell modulation of cell surface receptors preventing viral entry have been shown in other viral infections including HIV. Furthermore, A. paniculata is well known for its antimalaria activity (Herrmann 1996). Its activity is quite similar to that of chloroquine-a synthetic antimalaria compound derivative of the alkaloid quinine that has been proposed for co-medication with A. paniculata in this indication (Hafid et al. 2015)-and is currently investigated for the treatment of COVID-19 infection.

Based on these findings and its established status in Thai Traditional Medicine, the Health Ministry Thailand has approved the use of the extract to treat early stages of Covid-19 as a pilot program (The Straits Times. DEC 30, 2020).

As far as the specific mechanism of action of *A. paniculata* against viral infections is concerned, it has been experimentally demonstrated that the andrographolide 14-deoxy-11,12-dehydroandrographolide (DAP) (Fig. 1), a major component of the raw drug with a minimum content of 0.8% of the sum of andrographolide and DAP in the dried drug material according to Ph.Eur., exerts potent anti-influenza A virus activity against A/chicken/Hubei/327/2004 (H5N1), A/duck/Hubei/XN/2007 (H5N1), A/PR/8/34 (H1N1), A/NanChang/08/2010 (H1N1) and A/HuNan/01/2014 (H3N2) in vitro on A549 and MDCK cells and inhibits the replication of the H5N1 influenza virus by preventing the export of the viral ribonucleoprotein complexes from the nucleus (Cai et al. 2015).

For H5N1, DAP exhibited a CC50 (cytotoxic concentration required to reduce cell viability by 50% for uninfected cells determined by CCK-8 assay) and an IC50 (inhibition concentration to reduce the cytopathic effect (CPE) by 50% caused by A/chicken/Hubei/327/2004 (H5N1) in the same order of magnitude as the positive control Ribavirin (Cai et al. 2015).

Andrographolide itself was also shown to contribute to the overall anti-viral activity of *A. paniculata* extracts. In the case of the enterovirus D68 (EV-D68), that has emerged as a significant respiratory pathogen in recent years, it could be demonstrated in an in vitro virus model on human rhabdomyosarcoma RD cells (ATCC, CCL-136) that andrographolide prevents its replication by inhibiting the acidification of virus-containing endocytic vesicles, resulting in a dramatic inhibition of EV-D68 RNA replication (EC50=3.45 mM). In comparison, its median cytotoxic, lethal concentration was much higher at 75 mM (Wang et al. 2018).

A. paniculata in general and andrographolide in particular have also proven effective against the dengue virus (Paemanee et al. 2019). Here, the human HepG2 liver cell (ATCC Cat No. HB-8065) were infected with DENV 2 and subsequently incubated with andrographolide (50, 100, and 200  $\mu$ M). A proteomic based approach demonstrated an important role for Glucose regulated protein 78 (GRP78) and the unfolded protein response (UPR) mechanism in mediating the anti-dengue virus activity of andrographolide, which might, in part, explain the broad antiviral activity of andrographolide (Paemanee et al. 2019).

In this context, yet another andrographolide, 3,19isopropylideneandrographolide (IPAD) was shown to be effective against Herpes Simplex Virus. IPAD (22.50  $\mu$ M) completely suppressed ICP8 transcription and translation as well as DNA replication and HSV gD protein (Envelope glycoprotein D of the Human herpesvirus 1 (strain 17) (HHV-1) (Human herpes simplex virus 1)) expression in the tested virus strains in a Vero host cell model (Kongyingyoes et al. 2016). This envelope glycoprotein binds to the potential host cell entry receptors like TNFRSF14/HVEM, and NECTIN1 and may trigger fusion with host membrane by recruiting the fusion machinery.

A comparison of human patient data with those from a rat model showed that the pharmacokinetics of andrographolides are similar in both species. They are rapidly and almost completely absorbed (T1/2abs of about 25 min) into the blood (bioavailability= 91%, F=0.91) after oral administration at a therapeutic dose (20 mg/kg). Andrographolide binds to blood proteins and is distributed in blood and tissues within 1–2 h. The elimination half-time is in the range of 2–7 h (Panossian et al. 2000). A tissue distribution study revealed the highest concentration in kidney, followed by the liver, spleen, and brain, whereas an almost identical concentration was observed in heart and lungs (Bera et al. 2014).

It is however important to note that bioactivity of andrographolides is not limited to the anti-viral effect itself but also affects therapeutically relevant side effects of the infection. E.g. andrographolide was shown to inhibit Influenza A virus induced inflammation in a murine model through NF- $\kappa$ b and JAK-STAT signaling pathway (Ding et al. 2017).

In this context, it is important to note that *A. paniculata* has also been discussed as an Adaptogen (Panossian et al. 2021)—a category of natural compounds or herbal extracts that increase adaptability, resilience, and survival of organisms; they increase "the state of nonspecific resistance" of organisms to harmful factors, including bacterial and viral pathogens (Lazarev et al. 1959). In Ayurveda, the plants with traditionally use as adaptogens are referred to as Rasayana and are used as rejuvenating and for improving the overall health of anyone undergoing this treatment. It is therefore not surprising that *A. paniculata* is regarded as one of the

most important rasayana drugs (Thakur et al. 2014, 2015; Raina et al. 2013). In this context, the chemopreventive effects of *A. paniculata* extracts and Andrographolide were previously demonstrated (Sheeja and Kuttan 2006; Singh et al. 2009).

## Anti-viral activity of Kampo prescriptions such as Maoto

In Japan several Kampo prescriptions like Maoto, Kakkonto (Kurokawa et al. 1996; Okabayashi et al. 2014), Shahakusan (Hokari et al. 2012), Shoseiryuto (Nagai and Yamada 1994, 1998; Nagai et al. 1996; Yamada and Nagai 1998), Daiokanzoto (Watanabe 2018) and Hochuekkito (Dan et al. 2018) (Table 1) have been investigated for their effect against influenza virus infection.

All the above are traditionally prepared as decoctions (i.e. hot water extracts) according to the legal requirements of the current Japanese Pharmacopoeia (JPXVII, p. 22): Heat one-day dose of crude drugs with 400–600 ml of water until loss of about half the amount of added water spending more than 30 min, and filter through a cloth while warm.

Among all examined Kampo prescriptions, the most detailed information was available for Maoto (Ephedrae herba 4 g, Armeniacae semen 4 g, Cinnamomi cortex 3 g, and Glycyrrhizae radix 2 g). This prescription will therefore form the basis for the further discussion in the following paragraphs. Maoto (麻黄湯) was published already during the second century AD in the Shanghan Lun (傷寒論, Jap. Shoukanron)-one of the foundational texts of Ancient Chinese Medicine-under the Chinese name Ma-Huang-Tang, under which it is still used in TCM today. In Korean medicine, which developed from Ancient Chinese Medicine as a third sister system to TCM and Kampo, the identical prescription is referred to as Mahwang-tang (마황탕). Maoto is commonly applied for febrile diseases with symptoms like high fever and cough. Kampo prescriptions like Maoto that contain Ephedrae herba (Ephedra sinica Stapf) (Fig. 2) are also referred to as Mao-zai (麻黄剤).

