

# What's new in the aetiopathogenesis of vasculitis?

Paul A. Brogan

Received: 12 December 2006 / Revised: 5 January 2007 / Accepted: 8 January 2007 / Published online: 15 March 2007  
© IPNA 2007

**Abstract** The cause of the majority of childhood vasculitides is unknown although it is likely that a complex interaction between environmental factors and inherited host responses trigger the disease and determine the vasculitis phenotype. Epidemiological clues continue to implicate infectious triggers in Kawasaki syndrome (KS) and Henoch Schönlein purpura (HSP). Several genetic polymorphisms have now been described in KS and HSP which predispose to disease or predict disease severity. Anti-neutrophil cytoplasmic antibodies (ANCA) are now known to be directly involved in the pathogenesis of vascular injury in ANCA-associated vasculitides, although why some individuals develop ANCA in the first instance is not yet understood. Endothelial injury and repair are active areas of research in vasculitis. It is now possible to track endothelial injury non-invasively in children with vasculitis using surrogate markers of endothelial injury. The vasculogenic pathways involved in vascular repair following vasculitis, including endothelial progenitor cells, are beginning to be studied. It is anticipated that an improved understanding of the aetiopathogenesis of vasculitis in the young will ultimately shape future novel diagnostic and therapeutic approaches and will help us predict which children may develop premature arteriosclerosis in later life.

**Keywords** Aetiology · Child · Pathogenesis · Vasculitis

---

P. A. Brogan (✉)  
Department of Rheumatology, Institute of Child Health, Level 6,  
30 Guilford St.,  
London WC1N 1EH, UK  
e-mail: p.brogan@ich.ucl.ac.uk

## Introduction

Primary systemic vasculitis (PSV), while rare in children, is still associated with significant mortality and morbidity. Primary vasculitic syndromes include, amongst others, Henoch Schönlein purpura (HSP), Kawasaki syndrome (KS), polyarteritis nodosa (PAN), Takayasu disease (TD) and the anti-neutrophil cytoplasmic antibody (ANCA)-associated vasculitides (AAV), comprising Wegener's Granulomatosis (WG), microscopic polyangiitis, renal limited vasculitis and Churg-Strauss syndrome [1]. PSV is characterised by the presence of inflammation in the walls of blood vessels, with resultant tissue ischaemia and necrosis [1]. The cause of the majority of childhood vasculitides is unknown, although it is likely that a complex interaction between environmental factors, such as infections and inherited host responses, trigger the disease and determine the vasculitis phenotype [2, 3] This review summarises the findings of recent studies relating to the aetiopathogenesis of PSV and will be considered under the two broad headings of environmental triggers and host responses. This article will not focus on the therapy of PSV, nor will the article describe comprehensively all of the possible infective organisms now known to be associated with secondary infective vasculitis (very recently reviewed by Pagnoux et al. [4] and Naides [5]).

## Environmental triggers

### Epidemiological clues

A number of important epidemiological studies have recently been published describing the demographics of

PSV in the young [6–8], but as yet these have failed to definitively cast light on the aetiology of PSV in children. Epidemiological data are most readily available for the two commonest forms of PSV in children, HSP and KS, with relatively few data relating to the epidemiology of PAN or AAV in children. Common themes emerging relate to seasonality and ethnic predisposition, in particular in relation to KS, hinting at infectious precipitants and/or genetically determined host-responses. KS is commonest in Japan and in Japanese and other Oriental children living abroad [9]. Children aged 6 months to 5 years are the most susceptible, with peak incidence in children aged 9–11 months [10, 11]. Seasonal variation in KS incidence has been reported, with peak occurrence in the winter and spring months [10, 11]. Outbreaks of KS have been linked to weather patterns, with clusters of KS cases occurring in association with precipitation [12]. Direct person-to-person spread has not been observed, although in Japan the disease occurs more commonly in siblings of index cases, with an estimated peak incidence of 8–9% in siblings under the age of 2 years [13]. Currently, the estimated incidence of KS in the UK is 8.1 per 100,000 children under the age of 5 years – an incidence which has doubled since 1991 [14]. While this increase may truly indicate an increasing incidence, better case recognition may partly account for this rising trend. Whilst these observations suggest an infectious aetiology, the hunt for any single infectious agent has so far not proved fruitful, a fact most recently highlighted by the negative results that emerged from studies examining the potential link between coronavirus infection and KS in Taiwan [15]. It is likely that KS represents an extreme endpoint of any number of potential infectious agents in a genetically predisposed individual. This has prompted the recent change of nomenclature from Kawasaki disease to Kawasaki syndrome, which serves to focus attention on host responses rather than specific infectious agents.

HSP, similar to KS, also appears to have a seasonal variation, occurring more commonly in the autumn and winter [7, 16]. Yang et al. commented that the clustering of HSP in the autumn and winter, often preceded by upper respiratory infections, points to the possibility that HSP is infection-related [7], possibly to inciting agents such as streptococci [17], *Bartonella henselae* [18], parvovirus [19], HIV [20] and varicella [21], amongst others. As in KS, however, no single infectious agent has been identified, and it is likely that genetically controlled host responses determine whether or not an individual develops HSP in response to infectious triggers.

Epidemiological studies of AAV in adults have provided important clues relating to other environmental and genetic factors (see below) which may be involved in the aetiopathogenesis of these syndromes. The geographic

distribution of AAV disease occurrence in adults suggests that WG occurs more commonly in northern European countries than in southern European countries, with a twofold higher annual incidence being observed in Norway and England than in Lugo, Spain [22, 23]. The human leukocyte antigen (HLA) make-up of the Lugo population differed from that found in other Mediterranean areas and had a similar proportion of HLA-DRB1 alleles to that found in the UK population. Since no clear HLA association has been shown with any type of PSV other than Behcet's disease, Watts et al. suggest that environmental factors may be more important than genetic factors in WG [22, 23].

In this context there is a picture emerging of a potential relationship between occupational crystalline silica exposure and AAV that has been observed in a number of studies [24–28]. Overall exposure to silicon-containing compounds was found to be related to AAV, particularly WG, with an odds ratio of 2.5–5 [25–28]. The mechanisms by which silica may induce AAV are not well known. Silicon-containing compounds may be immune adjuvants, since silica particles are potent stimulators of lymphocytes, monocytes and macrophages. Silica particles may also stimulate neutrophil-reactive oxygen radical release and inflammatory cytokines, triggering exocytosis of the proteinase 3 and myeloperoxidase antigens on the surface of the polynuclear neutrophils, thus priming neutrophils for ANCA activation (see below). Whether this risk factor is important in WG in children or young adults is, however, questionable.

Other miscellaneous environmental risk factors for AAV in adults have been reported, such as smoking, which has been found by some researchers to have either a protective effect [29] or no effect at all [30, 31], putative triggering of WG by drug exposure, notably propylthiouracil or hydralazine [32], and links between malignancy and onset of WG [33]. These factors are, however, likely to be of less relevance as risk factors for paediatric AAV.

#### Infections and PSV

Many pathogens are known to either directly cause vasculitis or to act as potential triggers for PSV [4, 5]. Many of the viruses, bacteria and fungi that directly cause vasculitis demonstrate tissue tropism that includes the endothelium; other agents may bind to the vessel wall because the vascular endothelium expresses specific receptors for the pathogen. Even when the agent does not enter the endothelial cell, the immune response to the agent may still be focused at the endothelium because the pathogen is adherent to the endothelial cell surface, resulting in bystander injury to the endothelium and vessel.

## Superantigens and PSV

Superantigens (SAGs) are one of the environmental factors that have been proposed to modulate a number of autoimmune diseases, including vasculitis. The most compelling evidence for the involvement of SAG in the pathogenesis of vasculitis relates to KS, although this hypothesis has proved to be controversial [34]. SAGs are a class of immuno-stimulatory proteins of bacterial or viral origin with the ability to activate large fractions (5–30%) of the T cell population, and they are responsible for human toxic shock syndrome and some forms of gastroenteritis [3, 35]. Class II major histocompatibility complex (MHC)-positive endothelial cells operate as competent SAG-presenting cells for CD4 and CD8 lymphocytes *in vitro*. Dual signalling between endothelial cells and T cells results in V $\beta$ -restricted activation and adherence to endothelium, resulting in endothelial cell activation and injury [3]. Thus, SAGs could mechanistically result in severe endothelial and vascular injury and could account, in part, for the vascular injury associated with SAG-mediated diseases such as toxic shock syndrome.