Recently, Kampo clinical trials for influenza virus infection have been reported in Japan. E.g. Nagai et al. (2014) found that orally administered Maoto (0.9 and 1.6 g/kg/day) had significant anti-pyretic

<b>Maoto</b> 麻黄湯		Kakkonto 葛根湯		<b>Shahakusan</b> 瀉白散		Shoseiryuto 小青竜湯		Daiokanzoto 大黄甘草湯		Hochuekkito 補中益気湯	
										Angelicae sinensis radix	3 g
Armeniacae semen	4 g										
						Asiasari radix	3 g			A - two - 11 - w - 41	4 -
										Astragali radix	4 g
										Atractylodis mac. rhizoma	4 g
										Bupleuri radix	2 g
										Cimicifugae rhizoma	1 g
Cinnamomi cortex	3 g	Cinnamomi cortex	3 g			Cinnamomi cortex	3 g				
										Citri reticulatae pericarpium	2 g
Ephedrae herba	4 g	Ephedrae herba	4 g			Ephedrae herba	3 g				
										Ginseng radix	4 g
Glycyrrhizae radix	2 g	Glycyrrhizae radix	2 g	Glycyrrhizae radix	2 g	Glycyrrhizae radix	3 g	Glycyrrhizae radix	2 g	Glycyrrhizae radix	1 g
		Jujubae fructus	4 g							Jujubae fructus	2 g
				Lycii radicis cortex	4 g						
				Mori radicis cortex	4 g						
		Paeoniae radix	3 g			Paeoniae radix	3 g				
						Pinelliae rhizoma	6 g				
		Puerariae radix	8 g								
								Rhei rhizoma	4 g		
						Schisandrae fructus	3 g				
		Zingiberis rhizoma	1 g			Zingiberis rhizoma	3 g			Zingiberis rhizoma	1 g
				Long-grained rice	2 g						

Table 1 Anti-viral Kampo prescriptions including their one-day dose of crude drugs for decoction according to JPh

activity in influenza virus infected A/J mice after upper respiratory tract infection with the influenza virus A/PR/8/34. Administration of Maoto (0.8 and 1.3 g/kg/day) further significantly decreased the virus titers in both nasal and bronchoalveolar lavage fluids 52 h after infection and increased the anti-influenza virus antibodies IgM, IgA and IgG1 resulting in the binding of the virus (Nagai et al. 2014).

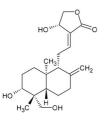
Masui et al. (2017) set up the culturing assay system for A549 cells which were infected with influenza virus A (PR8) in order to determine the virus titers in the culture supernatant, intracellular viral proteins and viral RNA. When the infected cells were treated with 400  $\mu$ g/ml of a commercial Maoto, the extract significantly reduced the virus titer as well as the production of viral surface proteins such as M2 and neuraminidase (NP), thus preventing viral entry (Masui et al. 2017).

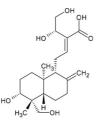
Maoto can also inhibit the uncoating of influenza virus. Furthermore, the inhibition of endosomal

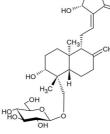
acidification by Maoto may prevent the release of the influenza virus through the inhibition of V-ATPase into the cytoplasm (Masui et al. 2017). Maoto is therefore the most favorable Kampo medicine for influenza virus and influenza illness.

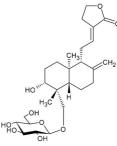
Nabeshima et al. (2012) investigated Maoto [2.5 g TID of commercial granules dissolved in warm water for 5 days] for the treatment of seasonal influenza in a randomized clinical trial. 28 influenza patients within 48 h of fever onset were randomly assigned to Maoto (n=10), Oseltamivir [75 mg BID for 5 days] (n=8), or Zanamivir [20 mg BID for 5 days] (n=10) and data collected for their total symptom score from self-reported symptom cards and the duration of fever (> 37.5 °C). No significant between-group differences were found for total symptom score among three groups without severe adverse effects. Nabeshima et al. (2012) thus demonstrated that Maoto affects the early phase of influenza virus infection, with an anti-influenza activity comparable to that of oseltamivir.







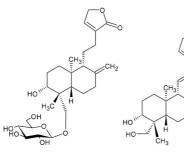


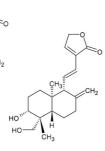


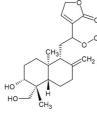
Andrographolide

- Andrographolic acid
- Andrographiside

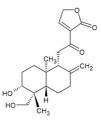
Deoxyandrographoside





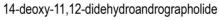


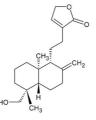
Deoxy-methoxy-andrographolide



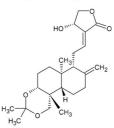
Neoandrographolide

Deoxy-oxo-andrographolide





Dideoxy-andrographolide



3,19-isopropylideneandrographolide

Fig. 1 Andrographis paniculata (Burm.f.) Nees and some of its active constituents Ephedrine Norephedrine Pseudoephedrine D-Norpseudoephedrine L-Methylephedrin L-Ephedrine D-Pseudo-ephedrine

As mentioned above, Maoto consists of the four individual raw drugs: Ephedrae herba 4 g (Fig. 2), Armeniacae semen 4 g (Fig. 3), Cinnamomi cortex 3 g (Fig. 4), and Glycyrrhizae radix 2 g (Fig. 5). When comparing the composition of this prescription with the other Kampo prescriptions with proven antiviral effects listed in the above table, three individual raw drugs seem most characteristic for this anti-viral indication, namely Ephedrae herba, Cinnamomi cortex, Glycyrrhizae radix.

Although ephedrine alkaloids in *Ephedra* spp. (Fig. 2) are sometimes regarded as the most important component in Maoto, when ephedrine was removed from Ephedra extract, typical side effect like excitement, sleep disorder, palpitations and gastrointestinal disorders could be eliminated in a mouse model (Takemoto et al. 2018). Nevertheless, the same Ephedrine alkaloids-free Ephedra extract (EFE) reduced formalin-induced pain in a dose-dependent manner in male ICR mice that were orally administered 350 mg/kg EFE, or 700 mg/kg Ephedra Herb extract for 3 days (Hyuga et al. 2016). EFE showed anti-influenza virus activity inhibiting the infection of MDCK cells incubated for 72 h in a twofold serial dilution of 10 µM oseltamivir, 50 µg/ml EFE, or 50 µg/ml Ephedra Herb extract with influenza virus A/WSN/33(H1N1) in a concentration-dependent manner (Hyuga et al. 2016). The authors therefore propose an active fraction of the condensed tannin mixture having molecular weight of 45,000 as an alternative active principle of Ephedra extract (Takemoto et al. 2018). These finding are especially interesting with regards to the Central Asian species Ephedra przewalskii Stapf-also used in the same traditional indications by the natives of both the Western Chinese region of Xinjiang and the Gobi desert in (Inner-) Mongolia-that has been experimentally demonstrated not to contain any significant amounts of ephedrine (Long et al. 2005).

Nevertheless, a study (Wei et al. 2019) aiming to screen antiviral components of the common Ephedrae herba drug confirmed the activity of L-methylephedrin (LMEP), L-ephedrine (LEP) and D-pseudoephedrine (DPEP) in MDCK cells infected by mouseadapted influenza virus A/PR8/34 (H1N1). After 24 h treatment, the virus load in the LMEP 31.25  $\mu$ g/ml, LEP 15.63 µg/ml and DPEP 15.63 µg/ml groups was significantly lower than that in oseltamivir positive control. In a male ICR mouse model, the mice were treated by gavage with oseltamivir (22 mg/kg), LEP or DPEP (40, 20, 10 mg/kg) solubilized in physiological saline for 7 days, resulting in a significant inhibition of mRNA expression levels of the TLR3, TLR4 and TLR7 signaling pathways and further down-regulated TNF- $\alpha$  levels and up-regulated IFN- $\beta$ levels (Wei et al. 2019). These Ephedra alkaloids therefore exert an antiviral effect in vitro which may be closely related to the inhibition of viral replication and the modulation of inflammatory response by adjusting the host's TLRs and RIG-1 (Retinoic acid Inducible Gene I)—an intracellular receptor of the innate immune system-pathways. Further, a vital fluorescence microscopic study (Mantani et al. 1999) showed that the extract of Ephedrae herba (100-400  $\mu$ g/ml) inhibited the acidification of endosomes and lysosomes in Madin-Darby canine kidney cells in a concentration-dependent manner, inhibiting the growth of influenza A/PR/8/34 (H1N1) (PR8) virus. Conversely, virus growth resumed concomitantly with the reappearance of acidified ELS after removal of the extract. The fact that its inhibitory effect was completely or partially reversed by FeCl<sub>3</sub>, a tanninreactive agent, indicates that tannins form an active fraction of the extract.

Morimoto et al. (1986) reported that Cinnamon bark contains the procyanidin heptamer cinnamtannin A3 that may contribute to the anti-influenza activity of Maoto. For example, Zhuang et al. (2009) found that the butanol fraction (Fr.2) of Cinnamomi Cortex extract (CC) showed the highest activities of both CC and Fr.2 on wild-type severe acute respiratory syndrome coronavirus (wtSARS-CoV) when the viruses were treated by the extracts before challenging with IC50 values of  $43.1 \pm 2.8$  and  $7.8 \pm 0.3 \,\mu$ g/ml and SI values of 8.4 and 23.1, respectively. Zhuang et al. (2009) were furthermore able to demonstrate that this extract could interfere with the clathrindependent endocytosis pathway using transferrin receptor (TfR) on Jurkat cells as an indicator.

Finally, in the case of Glycyrrhizae radix, the triterpene Glycyrrhizin (or glycyrrhizic acid or glycyrrhizinic acid) (GA) (Fig. 5) has been identified as

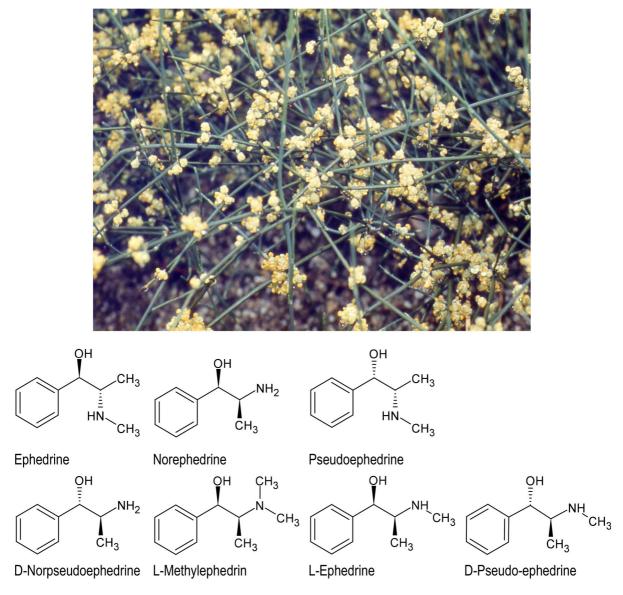


Fig. 2 Ephedra sinica Stapf and some of its active constituents

the principal bioactive ingredient with regards to its anti-viral, anti-inflammatory and hepatoprotective effects. Its anti-viral effects are manifold and have filled an entire review article already (Sun et al. 2019). For example, Hsieh C et al. (2012) were able to demonstrate that GA inhibitsPI3K/AKT signaling pathway regulated viral entry, via its neuraminidase inhibiting activity. Utsonomiya et al. (1997) investigated the anti-influenza effect of GA in influenza virus A2 (H2N2) infected BALB/c mice. The results demonstrated that GA may protect mice exposed to a lethal amount of influenza virus by the stimulation of INF- $\gamma$  production by T cells. The test compound, which consisted of one molecule of GA and two molecules of glucuronic acid was administered intraperitoneally (10 mg per kg of body weight) 1 day before infection and 1 and 4 days post-infection. All of the mice survived over the 21-day experimental period (Utsonomiya et al. 1997). In an in vitro herpes simplex virus-1 (HSV-1) infection model, (Lee et al. 2017) demonstrated that guercetin, a major component of *Glycyrrhiza uralensis*,



Fig. 3 Prunus armeniaca L.



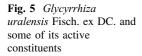
Fig. 4 Cinnamomum cassia (L.) J.Presl

significantly lowered HSV infectivity in Raw 264.7 cells, resulting in a dramatic decrease in plaque formation in Vero cells when they were incubated with infected cell lysates treated with quercetin. The same concentrations of quercetin further inhibited the expressions of HSV proteins (gD, ICP0) and genes (ICP0, UL13, UL52). Interestingly, quercetin in all tested concentrations specifically suppressed the expression of TLR-3, and this led to the inhibition of inflammatory transcriptional factors (NF- $\kappa$ B and IRF3) (Lee et al. 2017). Moreover, glycyrrhizin in

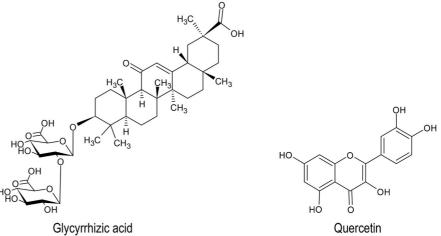
Glycyrrhizae radix is also reported to have antiinfluenza activity, as this drug selectively suppressed viral protein synthesis (IC50=0.27 mg/ml) in human influenza virus strain A/Udorn/72 (H3N2) on MDCK (+) host cells (Nomura et al. 2019).

Therefore, the anti-influenza activity of Maoto might be accelerated by the addition of the other component herbs besides Ephedrae herba, e.g. Cinnamomi cortex or Glycyrrhizae radix.

With the single exception of Armeniacae semen all the above raw drugs have entered the most recent







edition of the European Pharmacopoeia (PhEur 9) as part of an initiative to include East Asian raw drugs in the European regulatory framework in order to facilitate uniform and reliable quality control standards (Table 2).

The combination of Maoto and Andrographitis herba is referred to as Ma-Huang-Tang-Jia-Chuan-Xin-Lian in Chinese and in Korean as Mahwangtang-ga-cheonsimryeon (마황탕가천심린), respectively.