One important controversial issue is the ongoing debate regarding SAGs versus conventional antigens in the aetiopathogenesis of KS. There are similarities between the clinical and immunological features of KS and SAG toxin-mediated staphylococcal and streptococcal toxic shock syndromes, and scarlet fever [3].

The study by Abe et al. in 1992 was the first to describe the selective expansion of V $\beta$ 2 and V $\beta$ 8.1 T cells in KS [36], indicating T cell V $\beta$  skewing – the hallmark of a SAG-mediated process. Since then, many similar studies have examined T cell V $\beta$  repertoires in KS, or examined the prevalence of serological conversion or colonisation with SAG-producing organisms [37, 38]. Another important observation is that T cells infiltrating the walls of coronary arterial aneurysms and the intestinal mucosa of patients with KS show a skewed T cell V $\beta$  repertoire, with increased numbers of cells expressing V $\beta$ 2 [39, 40], again possibly indicative of a SAG-driven process.

However, Rowley et al. recently reported three fatal cases of KD and observed IgA plasma cell infiltration into the vascular wall during the acute phase of the illness. By examining the clonality of this IgA response using reverse transcriptase (RT)-PCR in lesional vascular tissue, these researchers observed that the IgA response was oligoclonal, suggesting a conventional Ag process rather than a SAG-driven one [41]. Synthetic versions of the prevalent IgA antibodies were then used in immunohistochemical experiments on acute KS and control tissues; several of the most prevalent IgA antibodies detected

antigen in lesional KS tissue, but not in control tissue. Further studies indicated that the antigen resides in cytoplasmic inclusion bodies [42], which is possibly consistent with aggregates of viral protein and nucleic acid. These important observations may suggest that there is a common infectious aetiology for KS, perhaps viral, and may cast doubt on the SAG hypothesis in KS.

We recently described the T cell V $\beta$  repertoires in a number of paediatric vasculitic syndromes [2]. Like others, we found evidence of V $\beta$ 2 expansions in peripheral blood of KS patients and skewing of the peripheral blood T cell V $\beta$  repertoire in 25 children with PSV, predominantly PAN. No such skewing of the V $\beta$  repertoire was observed in HSP patients. While these data provide impetus for further research into this contentious field, they do not resolve unequivocally the question of the role of SAGs in childhood vasculitic syndromes. That said, a recent multi-center review of Turkish children with PAN again has implicated streptococcal infection in the aetiopathogenesis of this vasculitic syndrome. Whether the mechanism is that of molecular mimicry of a conventional antigen or superantigen-mediated vascular injury is unknown [43]. Thus, the jury remains out on the SAG hypothesis in PSV of the young.

## *Staphylococcus aureus* and WG

Several groups have been actively researching the relationship between *Staphylococcus aureus* and WG [44]. There is an increased incidence of chronic nasal carriage of *S. aureus* in patients with WG, and this is associated with an increased risk of disease relapse [45]. Moreover relapses of WG can be reduced by the use of long-term trimethoprim-sulphamethoxazole [46], although it is currently unclear whether this efficacy results from eradication of the nasal carriage of *S. aureus* or by virtue of a potential anti-inflammatory effect of trimethoprim-sulphamethoxazole, which exerts its antibiotic effect by antagonising folic acid metabolism, a mechanism which could affect leucocytes. Possible mechanisms whereby *S. aureus* could result in flares of WG include SAG production and T and B cell activation, direct tropism of *S. aureus* for endothelial cells, with binding and internalisation of the organism by endothelial cells, or by priming of neutrophils (see below) [44]. Of note, however, is the observation that the carriage of staphylococcal strains that are positive for SAG genes in WG patients is not associated with the expected pattern of SAG-reactive T cells [47]. Of all these proposed mechanisms, the data currently suggest that the effects of *S. aureus* on endothelial cells and/or phagocytic cells may be the most plausible, with relatively scant evidence of a SAG effect based on studies of patterns of T cell V $\beta$  skewing.

## Host responses

### Genetic predisposition to vasculitides

Several investigators have examined potential genetic factors predisposing to vasculitis. Familial cases of vasculitis including PAN [48], KS [49], AAV [50] and TD [51] have been described. It is clear from these studies, however, that the predisposing genetic factors of PSV are complex and multiple and that they vary between different vasculitic syndromes. Previously genetic studies had focused on AAV, although a number of publications have recently appeared that report on important mutations and/or polymorphisms in KS and HSP.

### KS and novel genetic associations

Biezeveld et al. recently reported on the association of a mannose-binding lectin (MBL) genotype with cardiovascular abnormalities in KS [52, 53]. MBL is a macromolecule with collagenous and sugar-binding domains and is able to activate the complement pathway directly (i.e. independent of antibodies). MBL is thus the “third” limb of the complement cascade and is termed the MBL pathway. In particular, MBL binds to repeats of N-acetyl-glucosamine and mannose residues, expressed on the surface of many microbial antigens. A deficiency of MBL (found in up to a third of the healthy population) is associated with an increased risk of bacterial and viral infection in children [54]. Thus, MBL is an important component of the innate immune system, particularly in the acute phase of infections. Infants in the period between decreasing levels of maternal antibodies and the development of adaptive immunity are especially dependent on this molecule. In two studies of 90 Caucasian children with KS in The Netherlands, Biezeveld et al. observed what appears to be an ambiguous role for MBL in this context [52]. Infants with KS under 1 year of age were at higher risk of developing coronary artery aneurysms (CAA) if polymorphisms in the *MBL2* gene (correlating with low plasma MBL levels) were present (odds ratio: 15.7; 95% confidence interval: 1.4–176.5). Over the age of 1 year the reverse was true: the wild-type MBL genotype and normal plasma MBL levels were associated with a greater risk of CAA [53]. Thus, it appears that MBL modulates host responses and cardiovascular injury differently depending on age. In infants under the age of 1 year, normal MBL levels may protect from any putative infective organism triggering vascular injury in KS; whereas in older children who are not critically dependent on MBL for innate immunity, MBL, itself an acute phase protein, could contribute to endothelial injury by uncontrolled complement activation [53].

To date, there are only limited data examining the role of the MBL genotype in other vasculitides. Endo et al. detected the glomerular deposition of MBL and MBL-associated serine protease (MASP-1) as well as C3b/C3c, C5b-9, and C4-binding protein (C4-bp) in eight of ten patients with HSP nephritis, suggesting a role for MBL in glomerular injury associated with HSP [55].

Mutations and polymorphisms in several other miscellaneous genes have recently been described in KS, either as disease-susceptibility genes influencing the risk for CAA development or as a response to treatment. These are summarised in Table 1.

### Genetic polymorphisms and HSP

Several polymorphisms relating to disease susceptibility, severity and/or risk of renal involvement have recently been described. Many of these polymorphisms relate to cytokines or cell adhesion molecules involved in the modulation of inflammatory responses and endothelial cell activation, and may therefore have relevance in other autoimmune diseases as well as HSP. On the whole, studies of this nature have been hampered by relatively small patient numbers and thus lack the power to be definitive or necessarily applicable to all racial groups. These studies are summarised in Table 1.

### Genetic polymorphisms and ANCA associated vasculitides

A plethora of studies have examined multiple putative genes that may be involved in the aetiopathogenesis of AAV. These are summarised in Table 1.

### Autoantibodies and vascular injury: ANCA

Only recently has the evidence that ANCA are directly involved in the pathogenesis of vascular injury been documented. The most compelling evidence are reports of two neonates who developed pulmonary renal syndrome soon after birth from the trans-placental transfer of IgG MPO-ANCA from mothers with active microscopic polyangiitis (MPA) [56, 57]. Moreover, a mouse model recently described by Xiao et al. showed that the passive transfer of MPO-ANCA or anti-MPO lymphocytes (that make MPO-ANCA) into mice that lack functioning B or T cells (RAG  $-/-$ ) resulted in pauci-immune crescentic nephritis and small vessel vasculitis similar to that observed in humans with AAV [58]. Thus, the evidence that MPO-ANCA are directly involved in the pathogenesis of vascular injury is growing, but may still be somewhat debated for PR3-ANCA.

A plethora of publications describe in detail the mechanism of vascular injury mediated by ANCA, and

**Table 1** Genetic polymorphisms studied in KS, HSP, and AAV<sup>a</sup>

Molecule/genetic polymorphism	Vasculitis type		
	KS	HSP	AAV
Mannose binding lectin (MBL)	Ambiguous role for MBL influencing risk of coronary artery aneurysms (CAA); see main text	MBL and MBL-associated serine protease (MASP-1) detected in glomerular lesions of HSP- see main text	Not studied
Angiotensin converting enzyme (ACE)	ACE I/D polymorphism increases disease susceptibility [106]	No association of HSP nephritis with polymorphisms in ACE, albeit in studies involving small numbers of children [107–109]	Not studied
Matrix metalloproteinases (MMP)	MMP-3 6A/6A polymorphism results in higher frequency of CAA [110]	Genetics not studied; MMP-9 may be elevated in HSP [111]	Genetics not studied; MMPs expressed in lesional glomerular tissue [112]
Vascular endothelial growth factor (VEGF) and its receptor (KDR)	Polymorphisms of both contribute to increased CAA risk [113]	VEGF polymorphisms predispose to renal involvement [114]	Genetics not studied; VEGF elevated in active WG [115]
Interleukin 1 receptor antagonist (IL-1Ra)	Polymorphism associated with increased disease susceptibility [116]	Polymorphism predisposes to renal involvement [117]	Not studied
Interleukin 1β (IL-1β) [118]	No association found [116]	Polymorphism associated with renal involvement [118]	One study failed to identify associations between IL-1β polymorphisms and WG [119]
Tumour necrosis factor-alpha (TNF-α)	TNF-α-308A associated with increased intravenous immune globulin (IVIG) resistance [120]	TNF-alpha G-308A polymorphism not associated with HSP in Chinese patients [121]	One study failed to identify associations between TNFα polymorphisms and WG [119]
Interleukin-8 (IL-8)	Genetics not studied	Polymorphism associated with renal involvement [122]	Genetics not studied
Interleukin-10 (IL-10)	IL-10 gene promoter polymorphisms influence risk of CAA [120]	Genetics not studied	IL-10 (-1082) polymorphism associated with WG and MPA [123]
Chemokines	Chemokine receptor CCR5 and its ligand CCL3L1 influence disease susceptibility [124]	Genetics not studied	Genetics not studied
Familial Mediterranean Fever genotypes (MEFV gene mutation)	Genetics not studied	Mutations in MEFV found more commonly in Israeli and Turkish children with HSP [125, 126]	Genetics not studied
Human Leucocyte Antigens (HLA)	No consistent associations	Positivity for HLA-B35 predisposes to renal involvement in HSP [127]	No consistent associations
PAX2 (Paired box gene 2)	Genetics not studied	Polymorphisms in PAX2 predispose to renal involvement in HSP [128]	Genetics not studied
Nitric oxide and associated molecules	No association of eNOS and iNOS gene polymorphisms to the development of CAL in Japanese KS patients [129]	Inducible nitric oxide synthase 2A promoter polymorphism predisposes to renal involvement [130]	Genetics not studied
Cell adhesion molecules	Genetics not yet studied	Patients not carrying the codon ICAM-1 469 K/E genotype are at decreased risk of developing severe gastrointestinal complications [131]	Polymorphism in exon 11 of the CD18 gene associated with MPO-ANCA vasculitis; no relevant polymorphisms were identified for ICAM-1, E-selectin, CD11b, or human urokinase plasminogen activator receptor gene [132]

**Table 1** (continued)

Molecule/genetic polymorphism	Vasculitis type		
	KS	HSP	AAV
$\alpha$ -1-Antitrypsin	Genetics not studied	Isolated case reports of severe multi-systemic HSP and $\alpha$ -1-antitrypsin deficiency [133]	An association between PR3-ANCA and the deficient PiZZ phenotype has been described [134, 135]. Alpha-1-antitrypsin deficiency is not sufficient in itself to cause ANCA-positive vasculitides, but may act as an amplifying factor [136, 137]
Proteinase 3 (PR3)	Genetics not studied	Genetics not studied	Association with the A-564G polymorphism in the proteinase-3 promoter and WG [138]
Fc $\gamma$ receptors	No association for Fc $\gamma$ RIIA-131H/R, Fc $\gamma$ RIIB-232I/T, Fc $\gamma$ RIIIA-158 V/F and Fc $\gamma$ RIIIB-NA1/NA2 [139]	Genetics not studied	Possible association between NA1 allele of Fc $\gamma$ RIIIB in patients with WG and renal involvement [140]. Fc $\gamma$ R11-R131 and Fc $\gamma$ R11A-F158 may predispose to disease relapse [141].
CTLA-4 (cytotoxic T lymphocyte-associated antigen-4)	Genetics not studied	Genetics not studied	Polymorphism associated with WG [119]

<sup>a</sup> KS, Kawasaki Syndrome; HSP, Henoch Schönlein purpura; AAV, anti-neutrophil cytoplasmic antibody (ANCA)-associated vasculitides

these have been eloquently summarised in recent reviews [59, 60]. The most accepted model of pathogenesis proposes that ANCA activate cytokine-primed neutrophils, leading to bystander damage of endothelial cells and an escalation of inflammation with recruitment of mononuclear cells [61]. Neutrophil priming involves stimulation with an agent, such as lipopolysaccharide (LPS) or the cytokine tumour necrosis factor alpha (TNF- $\alpha$ ), at a concentration that does not of itself cause full functional activation but rather results in the translocation of ANCA-antigen (MPO and/or PR3) from intracellular granules to the cell surface. TNF- $\alpha$  priming and subsequent ANCA activation of neutrophils lead to a respiratory burst and degranulation, thereby generating the potential for endothelial injury. The binding of ANCA to antigens expressed on the surface of primed neutrophils is not enough in itself to cause full neutrophil activation; the binding of Fc receptors is also necessary [62]. Following neutrophil stimulation by ANCA, numerous cytotoxic mediators are released, including reactive oxygen species, chemokines, cytokines, proteolytic enzymes and nitric oxide (NO) [63]. The firm adhesion of activated neutrophils to endothelial cells results in endothelial cell damage and perhaps recruitment of other inflammatory cells including monocytes and T cells [64]. Following neutrophil activation by ANCA, neutrophils are driven down an accelerated but aberrant apoptotic pathway [65]. Thus, the neutrophils will develop the morphologic

features of apoptosis, but there is a failure in cell surface changes, such as bi-lipid cellular membrane phosphatidylserine (PS) externalisation [66, 67], which is normally an early feature of all apoptotic cells. This lack of PS externalisation means that phagocytes are less able to process the apoptotic neutrophils in a non-phlogistic fashion, explaining the well-characterised finding of leucocytoclasia that is often seen in the vasculitic lesions of certain vasculitic syndromes.

#### Autoantigen complementarity and AAV

The aforementioned clinical and experimental evidence support a role for ANCA in the pathogenesis of AAV, but it does not explain why ANCA develop in certain individuals. One emerging hypothesis which could cast light on this area is termed the complementarity theory [68]. The central premise for this theory is that anti-PR3 autoantibodies (PR3-ANCA) are generated by an immune response that initially is mounted against a peptide that is antisense or “complementary” to the autoantigen [68]. The implication is that infectious agents could trigger ANCA generation via microbial peptides which act as molecular mimics for complementary PR3. Indeed *S. aureus* contains a peptide sequence that mimics antisense PR3, as do the Ross River virus and *Entamoeba histolytica*, which are all infections previously associated with ANCA generation in

humans [69]. The theory of autoantigen complementarity could thus have important implications for the pathogenesis of other autoimmune diseases, but this needs further validation.

#### Downstream mediators of vascular injury

A number of studies have looked at other biomarkers predictive of CAA in KS. Two biomarkers deserve particular mention in this context: the matrix metalloproteinases (MMPs) and the S-100 proteins.