Another very interesting Kampo prescription is Kakkonto (葛根湯), which is listed with the official indication influenza virus. This prescription is relatively similar to Maoto, consisting of the raw drugs Puerariae radix, Ephedrae herba, Paeoniae radix, Jujubae fructus, Cinnamomi cortex, Glycyrrhizae

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radix, and Zingiberis rhizoma (Table 1). Especially Paeoniae radix contains gallotannin (Nishizawa et al. 1984) and may therefore be another very promising candidate for the treatment of the early corona virus (COVID-19) infection. This should be especially true for its combination with Andrographitis herba (Table 3).

In Chinese, this prescription is referred to as Ge-Gen-Tang-Jia-Chuan-Xin-Lian and in Korean as Galgeun-tang-ga-cheonsimryeon (갈근탕가천심린), respectively.

In the Kampo theory, viral infections as well as infections with bacteria and parasites are all subsumed under the concept external noxae (Jap. Gaija/ 外邪), the traditional indication of both Kakkonto and Maoto. Kakkonto is rather used in cases with

Table 2 麻黄湯加穿心蓮 (Maoto-ka-senshinren)

4 g
4 g
3 g
4 g
2 g

Table 3 葛根湯加穿心蓮 (Kakkonto-ka-senshinren)

Andrographitis herba	4 g
Cinnamomi cortex	3 g
Ephedrae herba	4 g
Glycyrrhizae radix	2 g
Jujubae fructus	4 g
Paeoniae radix	3 g
Puerariae radix	8 g
Zingiberis rhizoma	1 g

sweating and fever, whilst Maoto is used in patients with dry fever and cough. Thus, Kakkonto should also be suitable for the treatment of COVID-19, especially in earlier stage as the combination of fever (ca. 37 °C or higher) and pain fits its traditional indication.

To prevent the development of symptoms, Hochuekkito (Table 1) can be used. For prevention of pneumonia, Saikatsugekito (柴葛解肌湯) has been proposed, which can be combined with Kakkonto or Shosaikoto-ka-kikyo-sekko (小柴胡湯加桔梗石膏). In the stage of pneumonia, next to Western medical treatment, Seihaito (清肺湯) is an option. For the stage of recovery from pneumonia, also Seihaito or Chikujountanto (竹茹温胆湯) have been proposed. (Composition of minor mentioned prescriptions: Appendix 1).

During the recent COVID-19 outbreak, in China traditional Chinese medicine (TCM) was immediately included in the organized clinical response with great success. More than 3100 TCM staff were dispatched to Hubei province and TCM experts were fully integrated in the whole emergency medicine process. This includes a TCM scheme within the official guideline on diagnosis and treatment of COVID-19 (Anonymous 2020a). According to this TCM expert group tasked by the Chinese government

with the fight against COVID-19 in Wuhan, the best traditional prescription for the treatment of the infection is Seihaihaidokuto (清肺排毒湯) or Qing-Fei-Pai-Du-Tang in Chinese. Seihaihaidokuto has been officially promoted as a general prescription in the diagnosis and treatment plan of COVID-19 in China (Anonymous 2020a). This state-approved COVID-19 official formula is however very complex and consists of 21 individual herbal drugs (Table 4).

Seihaihaidokuto (清肺排毒湯) was developed by combining older prescriptions and thus includes Makyokansekito (麻杏甘石湯), Goreisan (五苓散), Shosaikoto (小柴胡湯), and Yakanmaoto (射干麻黄 湯). Goreisan (五苓散) was included in Seihaihaidokuto (清肺排毒湯) because the COVID-19 infection was shown to cause a burst in cytokine production, leading to swellings, inflammation, and diarrhea. Besides prescriptions that directly counteract the symptoms of the COVID-19 infections, Bofutsushosan (防風通聖散) was also included in order to counteract feelings of fullness of the belly and flatulence that typically accompany therapy with high doses of Ephedrae herba. The Chinese National Administration of Traditional Chinese Medicine reports that until the first week of February 2020, 214 COVID-19 patients in the provinces of Shanxi, Hebei, Heilongjiang and Shaanxi were treated with the combination prescription with overall effective rate  $\geq$  90%. In a majority of patients ( $\geq$  60%) symptoms were markedly improved and in many other the illness was at least stabilized (Zhao et al. 2020). In a subsequent study, 701 COVID-19 patients received the same treatment, with 130 patients (18.5%)completely cured, disappearance of the characteristic symptoms of COVID-19 such as fever and cough in further 51 patients (7.27%), improvement of said symptoms in 268 patients (38.2%), and stabilization of the state of disease in 212 patients (30.2%), respectively (Anonymous 2020b).

Although most experience with Seihaihaidokuto (清肺排毒湯) currently stems directly from Wuhan, successful treatments with very similar combinations such as the parallel administration of Kakkonto (葛根 湯), Shoseiryuto (小青竜湯), Goreisan (五苓散), Bofutsushosan (防風通聖散) have been reported from Japan. (Composition of minor mentioned prescriptions: Appendix 1).

In China—in addition to the abovementioned long established prescriptions—some newer TCM

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Table 4 清肺排毒湯	(Seihaihaidokuto)
---------------	-------------------

9 g
9 g
9 g
6 g
9 g
9 g
6 g
9 g
16 g
9 g
6 g
12 g
9 g
9 g
6 g
20 g
9 g
9 g
15 g
6 g
9 g

formulations have been used for COVID-19 therapy, many of which were developed during the 2002 Severe acute respiratory syndrome (SARS) epidemic (Liu et al. 2012). Of these innovative prescriptions, Shufeng Jiedu Jiaonang (疏风解毒胶囊)—Sofugedokukono (疏風解毒膠囊) in Japanese—has proven especially effective and has also entered the Chinese national treatment guidelines for COVID-19. In preclinical studies immunomodulatory and anti-inflammatory effects have been shown against severe actue respiratory syndrom SARS-CoV2-caused pneumonia (Tao et al. 2020). Clinical studies are on their way (Xia et al. 2020; Chen et al. 2021) (Table 5).

Another TCM prescription that first gained prominence during the 2002 SARS epidemic (Liu et al. 2012) for which clinical data are well established (Zheng et al. 2017) and which has been registered and used successfully for the treatment of COVID-19 in China is Fufang Yuxingcao Heiji (复方魚腥草合剤), whose activity against influenza viruses has also been demonstrated in vitro (Zu et al. 2010) (Table 6).

Further clinical examinations of this preparation for COVID-19 therapy are currently ongoing.

#### Table 5 疏风解毒胶囊 (Shufeng Jiedu Jiaonang)

Bupleuri radix	8 g
Forsythiae fructus	8 g
Glycyrrhizae radix	4 g
Isatidis radix	8 g
Patriniae herba	8 g
Phragmitis rhizoma	6 g
Polygoni cuspidati rhizoma	10 g
Verbenae herba	8 g

### Table 6 复方魚腥草合剂 (Fufang Yuxingcao Heiji)

Forsythiae fructus	0.6 g
Houttuyniae herba	6 g
Isatidis radix	1.5 g
Lonicerae flos	0.6 g
Scutellariae radix	1.5 g

### Conclusion

Maoto has the ability to act at all 3 steps important for viral proliferation: It has been shown to enhance the production of antibodies such as IgG, IgM and IgA against influenza virus (Nagai et al. 2014). Maoto further reduces the virus titer (of H1N1 in A549 cells) as well as the production of viral surface proteins such as M2 and neuraminidase (NP) hence preventing viral entry and release (Masui et al. 2017).