An imbalance between the MMPs and the tissue inhibitor of MMP (TIMP) has been implicated in the development of CAA in KS [70]. The extracellular matrix is maintained by a rigorously controlled balance between the synthesis and breakdown of its component proteins. MMPs and TIMPs play central roles in this process. Children with KS and high levels of MMP and/or a high MMP/TIMP ratio are more susceptible to coronary arterial lesions [71].

Calcium-binding proteins in the S-100 family, MRP-8 and MRP-14, are potential biomarkers for KS and CAA. These proteins form heterodimers and are secreted by neutrophils and monocytes in response to inflammatory signalling cascades. The MRP8/MRP14 heterodimer binds to microvascular endothelial cells and may participate in the genesis of a proinflammatory and prothrombotic state during systemic vasculitis [72]. Specifically, these proteins regulate the adhesion of neutrophils and monocytes to endothelial cells and are implicated in their transmigration into the vessel wall. Serum MRP-8/MRP-14 levels as well as mRNA expressions of MRP-8 and -14 in granulocytes were upregulated in acute KS and decreased dramatically within 24 h of intravenous immune globulin (IVIG) therapy [72]. The number of MRP-8/MRP-14-positive circulating endothelial cells (CECs, see below) was higher in patients with acute KD than in control patients, and the numbers increased significantly by 2 weeks after the onset of KD, especially in patients in whom coronary artery lesions developed [72].

One important implication of these observations is that these pathways could provide new therapeutic targets for IVIG non-responders or those with severe CAA and partial IVIG response.

#### Tumour necrosis factor alpha and vasculitis

There is much interest in the role of TNF- $\alpha$  as a downstream mediator of endothelial and vascular injury and the potential for a therapeutic blockade of this cytokine in a number of paediatric and adult vasculitic syndromes [73, 74]. The evidence for efficacy of TNF- $\alpha$  blockade in paediatric vasculitis currently remains anecdotal – albeit promising – and an important addition to the armamentar-

ium for diseases recalcitrant to conventional therapy [75, 76].

#### Tracking endothelial injury in vasculitis

A number of groups have recently described methods for detecting endothelial cell components in blood which may allow the assessment of the molecular events associated with vascular injury. Two potentially informative methods have been reported: endothelial microparticles (EMPs) and circulating detached mature endothelial cells (CECs).

EMPs are released from the cell surface in response to a variety of stimuli associated with endothelial injury, including vasculitis [77], atherosclerosis and acute coronary events [78] and thrombotic thrombocytopenic purpura [79]. We have previously demonstrated that it is possible to monitor endothelial injury in vasculitis in children through the detection of circulating EMPs expressing E-selectin, or CD105, in effect providing a window to the activated endothelium [80].

CECs are necrotic or highly activated endothelial cells which detach from the vessel wall [81]. They have recently been shown to be a useful clinical marker of endothelial injury in vasculitis in adults [82–84], stroke [85] and pulmonary hypertension [86]. Woywodt et al. suggest that CECs correlate with disease activity in adults with ANCA vasculitis and are thus useful both diagnostically and also for monitoring disease activity in response to therapy [84]. Our preliminary data (PB et al., unpublished) suggest that CECs are also elevated in children with active primary systemic vasculitis and that they may also be used as a novel clinical tool to monitor disease activity in response to treatment.

#### Vascular repair mechanisms

A recently emerging concept now supported by several animal and human studies is that bone marrow-derived endothelial progenitor cells (EPCs) are involved in both normal physiological and pathophysiological repair mechanisms, for example during myocardial vascular regeneration following myocardial infarction and in post-transplant arteriosclerosis [85–87]. The discovery of EPCs by Asahara et al. represented a major advance which has altered our understanding of endothelial repair and the process of neovascularisation and focused considerable research interest in these areas [88]. These researchers showed that CD34 + peripheral blood mononuclear cells from healthy adults acquire an endothelial cell-like phenotype *in vitro* and are incorporated into regenerating capillaries in a murine hind-limb ischaemia model, findings subsequently confirmed by several groups.

The number and the function of circulating EPCs influence cardiovascular risk and endothelial responses to insults associated with endothelial injury in humans. Disease and pre-disease states studied to date include: conventional atherosclerotic risk factors [87, 89]; connective tissue/inflammatory diseases, including rheumatoid arthritis [90]; systemic sclerosis [91]; Wegener's granulomatosis [92]; Kawasaki disease [93] (although data relating to children are limited to this latter small study). EPCs are known to originate in the bone marrow, and they function normally via homing and adherence to regions of damage [94, 95]. EPC responses to the treatment of vasculitis in children have yet to be studied, but it is an area of ongoing active research.

Does vasculitis in the young predispose to premature atherosclerosis?

Several key aspects of the long-term outcome of vasculitis in the young remain of ongoing concern and are currently being best studied in KS. Most importantly, controversy continues as to whether KS constitutes a risk factor for premature atherosclerosis. Histological findings seen in KS arteries at sites of previous aneurismal lesions long after disease resolution appear to be indistinguishable from atherosclerosis [96]. Dhillon et al. studied vascular responses to reactive hyperaemia in the brachial artery using high-resolution ultrasound [97]. Flow-mediated dilation (an endothelial-dependent response) was reduced in KS patients compared with control subjects many years after the illness, even in patients without detectable early coronary artery involvement. In a long-term case control study, Albisetti et al. showed that KS patients were more likely to have impairment of the fibrinolytic system, another marker of endothelial dysfunction, and this again was unrelated to the degree of coronary artery involvement [98]. Pilla et al. demonstrated reduced arterial distensibility (an independent risk factor for cardiovascular morbidity and mortality in adults), as assessed using ultrasound pulse wave velocity in the brachio-radial arterial segments of 43 children who had KD a median of 3 years previously [99]; this finding has been recently confirmed by Cheung et al. [100] and also documented in children with PAN [101].

In contrast, another recent cross-sectional study by McCrindle failed to demonstrate differences in brachial artery reactivity or carotid intima-media thickness following KS, although there was some degree of impaired blood pressure control documented on 24-h ambulatory blood pressure monitoring in KD cases [102].

Perhaps more important than these studies of novel surrogate markers for vascular injury are the long-term epidemiological data from Japan. In the fifth look at long-term outcomes of a cohort of 6576 patients with KD

enrolled between 1982 and 1992, the mortality rate for patients without cardiac sequelae in the acute phase of the disease and female patients with sequelae did not differ from that of the normal population [103]. The mortality rate of males with cardiac sequelae was, however, 2.4-fold higher than that of the normal population. Thus, the long-term outlook for patients with coronary involvement due to KS, particularly males, must remain guarded at the present time.

## Conclusion and future directions

This is an exciting time for vasculitis research. Our understanding of the pathogenesis is advancing, as is our approach to novel therapeutic approaches, including the increasing use of biologics, although formal clinical trials involving children are still lacking in this area. Significant other challenges are looming. These include establishing international research networks and databases to obtain adequate patient numbers, thereby enabling powering of studies; validation of classification criteria suitable for the paediatric vasculitides [104, 105]; the development of tools to allow reliable non-invasive monitoring of disease activity; and the development of robust core outcome variables that can be used to assess outcomes in clinical trials. Lastly, there remains the question regarding longer term cardiovascular morbidity in children who survive vasculitis. Ultimately it must be anticipated that advances in our understanding of the environmental triggers and host responses will shape future novel therapeutic approaches to PSV in the young.