In classical Kampo theory, viral infections as well as infections with bacteria and parasites are all subsumed under the concept external noxae. Maotoka-senshinren, i.e. Maoto complemented by Andrographitis herba—a drug with significant anti-viral activity in its own right—can be recommended for the treatment of those infectious diseases that are characterized by fever.

A development of resistance against Maoto-kasenshinren is not to be expected, as several thousand individual phytochemical constituents are contained in the full extract mixture the continuous application of which should make it almost impossible for the virus to adapt. Furthermore, as stated above, the Maoto prescription has been in continuous and safe use since the second century AD with no known cases of resistance development. Therefore, we strongly suggest that the combination might be used for influenza viruses and tested for the new corona virus, SARS-COV2 that is currently spreading throughout the world.

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**Appendix 1** 

See Table 7.

 Table 7
 Minor prescriptions

4 g	
2 g	
2.5 g	
1 g	
6 g	
2 g	
3 g	
8 g	
2 g	
1 g	
6 g	
2 g	
2 g	
10 g	
2 g	
5 g	
3 g	
3 g	
4 g	
3 g	
2 g	
2 g	
2 g	
	2 g 2.5 g 1 g 6 g 2 g 3 g 8 g 2 g 1 g 6 g 2 g 2 g 1 0 g 2 g 3 g 3 g 3 g 3 g 3 g 3 g 3 g 3

Table 7 continued

Chiri reticulatae pericarpium         2 g           Fritilariae thunbergin bulbus         2 g           Giycyrthizae radix         1 g           Jujuboe Tructus         2 g           Optiopogonis radix         3 g           Platycodonis radix         2 g           Optiopogonis radix         2 g           Poriae selectorium         3 g           Scutellariae radix         2 g           Zingiberis finizoma         1 g           Scutellariae radix         2 g           Aurantii fuctus immaturus         2 g           Bunbusce callis in taenias         3 g           Bupleuri radix         3 g           Cyperi faizoma         1 g           Cyperi faizoma         1 g           Glycyrthizae radix         1 g           Cyperi faizoma         1 g           Glycyrthizae radix         1 g           Optiopogonis radix         3 g           Glycyrthizae radix         1 g           Cyperi faizoma         1 g           Glycyrthizae radix         1 g           Optiopogonis radix         3 g           Portae selectorium         3 g           Portae selectorium         3 g           Strucellariae terdix	Table / continued		
Gardeniae fructus     2 g       Grycyntrize radix     1 g       Jujbae fructus     2 g       Mori radicis cortex     2 g       Ophiopogonis radix     3 g       Pariae solerotium     3 g       Schiandrae fructus     1 g       Schiandrae fructus     2 g       Zingiberis rhizoma     1 g       Chikujountanto (代始温思想)     1 g       Aurantii fructus immatrurus     2 g       Bambusac calls in taenias     3 g       Citri reficulatus pericampium     2 g       Ginseng radix     1 g       Cyperi rhizoma     1 g       Cyperi rhizoma     2 g       Ginseng radix     1 g       Cyperi rhizoma     2 g       Ginseng radix     1 g       Ophiopogonis radix     3 g       Portiae sclerotium     3 g       Zingiberis rhizoma     1 g       Ophiopogonis radix     3 g       Portiae sclerotium     3 g       Zingiberis rhizoma     1 g       Ophiopogonis radix     2 g       Armeniaca semen     9 g     4 g       Gyperimibers andix     6 g     2 g       Grypurimibers radix     6 g     2 g       Ophioponibrosum     6 g     2 g       Ophioponibrosum     6 g	Citri reticulatae pericarpium	2 g	
Glycyrhizac radix     1 g       Jujubac fractus     2 g       Moir indicis cortex     2 g       Ophiopogonis radix     3 g       Plarycodonis radix     2 g       Poriae sclerorium     3 g       Scutellariae radis     2 g       Zugiberis rihozoma     1 g       Chikujountanto (竹街温胆湯)     1       Aurantii fractus immaturus     2 g       Bambusea caulis in taenias     3 g       Bupleuri radix     3 g       Copridis rihozoma     1 g       Cyperi rihizoma     2 g       Glycyrrihizac radix     1 g       Ophiopogonis radix     3 g       Particulate pericarptum     2 g       Copridis rihozoma     1 g       Ophiopogonis radix     3 g       Particulate pericarptum     2 g       Ophiopogonis radix     3 g       Particulate pericarptum     2 g       Ophiopogonis radix     3 g       Partis elizoma     1 g       Ophiopogonis radix     3 g       Partis elizoma     1 g       Ophiopogonis radix     3 g       Partis ributoma     1 g       Ophiopogonis radix     2 g       Ophiopogonis radix     3 g       Differis ributoma     1 g       Cipyeri ributoma     1 g<	Fritillariae thunbergii bulbus	2 g	
Jajubae fructus     2 g       Moir radicis cortex     2 g       Ophiopogonis radix     2 g       Platycodonis radix     2 g       Priate sclerotium     3 g       Schisandrae fructus     1 g       Scutellariae radix     2 g       Zingleris rhizoma     1 g       Chikujountano (竹倉鼎即得)     2 g       Auranti fructus immaturus     2 g       Bambusae caulis in taenias     3 g       Bupleuri radix     3 g       Chiri circiculatae pericarpium     2 g       Copidis rhizoma     1 g       Ophiopogonis radix     3 g       Binolize radix     1 g       Ophiopogonis radix     3 g       Pinelliae rhizoma     1 g       Ophiopogonis radix     2 g       Portiae sclerotium     3 g       Zingiberis rhizoma     1 g       Ophiopogonis radix     2 g       Portiae sclerotium     3 g       Zingiberis rhizoma     1 g       Makyokanschio (係音甘音湯) E     Makyosekikanto (條 音右音影) E       Armentiace semen     9 g     4 g       Cypeur fibrosum     1 g       Grypyprinize radix     6 g       Grypyprinize radix     6 g       Ophioposite racecorphalae     4 g       Phiorona     2 g	Gardeniae fructus	2 g	
Mori radicis cortex2 gOptiopogonis radix3 gPlatycodonis radix2 gPorias scleorium3 gSchiandrae fructus1 gScutellariae radix2 gZingiberis rhizoma1 gChitujountanto (竹浜温胆湯)1Aurati fructus immaturus2 gBambusac callis in taenias3 gBupleuri radix3 gCoptidis rhizoma1 gCoptidis rhizoma1 gOphiopogonis radix3 gCiri reticultase pericarpium2 gGinseng radix1 gOphiopogonis radix3 gPinelliae rhizoma5 gPlaycodi radix2 gPoriae sclerolium3 gArmeniacae semen9 g4 g4 gGlycyrrhizae radix6 gOptiopiosum18 gIo g2 gPoriae sclerolium2 gPoriae sclerolium3 gPure taita introma6 gArmeniacae semen9 g4 g10 gGorsent (Ti-Sth)10 gArmeniacae semen9 g4 g10 gPoriae sclerolium3 gPoriae sclerolium3 gPoriae sclerolium4 gArmeniacae semen9 g4 g10 gGorsent (Ti-Sth)10 gArmeniacae semen9 g4 g10 gPoriae sclerolium4 gArmeniacae semen4 gPoriae sclerolium4 gArmeniacae semen<	Glycyrrhizae radix	1 g	
Ophiopogonis radix       3 g         Play codonis radix       2 g         Poriae sclerotium       3 g         Schisandra fructus       1 g         Scuellariae radix       2 g         Zingiberis rhizoma       1 g         Chikajountanto (竹當選問題)	Jujubae fructus	2 g	
Platycodonis radix       2 g         Poriae sclerotium       3 g         Schisandne fructus       1 g         Schisandne fructus       2 g         Zingiberis rhizoma       1 g         Chikujountanto (行街温胆湯)       2         Bambusae caulis in taenias       3 g         Bupleuri radix       3 g         Christiouta pericarpium       2 g         Coptidis rhizoma       1 g         Cyperi rhizoma       2 g         Ginseng radix       1 g         Ophiopogonis radix       3 g         Pinelliae rhizoma       5 g         Platycoi radix       2 g         Ophiopogonis radix       1 g         Ophiopogonis radix       2 g         Poriae sclerotium       3 g         Zingiberis rhizoma       1 g         Makyokansekito (隊 TélTáliš) H       ************************************	Mori radicis cortex	2 g	
Poriae sclerotium       3 g         Schiandrae fructus       1 g         Scutellariae radix       2 g         Zingiberis rhizoma       1 g         Chikujountanto (竹筎温胆湯)       2         Auranti fructus immaturus       2 g         Bambusae caulis in taenias       3 g         Citri reticulatae pericarpium       2 g         Copidis rhizoma       1 g         Cyperi rhizona       2 g         Ginseng radix       1 g         Ophiopogonis radix       3 g         Pinelliae rhizoma       3 g         Pinelliae rhizoma       3 g         Zingiberis rhizoma       3 g         Ophiopogonis radix       2 g         Portae sclerotium       3 g         Zingiberis rhizoma       1 g         Ophiopogonis radix       2 g         Portae sclerotium       3 g         Armenicae semen       9 g       4 g         Glycyrrhizae radix       6 g       2 g         Goreisan (茄花常想)       1       1         Goreisan (茄花常数)       1 g       1         Armenicae semen       9 g       4 g         Glycyrrhizae radix       6 g       2 g         Goreisan (茄花常想)       1 <td>Ophiopogonis radix</td> <td>3 g</td> <td></td>	Ophiopogonis radix	3 g	
Schisandrae fructus       1         Settellariae radix       2         Stangberis rhizoma       1         Aurantii fructus immaturus       2         Aurantii fructus immaturus       2         Bambusae caulis in taenias       3         Bupleuri radix       3         Ctrir reiculatae pericarpium       2         Coptidis rhizoma       1         Glycyrrhizae radix       1         Glycyrrhizae radix       1         Glycyrrhizae radix       1         Pohlopogonis radix       3         Barbuse caulis in taenias       3         Glycyrrhizae radix       1         Glycyrrhizae radix       1         Pohlopogonis radix       3         Barbusoti (M& 2       2         Poriae sclerotium       3         Barbusoti (M& 2       2         Poriae sclerotium       3         Auskyokarschik (M& 2       2         Poriae sclerotium       9         Armeniace semen       9         9       9         Glycyrhizae radix       6         Glycyrhizae radix       6         Goreisan (Æ & 10)       2         Goreisan (Æ & 10)       10 </td <td>Platycodonis radix</td> <td>2 g</td> <td></td>	Platycodonis radix	2 g	