**Acknowledgments** I would like to thank Professor Michael Dillon for helpful comments on this manuscript.

## References

1. Brogan PA, Dillon MJ (2000) Vasculitis from the pediatric perspective. *Curr Rheumatol Rep* 2:411–416
2. Brogan PA, Shah V, Bagga A, Klein N, Dillon MJ (2003) T cell Vbeta repertoires in childhood vasculitides. *Clin Exp Immunol* 131:517–527
3. Brogan PA, Shah V, Klein N, Dillon MJ (2004) Vbeta-restricted T cell adherence to endothelial cells: a mechanism for superantigen-dependent vascular injury. *Arthritis Rheum* 50:589–597
4. Pagnoux C, Cohen P, Guillemin L (2006) Vasculitides secondary to infections. *Clin Exp Rheumatol* 24:S71–S81
5. Nades SJ (2002) Known causes of vasculitis in man. *Cleve Clin J Med* 69[Suppl 2]:S115–S119
6. Gardner-Medwin, JM, Dolezalova P, Cummins C, Southwood TR (2002) Incidence of Henoch-Schonlein purpura, Kawasaki disease, and rare vasculitides in children of different ethnic origins. *Lancet* 360:1197–1202



7. Yang YH, Hung CF, Hsu CR, Wang LC, Chuang YH, Lin YT, Chiang BL (2005) A nationwide survey on epidemiological characteristics of childhood Henoch-Schonlein purpura in Taiwan. *Rheumatology (Oxford)* 44:618–622
8. Dolezalova P, Telekesova P, Nemcova D, Hoza J (2004) Incidence of vasculitis in children in the Czech Republic: 2-year prospective epidemiology survey. *J Rheumatol* 31:2295–2299
9. Brogan PA, Bose A, Burgner D, Shingadia D, Tulloh R, Michie C, Klein N, Booy R, Levin M, Dillon MJ (2002) Kawasaki disease: an evidence based approach to diagnosis, treatment, and proposals for future research. *Arch Dis Child* 86:286–290
10. Newburger JW, Taubert KA, Shulman ST, Rowley AH, Gewitz MH, Takahashi M, McCrindle BW (2003) Summary and abstracts of the Seventh International Kawasaki Disease Symposium. *Hakone Jpn Pediatr Res* 53:153–157
11. Shulman ST, Rowley AH (2004) Advances in Kawasaki disease. *Eur J Pediatr* 163:285–291
12. Bronstein, DE, Dille AN, Austin JP, Williams CM, Palinkas LA, Burns JC (2000) Relationship of climate, ethnicity and socio-economic status to Kawasaki disease in San Diego County, 1994 through 1998. *Pediatr Infect Dis J* 19:1087–1091
13. Fujita Y, Nakamura Y, Sakata K, Hara N, Kobayashi M, Nagai M, Yanagawa H, Kawasaki T (1989) Kawasaki disease in families. *Pediatrics* 84:666–669
14. Harnden A, Alves B, Sheikh A (2002) Rising incidence of Kawasaki disease in England: analysis of hospital admission data. *BMJ* 324:1424–1425
15. Chang L, Chiang BL, Kao CL, Wu MH, Chen PJ, Berkhout B, Yang HC, Huang LM (2006) Lack of association between infection with a novel human coronavirus (HCoV), HCoV-NH, and Kawasaki disease in Taiwan. *J Infect Dis* 193:283–286
16. Kim S, Dedeoglu F (2005) Update on pediatric vasculitis. *Curr Opin Pediatr* 17:695–702
17. Al-Sheyyab M, Batiha A, el-Shanti H, Daoud A (1999) Henoch-Schonlein purpura and streptococcal infection: a prospective case-control study. *Ann Trop Paediatr* 19:253–255
18. Ayoub EM, McBride J, Schmiederer M, Anderson B (2002) Role of *Bartonella henselae* in the etiology of Henoch-Schonlein purpura. *Pediatr Infect Dis J* 21:28–31
19. Cioc AM, Sedmak DD, Nuovo GJ, Dawood MR, Smart G, Magro CM (2002) Parvovirus B19 associated adult Henoch Schonlein purpura. *J Cutan Pathol* 29:602–607
20. Hidaka H, Okada T, Matsumoto H, Yoshino M, Nagaoka Y, Takeguchi F, Iwasawa H, Tomaru R, Wada T, Shimizu T, Ohtani M, Yamanaka K, Fukutake K, Nakao T (2003) Henoch-Schonlein purpura nephritis in a patient infected with the human immunodeficiency virus. *Nippon Jinzo Gakkai Shi* 45:387–392
21. Kalman S, Ibrahim AH, Atay A (2005) Henoch-Schonlein purpura in a child following varicella. *J Trop Pediatr* 51:240–241
22. Watts RA, Gonzalez-Gay MA, Lane SE, Garcia-Porrúa C, Benthams G, Scott DG (2001) Geoepidemiology of systemic vasculitis: comparison of the incidence in two regions of Europe. *Ann Rheum Dis* 60:170–172
23. Watts RA, Lane SE, Scott DG, Koldingsnes W, Nossent H, Gonzalez-Gay MA, Garcia-Porrúa C, Benthams GA (2001) Epidemiology of vasculitis in Europe. *Ann Rheum Dis* 60:1156–1157
24. Mahr AD, Neogi T, Merkel PA (2006) Epidemiology of Wegener's granulomatosis: Lessons from descriptive studies and analyses of genetic and environmental risk determinants. *Clin Exp Rheumatol* 24:S82–S91
25. Moulin P, Lehucher-Michel MP (2004) Wegener's disease and exposure to silica. Study of the physiopathological mechanisms. *Presse Med* 33:1349–1351
26. Nuyts GD, Van Vlem E, De Vos A, Daelemans RA, Rorive G, Elseviers MM, Schurgers M, Segaert M, D'Haese PC, De Broe ME (1995) Wegener granulomatosis is associated to exposure to silicon compounds: a case-control study. *Nephrol Dial Transplant* 10:1162–1165
27. Rosenman KD, Moore-Fuller M, Reilly MJ (2000) Kidney disease and silicosis. *Nephron* 85:14–19
28. Stratta P, Messuerotti A, Canavese C, Coen M, Luccoli L, Bussolati B, Giorda L, Malavenda P, Cacciabue M, Bugiani M, Bo M, Ventura M, Camussi G, Fubini B (2001) The role of metals in autoimmune vasculitis: epidemiological and pathogenic study. *Sci Total Environ* 270:179–190
29. Haubitz M, Woywodt A, de Groot K, Haller H, Goebel U (2005) Smoking habits in patients diagnosed with ANCA associated small vessel vasculitis. *Ann Rheum Dis* 64:1500–1502
30. Hogan SL, Satterly KK, Dooley MA, Nachman PH, Jennette JC, Falk RJ (2001) Silica exposure in anti-neutrophil cytoplasmic autoantibody-associated glomerulonephritis and lupus nephritis. *J Am Soc Nephrol* 12:134–142
31. Lane SE, Watts RA, Benthams G, Innes NJ, Scott DG (2003) Are environmental factors important in primary systemic vasculitis? A case-control study. *Arthritis Rheum* 48:814–823
32. Merkel PA (1998) Drugs associated with vasculitis. *Curr Opin Rheumatol* 10:45–50
33. Pankhurst T, Savage CO, Gordon C, Harper L (2004) Malignancy is increased in ANCA-associated vasculitis. *Rheumatology* 43:1532–1535
34. Barron KS (2002) Kawasaki disease: etiology, pathogenesis, and treatment. *Cleve Clin J Med* 69[Suppl 2]:SII69–SII78
35. Li H, Llera A, Mariuzza RA (1998) Structure-function studies of T-cell receptor-superantigen interactions. *Immunol Rev* 163:177–186
36. Abe J, Kotzin BL, Jujo K, Melish ME, Glode MP, Kohsaka T, Leung DY (1992) Selective expansion of T cells expressing T-cell receptor variable regions V beta 2 and V beta 8 in Kawasaki disease. *Proc Natl Acad Sci USA* 89:4066–4070
37. Leung DY, Meissner HC, Shulman ST, Mason WH, Gerber MA, Glode MP, Myones BL, Wheeler JG, Ruthazer R, Schlievert PM (2002) Prevalence of superantigen-secreting bacteria in patients with Kawasaki disease. *J Pediatr* 140:742–746
38. Matsubara K, Fukaya T, Miwa K, Shibayama N, Nigami H, Harigaya H, Nozaki H, Hirata T, Baba K, Suzuki T, Ishiguro A (2006) Development of serum IgM antibodies against superantigens of *Staphylococcus aureus* and *Streptococcus pyogenes* in Kawasaki disease. *Clin Exp Immunol* 143:427–434
39. Leung DY, Giorno RC, Kazemi LV, Flynn PA, Busse JB (1995) Evidence for superantigen involvement in cardiovascular injury due to Kawasaki syndrome. *J Immunol* 155:5018–5021
40. Yamashiro Y, Nagata S, Oguchi S, Shimizu T (1996) Selective increase of V beta 2+ T cells in the small intestinal mucosa in Kawasaki disease. *Pediatr Res* 39:264–266
41. Rowley AH, Shulman ST, Spike BT, Mask CA, Baker SC (2001) Oligoclonal IgA response in the vascular wall in acute Kawasaki disease. *J Immunol* 166:1334–1343
42. Rowley AH, Baker SC, Shulman ST, Garcia FL, Guzman-Cottrill JA, Chou P, Terai M, Kawasaki T, Kalelkar MB, Crawford SE (2004) Detection of antigen in bronchial epithelium and macrophages in acute Kawasaki disease by use of synthetic antibody. *J Infect Dis* 190:856–865
43. Ozen S, Bakkaloglu A, Dusunel R, Soylemezoglu O, Ozaltin F, Poyrazoglu H, Kasapcopur O, Ozkaya O, Yalcinkaya F, Balat A, Kural N, Donmez O, Alpaya H, Anarat A, Mir S, Gur-Guven A, Sonmez F, Gok F (2007) Childhood vasculitides in Turkey: a nationwide survey. *Clin Rheumatol* 26:196–200
44. Pupa ER, Tervaert JW (2003) The relation between *Staphylococcus aureus* and Wegener's granulomatosis: current knowledge and future directions. *Intern Med* 42:771–780
45. Stegeman CA, Tervaert JW, Sluiter WJ, Manson WL, de Jong PE, Kallenberg CG (1994) Association of chronic nasal carriage