Scutellaria radix       2 g         Zingiberis rhizoma       1 g         Chikujountanto (竹每溫胆湯)       3         Aurantii furctus immaturus       2 g         Bambusae caulis in taenias       3 g         Bupleuri radix       3 g         Coptidis rhizoma       1 g         Coptidis rhizoma       1 g         Cyperi rhizoma       2 g         Ginseng radix       1 g         Ophiopogoins radix       3 g         Pinelliae rhizoma       5 g         Platycodi radix       2 g         Oprisongoins radix       3 g         Zingiberis rhizoma       1 g         Makyokansekito (旅音甘石湯)       Makyosekikanto (旅音甘石湯) H         rl	Poriae sclerotium	3 g	
Zingiberis rhizoma1 gChikujountanto (竹筎温即弱)2Aurantii fructus immaturus2 gBambusae caluis in taenias3 gBupleuri radix3 gCitri reticulatae pericarpium2 gCopidis rhizoma1 gCyperi rhizoma2 gGinseng radix1 gGlycyrrhizae radix1 gOphiopogonis radix3 gPinelliae rhizoma5 gPlatycodi radix2 gPoriae sclerotium3 gArmeniacae semen9 g4 gGlycyrrhizae radix6 gGypen ribrowa1 gArmeniacae semen9 g4 gGlycyrhizae radix6 gGypen ribrowa3 gInstantis rhizoma1 gArmeniacae semen9 g4 gGlycyrhizae radix6 gGypeum fibrosum18 gIo generative10 gGoreisan (五苓散)1Atractylodis macrocephale4 gPoriae sclerotium3 gPoriae sclerotium3 gHizoma1 gGinseng radix6 gGreisan (五苓散)1 gAtractylodis macrocephale4 gPoriae sclerotium4 gPoriae sclerotium2 gNosaikoto (小柴樹陽)1Upperi radix6 gGinseng radix2 gGingeng radix2 gGipcyrhizae radix2 gPoriae sclerotium2 gPoriae sclerotium2 gIbpleai radix2 g	Schisandrae fructus	1 g	
Chikujountanto (竹菊温胆湯)       2 g         Auranti fructus immaturus       2 g         Bambusae caulis in taenias       3 g         Bupleuri radix       3 g         Citri reticulatae pericarpium       2 g         Copidis fnizoma       1 g         Cyperi rhizoma       2 g         Ginseng radix       1 g         Ophiopogonis radix       3 g         Pinelliae rhizoma       5 g         Platycodi radix       2 g         Poriae sclerotium       3 g         Zingiberis rhizoma       1 g         Makyosekikanto (旅春甘石湯) r       ************************************	Scutellariae radix	2 g	
Aurantii fructus immaturus2 gBambusae caulis in taenias3 gBupleuri radix3 gCoptidis inizoma2 gCoptidis inizoma1 gCyperi rhizoma2 gGinseng radix1 gOphiopogonis radix3 gPinelliae rhizoma5 gPlatycodi radix2 gPoriae sclerotium3 gZingiberis rhizoma1 gMakyokansektio (methaffa)1 g $p^{-1}$ 3 gZingiberis rhizoma1 gMakyokansektio (methaffa)9 g $p^{-1}$ 9 gArmenjacae semen9 g9 g4 gGlycymhizae radix6 gGlycymhizae radix6 gGlycymhizae radix6 gImmaturus6 gGlycymhizae radix6 gArmerytolis nacrocephalae6 grhizoma3 gPoriae sclerotium4 grhizoma1 gCinnamoni cortex3 gPolypor sclerotium4 gShocaiktot (rhighi)2 gBupleuri radix6 gGinseng radix2 gShocaiktot (rhighi)2 gBupleuri radix2 gPoriae celerotium2 gBupleuri radix2 gPinelliae rhizoma2 gPinelliae rhizoma2 gPinelliae rhizoma2 gPinelliae rhizoma3 gPortex celerotium3 gPortex celerotium5 gPortex celerotium2 g	Zingiberis rhizoma	1 g	
Bambusae caulis in taenias       3 g         Bupleuri radix       3 g         Chri reticulatae pericarpium       2 g         Coptidis rhizoma       1 g         Cyperi rhizoma       2 g         Ginseng radix       1 g         Ophiopogonis radix       3 g         Pinelliae rhizoma       5 g         Pinelliae rhizoma       3 g         Zingiberis rhizoma       3 g         Pinelliae rhizoma       3 g         Zingiberis rhizoma       1 g         Makyosknsekito (#AETHTAB)       1 g         Makyosknsekito (#AETHTAB)       1 g         Ameniacae semen       9 g       4 g         Ephedrae herba       9 g       4 g         Glycyntrize radix       6 g       2 g         Gypuntibrosum       18 g       10 g         Goreisan (五苓散)       10 g       10 g         Goreisan (五苓散)       4 g       10 g         Cinnamoni cortex       3 g       10 g         Polyopi sclerotium       4 g       10 g         Shosaikoto (小柴樹湯)       10 g       10 g         Poriae sclerotium       4 g       10 g         Shosaikoto (小柴樹湯)       2 g       10 g         Shosai	Chikujountanto (竹茹温胆湯)		
Bupleuri radix3 gCtri reticulatae pericarpium2 gCoptidis rhizoma1 gCyperi rhizoma2 gGinseng radix1 gGlycyrrhizae radix1 gOphiopogonis radix3 gPinelliae rhizoma3 gPoriae sclearotium3 gPoriae sclearotium3 gZingiberis rhizoma1 gMakyoschikanto (條杏甘石湯) 中1 gArmeniacae semen9 g4 gEphedrae herba9 g4 gGlycyrrhizae radix6 g2 gGypsum fibrosum6 g2 gGoreian (五苓散)18 g10 gArmeniacie selentium4 gPoriae sclearotium6 gArractylodis macrocephalae4 grhizoma3 gPoriar sclearotium6 gShosaikoto (小柴胡湯)3 gPoriar sclearotium6 gJupbeleri radix6 gGinseng radix2 gJupber fractus6 gJupber fractus2 gPoriar sclearotium2 gPinelliae rhizoma2 gFinandir fractus2 gPinelliae rhizoma2 gPinelliae rhizoma2 gPinelliae rhizoma3 g	Aurantii fructus immaturus	2 g	
Ciri reticulatae pericarpium2 gCoptidis rhizoma1 gCyperi rhizoma2 gGinseng radix1 gOphiopogonis radix3 gPinelliae rhizoma5 gPlatycodi radix2 gPoriae sclerotium3 gZingberis rhizoma1 gMakyokansekito (麻杏甘石湯) 中Makyosekikanto (麻杏甘石湯) 市Armeniacae semen9 g4 gGlycyrrhizae radix6 g2 gGysum fibrosum18 g10 gGoreisan (五苓散)8 g10 gArractylodis macrocephalae rhizoma3 gCinnamomi cortex3 g3 gPoriae sclerotium4 gShosaikoto (小柴胡湯)4 gPoriae sclerotium2 gJupubae fructus2 gJupubae fructus5 g	Bambusae caulis in taenias	3 g	
Coptidis rhizoma1 gCyperi rhizoma2 gGinseng radix1 gGlycyrrhizae radix1 gOphiopogonis radix3 gPinelliae rhizoma5 gPlatycodi radix2 gPoriae sclerotium3 gZingiberis rhizoma1 gMakyokansekito (除杏甘石湯)*********************************	Bupleuri radix	3 g	
Cyperi rhizoma2 gGinseng radix1 gGlycyrrhizae radix1 gOptioogonis radix3 gPinelliae rhizoma5 gPlatycodi radix2 gPoriae sclerotium3 gZingiberis rhizoma1 gMakyoskansekito (孫杏甘石湯) 中7 gArmeniacae semen9 g4 gGlycyrthizae radix6 g2 gGysum fibrosum18 g10 gGoreisan (五苓散)1Alismatis rhizoma6 g2 gAnatopiona3 g10 gGoreisan (五苓散)4 gCinnamomi cortex3 g10 gPoriae sclerotium4 gPoriae sclerotium6 gGinseng (五苓散)4 gPoriae sclerotium6 gGinseng (五苓散)10 gJupibae fractura2 gJupibae fractura6 gGinseng (古苓散)1Jupibae fractura3 gPoriae sclerotium4 gShosaikoto (小學胡湯)1Jupibae fractura2 gJupibae fractura2 gJupibae fractura2 gJupibae fractura2 gJupibae fractura2 gJupibae fractura5 gShosaikoto2 gJupibae fractura5 gJupibae fractura5 gJupibae fractura5 g	Citri reticulatae pericarpium	2 g	
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Goreisan (五苓散)       6 g         Alismatis rhizoma       6 g         Atractylodis macrocephalae       4 g         rhizoma       3 g         Cinnamomi cortex       3 g         Polypori sclerotium       4 g         Poriae sclerotium       4 g         Shosaikoto (小柴胡湯)       5 g         Image: Polyporize radix       2 g         Glycyrrhizae radix       2 g         Jujubae fructus       2 g         Pinelliae rhizoma       5 g	Glycyrrhizae radix	6 g	2 g
Alismatis rhizoma       6 g         Atractylodis macrocephalae       4 g         rhizoma       3 g         Cinnamomi cortex       3 g         Polypori sclerotium       4 g         Poriae sclerotium       4 g         Shosaikoto (小柴胡湯)       6 g         Ginseng radix       6 g         Glycyrrhizae radix       2 g         Jujubae fructus       2 g         Pinelliae rhizoma       5 g		18 g	10 g
Atractylodis macrocephalae       4 g         rhizoma       3 g         Cinnamomi cortex       3 g         Polypori sclerotium       4 g         Poriae sclerotium       4 g         Poriae sclerotium       6 g         Shosaikoto (小柴胡湯)       6 g         Ginseng radix       2 g         Glycyrrhizae radix       2 g         Jujubae fructus       2 g         Pinelliae rhizoma       5 g	Goreisan (五苓散)		
rhizoma Cinnamomi cortex 3 g Polypori sclerotium 4 g Poriae sclerotium 4 g Shosaikoto (小柴胡湯) Bupleuri radix 6 g Ginseng radix 2 g Glycyrrhizae radix 2 g Jujubae fructus 2 g	Alismatis rhizoma		
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Poriae sclerotium     4 g       Shosaikoto (小柴胡湯)     6 g       Bupleuri radix     6 g       Ginseng radix     2 g       Glycyrrhizae radix     2 g       Jujubae fructus     2 g       Pinelliae rhizoma     5 g	Cinnamomi cortex	3 g	
Shosaikoto (小柴胡湯)         Bupleuri radix       6 g         Ginseng radix       2 g         Glycyrrhizae radix       2 g         Jujubae fructus       2 g         Pinelliae rhizoma       5 g	Polypori sclerotium	4 g	
Bupleuri radix6 gGinseng radix2 gGlycyrrhizae radix2 gJujubae fructus2 gPinelliae rhizoma5 g		4 g	
Ginseng radix2 gGlycyrrhizae radix2 gJujubae fructus2 gPinelliae rhizoma5 g	Shosaikoto (小柴胡湯)		
Glycyrrhizae radix2 gJujubae fructus2 gPinelliae rhizoma5 g	Bupleuri radix	6 g	
Jujubae fructus2 gPinelliae rhizoma5 g		2 g	
Pinelliae rhizoma 5 g		2 g	
6	Jujubae fructus	2 g	
Scutellariae radix 3 g			
	Scutellariae radix	3 g	