- of *Staphylococcus aureus* and higher relapse rates in Wegener granulomatosis. *Ann Intern Med* 120:12–17
46. Stegeman CA, Tervaert JW, de Jong PE, Kallenberg CG (1996) Trimethoprim-sulfamethoxazole (co-trimoxazole) for the prevention of relapses of Wegener's granulomatosis. Dutch Co-Trimoxazole Wegener Study Group. *N Engl J Med* 335:16–20
  47. Popa ER, Stegeman CA, Bos NA, Kallenberg CG, Tervaert JW (2003) Staphylococcal superantigens and T cell expansions in Wegener's granulomatosis. *Clin Exp Immunol* 132:496–504
  48. Mason JC, Cowie MR, Davies KA, Schofield JB, Cambridge J, Jackson J, So A, Allard SA, Walport MJ (1994) Familial polyarteritis nodosa. *Arthritis Rheum* 37:1249–1253
  49. Dergun M, Kao A, Hauger SB, Newburger JW, Burns JC (2005) Familial occurrence of Kawasaki syndrome in North America. *Arch Pediatr Adolesc Med* 159:876–881
  50. Manganelli P, Giacosa R, Fietta P, Zanetti A, Neri TM (2003) Familial vasculitides: Churg-Strauss syndrome and Wegener's granulomatosis in 2 first-degree relatives. *J Rheumatol* 30:618–621
  51. Kodama K, Kida O, Morotomi Y, Tanaka K (1986) Male siblings with Takayasu's arteritis suggest genetic etiology. *Heart Vessels* 2:51–54
  52. Biezeveld MH, Kuipers IM, Geissler J, Lam J, Ottenkamp JJ, Hack CE, Kuijpers TW (2003) Association of mannose-binding lectin genotype with cardiovascular abnormalities in Kawasaki disease. *Lancet* 361:1268–1270
  53. Biezeveld MH, Geissler J, Weverling GJ, Kuipers IM, Lam J, Ottenkamp J, Kuijpers TW (2006) Polymorphisms in the mannose-binding lectin gene as determinants of age-defined risk of coronary artery lesions in Kawasaki disease. *Arthritis Rheum* 54:369–376
  54. Turner MW, Hamvas RM (2000) Mannose-binding lectin: structure, function, genetics and disease associations. *Rev Immunogenet* 2:305–322
  55. Endo M, Ohi H, Ohsawa I, Fujita T, Matsushita M (2000) Complement activation through the lectin pathway in patients with Henoch-Schonlein purpura nephritis. *Am J Kidney Dis* 35:401–407
  56. Bansal PJ, Tobin MC (2004) Neonatal microscopic polyangiitis secondary to transfer of maternal myeloperoxidase-antineutrophil cytoplasmic antibody resulting in neonatal pulmonary hemorrhage and renal involvement. *Ann Allergy Asthma Immunol* 93:398–401
  57. Schlieben DJ, Korbet SM, Kimura RE, Schwartz MM, Lewis EJ (2005) Pulmonary-renal syndrome in a newborn with placental transmission of ANCA. *Am J Kidney Dis* 45:758–761
  58. Xiao H, Heeringa P, Hu P, Liu Z, Zhao M, Aratani Y, Maeda N, Falk RJ, Jennette JC (2002) Antineutrophil cytoplasmic autoantibodies specific for myeloperoxidase cause glomerulonephritis and vasculitis in mice. *J Clin Invest* 110:955–963
  59. Jennette JC, Xiao H, Falk RJ (2006) Pathogenesis of vascular inflammation by anti-neutrophil cytoplasmic antibodies. *J Am Soc Nephrol* 17:1235–1242
  60. Morgan MD, Harper L, Williams J, Savage C (2006) Antineutrophil cytoplasm-associated glomerulonephritis. *J Am Soc Nephrol* 17:1224–1234
  61. Harper L, Savage CO (2000) Pathogenesis of ANCA-associated systemic vasculitis. *J Pathol* 190:349–359
  62. Mulder AH, Stegeman CA, Kallenberg CG (1995) Activation of granulocytes by anti-neutrophil cytoplasmic antibodies (ANCA) in Wegener's granulomatosis: a predominant role for the IgG3 subclass of ANCA. *Clin Exp Immunol* 101:227–232
  63. Savage CO, Harper L, Holland M (2002) New findings in pathogenesis of antineutrophil cytoplasm antibody-associated vasculitis. *Curr Opin Rheumatol* 14:15–22
  64. Tipping PG, Holdsworth SR (2006) T cells in crescentic glomerulonephritis. *J Am Soc Nephrol* 17:1253–1263
  65. Harper L, Ren Y, Savill J, Adu D, Savage CO (2000) Antineutrophil cytoplasmic antibodies induce reactive oxygen-dependent dysregulation of primed neutrophil apoptosis and clearance by macrophages. *Am J Pathol* 157:211–220
  66. Harper L, Cockwell P, Adu D, Savage CO (2001) Neutrophil priming and apoptosis in anti-neutrophil cytoplasmic autoantibody-associated vasculitis. *Kidney Int* 59:1729–1738
  67. Zwaal RF, Schroit AJ (1997) Pathophysiologic implications of membrane phospholipid asymmetry in blood cells. *Blood* 89:1121–1132
  68. Pendergraft WF III, Preston GA, Shah RR, Tropsha A, Carter CW Jr, Jennette JC, Falk RJ (2004) Autoimmunity is triggered by cPR-3(105–201), a protein complementary to human autoantigen proteinase-3. *Nat Med* 10:72–79
  69. Pendergraft WF III, Pressler BM, Jennette JC, Falk RJ, Preston GA (2005) Autoantigen complementarity: a new theory implicating complementary proteins as initiators of autoimmune disease. *J Mol Med* 83:12–25
  70. Senzaki H (2006) The pathophysiology of coronary artery aneurysms in Kawasaki disease: role of matrix metalloproteinases. *Arch Dis Child* 91:847–851
  71. Senzaki H, Masutani S, Kobayashi J, Kobayashi T, Nakano H, Nagasaka H, Sasaki N, Asano H, Kyo S, Yokote Y (2001) Circulating matrix metalloproteinases and their inhibitors in patients with Kawasaki disease. *Circulation* 104:860–863
  72. Hirono K, Foell D, Xing Y, Miyagawa-Tomita S, Ye F, Ahlmann M, Vogl T, Futatani T, Rui C, Yu X, Watanabe K, Wanatabe S, Tsubata S, Uese K, Hashimoto I, Ichida F, Nakazawa M, JRoth J, Miyawaki T (2006) Expression of myeloid-related protein-8 and -14 in patients with acute Kawasaki disease. *J Am Coll Cardiol* 48:1257–1264
  73. Booth AL, Harper L, Hammad T, Bacon P, Griffith M, Levy J, Savage C, Pusey C, Jayne D (2004) Prospective study of TNFalpha blockade with infliximab in anti-neutrophil cytoplasmic antibody-associated systemic vasculitis. *Am Soc Nephrol* 15:717–721
  74. Booth AD, Jayne DR, Kharbanda RK, McEniery CM, Mackenzie IS, Brown J, Wilkinson IB (2004) Infliximab improves endothelial dysfunction in systemic vasculitis: a model of vascular inflammation. *Circulation* 109:1718–1723
  75. Stenbog EV, Windelborg B, Horlyck A, Herlin T (2006) The effect of TNFalpha blockade in complicated, refractory Kawasaki disease. *Scand J Rheumatol* 35:318–321
  76. Wilkinson NM, Erendzhinova E, Zeff A, Cabral DA (2006) Infliximab as rescue therapy in three cases of paediatric Wegener's granulomatosis. *Rheumatology (Oxford)* 45:1047–1048
  77. Brogan PA, Shah V, Brachet C, Harnden A, Mant D, Klein N, Dillon MJ (2004) Endothelial and platelet microparticles in vasculitis of the young. *Arthritis Rheum* 50:927–936
  78. Bernal-Mizrachi L, Jy W, Fierro C, Macdonough R, Velazquez HA, Purow J, Jimenez JJ, Horstman LL, Ferreira A, de Marchena E, Ahn YS (2004) Endothelial microparticles correlate with high-risk angiographic lesions in acute coronary syndromes. *Int J Cardiol* 97:439–446
  79. Jimenez JJ, Jy W, Mauro LM, Horstman LL, Ahn YS (2001) Elevated endothelial microparticles in thrombotic thrombocytopenic purpura: findings from brain and renal microvascular cell culture and patients with active disease. *Br J Haematol* 112:81–90
  80. Brogan PA, Dillon MJ (2004) Endothelial microparticles and the diagnosis of the vasculitides. *Intern Med* 43:1115–1119
  81. Woywodt A, Bahlmann FH, de Groot K, Haller H, Haubitz M (2002) Circulating endothelial cells: life, death, detachment and