Table 7 continued

Zingiberis rhizoma	4 g	
Yakanmaoto (射干麻黄湯)		
Asiasari radix	3 g	
Asteris radix et rhizoma	6 g	
Belamcandae rhizoma	9 g	
Ephedrae herba	9 g	
Farfarae flos	6 g	
Jujubae fructus	3	fruits
Pinelliae rhizoma	9 g	
Schisandrae fructus	3 g	
Zingiberis rhizoma	9 g	
Bofutsushosan (防風通聖散)		
Angelicae sinensis radix	2 g	
Atractylodis macrocephalae rhizoma	3 g	
Cnidii rhizoma	2 g	
Ephedrae herba	2 g	
Forsythiae fructus	2 g	
Gardeniae fructus	2 g	
Glycyrrhizae radix	2 g	
Gypsum fibrosum	3 g	
Menthae haplocalycis herba	2 g	
Natrii sulfus	2 g	
Paeoniae radix	2 g	
Platycodi radix	2 g	
Rhei rhizoma	2 g	
Saposhnikoviae radix	2 g	
Schizonepetae herba	2 g	
Scutellariae radix	2 g	
Talcum crystallinum	5 g	
Zingiberis rhizoma	2 g	

### Appendix 2

See Table 8.

Table 8 Taxonomy of East Asian herbal drugs

Drug	Kanji	Accepted name https://mpns.science.kew.org/mpns-portal/		
Agastachis herba	藿香	Agastache rugosa (Fisch. & C.A.Mey.) Kuntze		
Alismatis rhizoma	澤瀉	Alisma plantago-aquatica subsp. orientale (Sam.) Sam		
Andrographitis herba	穿心蓮	Andrographis paniculata (Burm.f.) Nees		
Angelicae sinensis radix	當歸	Angelica sinensis (Oliv.) Diels		
Armeniacae semen	杏仁	Prunus armeniaca L.		
Asiasari radix	細辛	Asarum sieboldii Miq		
Asparagi radix	天門冬	Asparagus cochinchinensis (Lour.) Merr		
Asteris radix et rhizoma	紫菀	Aster tataricus L.f		
Astragali radix	黄芪	Astragalus mongholicus Bunge		
Atractylodis macrocephalae rhizoma	白朮	Atractylodes macrocephala Koidz		
Aurantii fructus immaturus	枳實	Citrus  imes aurantium L.		
Bambusae caulis in taenias	竹茹	Bambusa beecheyana Munro		
Belamcandae rhizoma	射干	Iris domestica (L.) Goldblatt & Mabb		
Bupleuri radix	柴胡	Bupleurum falcatum L. (used in Japan & Korea)		
		Bupleurum chinense DC. (used in China)		
Cimicifugae rhizoma	升麻	Actaea dahurica (Turcz. ex Fisch. & C.A.Mey.) Franch		
Cinnamomi cortex	桂皮	Cinnamomum cassia (L.) J.Presl		
Cinnamomi ramulus	桂枝	Cinnamomum cassia (L.) J.Presl		
Citri reticulatae pericarpium	陳皮	Citrus × aurantium L.		
Cnidii rhizoma	川芎	Ligusticum officinale (Makino) Kitag		
Coptidis rhizoma	黄連	Coptis chinensis Franch		
Cyperi rhizoma	香附	Cyperus rotundus L.		
Dioscoreae rhizoma	山薬	Dioscorea japonica Thunb		
Ephedrae herba	麻黄	Ephedra sinica Stapf		
Farfarae flos	款冬	Tussilago farfara L.		
Forsythiae fructus	連翹	Forsythia suspensa (Thunb.) Vahl		
Fritillariae thunbergii bulbus	浙貝母	Fritillaria thunbergii Miq		
Gardeniae fructus	梔子	Gardenia jasminoides J.Ellis		
Ginseng radix	人参	Panax ginseng C.A.Mey		
Glycyrrhizae radix	甘草	Glycyrrhiza uralensis Fisch. ex DC		
Houttuyniae herba	十薬	Houttuynia cordata Thunb		
Isatidis radix	藍草	Isatis indigotica Fort		
Jujubae fructus	大棗	Ziziphus jujuba Mill		
Lonicerae flos	金銀花	Lonicera japonica Thunb		
Lycii radicis cortex	地骨皮	Lycium chinense Mill		
Menthae haplocalycis herba	薄荷	Mentha canadensis L.		
Mori radicis cortex	桑白皮	Morus alba L.		
Ophiopogonis radix	麥冬	Ophiopogon japonicus (Thunb.) Ker Gawl		
Paeoniae radix	芍薬	Paeonia lactiflora Pall		
Patriniae herba	敗醤葉	Patrinia scabiosifolia Fischer ex Treviranus		
Phragmitis rhizoma	蘆根	Phragmites communis Trin		
Pinelliae rhizoma	半夏	Pinellia ternata (Thunb.) Makino		
Platycodonis radix	桔梗	Platycodon grandiflorus (Jacq.) A.DC		
Polygoni cuspidati rhizoma	虎杖根	Polygonum cuspidatum Sieb.et Zucc		
Polypori sclerotium	豬苓	Polyporus umbellatus (Pers.) Fries		

Table 8 continued

Drug	Kanji	Accepted name https://mpns.science.kew.org/mpns-portal/	
Poriae sclerotium	茯苓	Poria cocos (Schw.) Wolf	
Puerariae radix	葛根	Pueraria montana var. lobata (Willd.) Maesen & S.M. Almeida ex Sanjappa & Predeep	
Rhei rhizoma	大黄	Rheum palmatum L.	
Saposhnikoviae radix	防風	Saposhnikovia divaricata (Turcz. ex Ledeb.) Schischk	
Schisandrae fructus	五味子	Schisandra chinensis (Turcz.) Baill	
Schizonepetae herba	荊芥	Nepeta tenuifolia Benth	
Scutellariae radix	黄芩	Scutellaria baicalensis Georgi	
Verbenae herba	馬鞭草	Verbena officinalis L.	
Zingiberis rhizoma	生姜	Zingiber officinale Roscoe	

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