- repair of the endothelial cell layer. *Nephrol Dial Transplant* 17:1728–1730
82. Woywodt A, Streiber F, de Groot K, Regelsberger H, Haller H, Haubitz M (2003) Circulating endothelial cells as markers for ANCA-associated small-vessel vasculitis. *Lancet* 361:206–210
  83. Woywodt A, Blann AD, Kirsch T, Erdbruegger U, Banzet N, Haubitz M, Dignat-George F (2006) Isolation and enumeration of circulating endothelial cells by immunomagnetic isolation: proposal of a definition and a consensus protocol. *J Thromb Haemost* 4:671–677
  84. Woywodt A, Goldberg C, Kirsch T, de Groot K, Erdbruegger U, Haller H, Haubitz M (2006) Circulating endothelial cells in relapse and limited granulomatous disease due to ANCA associated vasculitis. *Ann Rheum Dis* 65:164–168
  85. Nadar SK, Lip GY, Lee KW, Blann AD (2005) Circulating endothelial cells in acute ischaemic stroke. *Thromb Haemost* 94:707–712
  86. Bull TM, Golpon H, Hebbel RP, Solovey A, Cool CD, Tuder RM, Geraci MW, Voelkel NF (2003) Circulating endothelial cells in pulmonary hypertension. *Thromb Haemost* 90:698–703
  87. Hill JM, Zalos G, Halcox JP, Schenke WH, Waclawiw MA, Quyyumi AA, Finkel T (2003) Circulating endothelial progenitor cells, vascular function, and cardiovascular risk. *N Engl J Med* 348:593–600
  88. Asahara T, Murohara, T Sullivan A, Silver M, van der Zee R, Li T, Witzenbichler B, Schatteman G, Isner JM (1997) Isolation of putative progenitor endothelial cells for angiogenesis. *Science* 275:964–967
  89. Werner N, Kosiol S, Schiegl T, Ahlers P, Walenta K, Link A, Bohm M, Nickenig G (2005) Circulating endothelial progenitor cells and cardiovascular outcomes. *N Engl J Med* 353:999–1007
  90. Herbrig K, Haensel S, Oelschlaegel U, Pistrosch F, Foerster S, Passauer J (2006) Endothelial dysfunction in patients with rheumatoid arthritis is associated with a reduced number and impaired function of endothelial progenitor cells. *Ann Rheum Dis* 65:157–163
  91. Kuwana M, Okazaki Y, Yasuoka H, Kawakami Y, Ikeda Y (2004) Defective vasculogenesis in systemic sclerosis. *Lancet* 364:603–610
  92. Holmen C, Elsheikh E, Stenvinkel P, Qureshi AR, Pettersson E, Jalkanen S, Sumitran-Holgersson S (2005) Circulating inflammatory endothelial cells contribute to endothelial progenitor cell dysfunction in patients with vasculitis and kidney involvement. *J Am Soc Nephrol* 16:3110–3120
  93. Nakatani K, Takeshita S, Tsujimoto H, Kawamura Y, Tokutomi T, Sekine I (2003) Circulating endothelial cells in Kawasaki disease. *Clin Exp Immunol* 131:536–540
  94. Jin H, Aiyer A, Su J, Borgstrom P, Stupack D, Friedlander M, Varner J (2006) A homing mechanism for bone marrow-derived progenitor cell recruitment to the neovasculature. *J Clin Invest* 116:652–662
  95. Kogata N, Arai Y, Pearson JT, Hashimoto K, Hidaka K, Koyama T, Somekawa S, Nakaoka Y, Ogawa M, Adams RH, Okada M, Mochizuki N (2006) Cardiac ischemia activates vascular endothelial cadherin promoter in both preexisting vascular cells and bone marrow cells involved in neovascularization. *Circ Res* 98:897–904
  96. Naoe S, Takahashi K, Masuda H, Tanaka N (1991) Kawasaki disease. With particular emphasis on arterial lesions. *Acta Pathol Jpn* 41:785–797
  97. Dhillion R, Clarkson P, Donald AE, Powe AJ, Nash M, Novelli V, Dillon MJ, Deanfield JE (1996) Endothelial dysfunction late after Kawasaki disease. *Circulation* 94:2103–2106
  98. Albisetti M, Chan AK, McCrindle BW, Wong D, Vegh P, Adams M, Dinyari M, Monagle P, Andrew M (2003) Fibrinolytic response to venous occlusion is decreased in patients after Kawasaki disease. *Blood Coagul Fibrin* 14:181–186
  99. Pilla C, Cheung YF, Brogan PA, Dillon MJ, Redington AN (2000) Chronically reduced arterial distensibility in kawasaki disease: further evidence for the beneficial effects of immunoglobulin (Abstract). *Circulation* 102[Supplement 2]:830–831
  100. Cheung YF, Ho MH, Tam SC, Yung TC (2004) Increased high sensitivity C reactive protein concentrations and increased arterial stiffness in children with a history of Kawasaki disease. *Heart* 90:1281–1285
  101. Cheung YF, Brogan PA, Pilla CB, Dillon MJ, Redington AN (2002) Arterial distensibility in children and teenagers: normal evolution and the effect of childhood vasculitis. *Arch Dis Child* 87:348–351
  102. Silva AA, Maeno Y, Hashmi A, Smallhorn JF, Silverman ED, McCrindle BW (2001) Cardiovascular risk factors after Kawasaki disease: a case-control study. *J Pediatr* 138:400–405
  103. Nakamura Y, Yanagawa H, Harada K, Kato H, Kawasaki T (2002) Mortality among persons with a history of Kawasaki disease in Japan: the fifth look. *Arch Pediatr Adolesc Med* 156:162–165
  104. Dillon MJ, Ozen S (2006) A new international classification of childhood vasculitis. *Pediatr Nephrol* 21:1219–1222
  105. Ozen S, Ruperto N, Dillon MJ, Bagga A, Barron K, Davin JC, Kawasaki T, Lindsley C, Petty RE, Prieur AM, Ravelli A, Woo P (2006) EULAR/PRES endorsed consensus criteria for the classification of childhood vasculitides. *Ann Rheum Dis* 65:936–941
  106. Shim YH, Kim HS, Sohn S, Hong YM (2006) Insertion/deletion polymorphism of angiotensin converting enzyme gene in Kawasaki disease. *J Korean Med Sci* 21:208–211
  107. Dudley J, Afifi E, Gardner A, Tizard EJ, McGraw ME (2000) Polymorphism of the ACE gene in Henoch-Schonlein purpura nephritis. *Pediatr Nephrol* 14:218–220
  108. Brodkiewicz A, Ciechanowicz A, Urbanska A, Peregud-Pogorzelski J, Dzienski P, Subicka D, Fydryk J (2000) The I/D polymorphism of the ACE gene in children with Henoch-Schoenlein purpura. *Pol Merk Lekarski* 8:236–238
  109. Amoroso A, Danek G, Vatta S, Crovella S, Berrino M, Guarrera S, Fasano ME, Mazzola G, Amore A, Gianoglio B, Peruzzi L, Coppo R (1998) Polymorphisms in angiotensin-converting enzyme gene and severity of renal disease in Henoch-Schoenlein patients. Italian Group of Renal Immunopathology. *Nephrol Dial Transplant* 13:3184–3188
  110. Park JA, Shin KS, Kim YW (2005) Polymorphism of matrix metalloproteinase-3 promoter gene as a risk factor for coronary artery lesions in Kawasaki disease. *J Korean Med Sci* 20:607–611
  111. Zou CC, Zhao ZY, Tang LF, Liang L (2006) Plasma levels of matrix metalloproteinase-9 in Henoch-Schonlein purpura. *Scand J Rheumatol* 35:52–55
  112. Sanders JS, van Goor H, Hanemaaijer R, Kallenberg CG, Stegeman CA (2004) Renal expression of matrix metalloproteinases in human ANCA-associated glomerulonephritis. *Nephrol Dial Transplant* 19:1412–1419
  113. Kariyazono H, Ohno T, Khajoev V, Ihara K, Kusuhara K, Kinukawa N, Mizuno Y, Hara T (2004) Association of vascular endothelial growth factor (VEGF) and VEGF receptor gene polymorphisms with coronary artery lesions of Kawasaki disease. *Pediatr Res* 56:953–959
  114. Rueda B, Perez-Armengol C, Lopez-Lopez S, Garcia-Porrúa C, Martín J, Gonzalez-Gay MA (2006) Association between functional haplotypes of vascular endothelial growth factor and renal complications in Henoch-Schonlein purpura. *J Rheumatol* 33:69–73
  115. Li CG, Reynolds I, Ponting JM, Holt PJ, Hillarby MC, Kumar S (1998) Serum levels of vascular endothelial growth factor

- (VEGF) are markedly elevated in patients with Wegener's granulomatosis. *Br J Rheumatol* 37:1303–1306
116. Wu SF, Chang JS, Wan L, Tsai CH, Tsai FJ (2005) Association of IL-1Ra gene polymorphism, but no association of IL-1beta and IL-4 gene polymorphisms, with Kawasaki disease. *J Clin Lab Anal* 19:99–102
  117. Amoli MM, Thomson W, Hajeer AH, Calvino MC, Garcia-Porrúa C, Ollier WE, Gonzalez-Gay MA (2002) Interleukin 1 receptor antagonist gene polymorphism is associated with severe renal involvement and renal sequelae in Henoch-Schonlein purpura. *J Rheumatol* 29:1404–1407
  118. Amoli MM, Calvino MC, Garcia-Porrúa C, Llorca J, Ollier WE, Gonzalez-Gay MA (2004) Interleukin 1beta gene polymorphism association with severe renal manifestations and renal sequelae in Henoch-Schonlein purpura. *J Rheumatol* 31:295–298
  119. Huang D, Giscombe R, Zhou Y, Lefvert AK (2000) Polymorphisms in CTLA-4 but not tumor necrosis factor-alpha or interleukin 1beta genes are associated with Wegener's granulomatosis. *J Rheumatol* 27:397–401
  120. Yang J, Li CR, Li YB, Li RX, Sun LB, Huang HJ, Wang GB (2003) The correlation between Kawasaki disease and polymorphisms of Tumor necrosis factor alpha and interleukin-10 gene promoter. *Zhonghua Er Ke Za Zhi* 41:598–602
  121. Yang YH, Lai HJ, Kao CK, Lin YT, Chiang BL (2004) The association between transforming growth factor-beta gene promoter C-509T polymorphism and Chinese children with Henoch-Schonlein purpura. *Pediatr Nephrol* 19:972–975
  122. Amoli MM, Thomson W, Hajeer AH, Calvino MC, Garcia-Porrúa C, Ollier WE, Gonzalez-Gay MA (2002) Interleukin 8 gene polymorphism is associated with increased risk of nephritis in cutaneous vasculitis. *J Rheumatol* 29:2367–2370
  123. Bartfai Z, Gaede KI, Russell KA, Murakozy G, Muller-Quernheim J, Specks U (2003) Different gender-associated genotype risks of Wegener's granulomatosis and microscopic polyangiitis. *Clin Immunol* 109:330–337
  124. Burns JC, Shimizu C, Gonzalez E, Kulkarni H, Patel S, Shike H, Sundel RS, Newburger JW, Ahuja SK (2005) Genetic variations in the receptor-ligand pair CCR5 and CCL3L1 are important determinants of susceptibility to Kawasaki disease. *J Infect Dis* 192:344–349
  125. Gershoni-Baruch R, Broza Y, Brik R (2003) Prevalence and significance of mutations in the familial Mediterranean fever gene in Henoch-Schonlein purpura. *J Pediatr* 143:658–661
  126. Tunca M, Akar S, Onen F, Ozdogan H, Kasapcopur O, Yalcinkaya F, Tutar E, Ozen S, Topaloglu R, Yilmaz E, Arici M, Bakkaloglu A, Besbas N, Akpolat T, Dinc A, Erken E (2005) Familial Mediterranean fever (FMF) in Turkey: results of a nationwide multicenter study. *Medicine* 84:1–11
  127. Amoli MM, Thomson W, Hajeer AH, Calvino MC, Garcia-Porrúa C, Ollier WE, Gonzalez-Gay MA (2002) HLA-B35 association with nephritis in Henoch-Schonlein purpura. *J Rheumatol* 29:948–949
  128. Yi ZW, Fang XL, Wu XC, He XJ, He QN, Dang XQ, Zhu CP, Mo SH (2006) Role of PAX2 gene polymorphisms in Henoch-Schonlein purpura nephritis. *Nephrology* 11:42–48
  129. Khajooe V, Kariyazono H, Ohno T, Ihara K, Mizuno Y, Kusuhara K, Hara T (2003) Inducible and endothelial constitutive nitric oxide synthase gene polymorphisms in Kawasaki disease. *Pediatr Int* 45:130–134
  130. Martin J, Paco L, Ruiz MP, Lopez-Nevot MA, Garcia-Porrúa C, Amoli MM, Calvino MC, Ollier WE, Gonzalez-Gay MA (2005) Inducible nitric oxide synthase polymorphism is associated with susceptibility to Henoch-Schonlein purpura in northwestern Spain. *J Rheumatol* 32:1081–1085
  131. Amoli MM, Matthey DL, Calvino MC, Garcia-Porrúa C, Thomson W, Hajeer AH, Ollier WE, Gonzalez-Gay MA (2001) Polymorphism at codon 469 of the intercellular adhesion molecule-1 locus is associated with protection against severe gastrointestinal complications in Henoch-Schonlein purpura. *J Rheumatol* 28:1014–1018
  132. Gencik M, Meller S, Borgmann S, Sitter T, Menezes Saecker AM, Fricke H, Epplen JT (2000) The association of CD18 alleles with anti-myeloperoxidase subtypes of ANCA-associated systemic vasculitides. *Clin Immunol* 94:9–12
  133. Patterson CC, Ross P Jr, Pope-Harman AL, Knight DA, Magro CM (2005) Alpha-1 anti-trypsin deficiency and Henoch-Schonlein purpura associated with anti-neutrophil cytoplasmic and anti-endothelial cell antibodies of immunoglobulin-A isotype. *J Cutan Pathol* 32:300–306
  134. Esnault VL, Testa A, Audrain M, Roge C, Hamidou M, Barrier JH, Sesboue R, Martin JP, Lesavre P (1993) Alpha 1-antitrypsin genetic polymorphism in ANCA-positive systemic vasculitis. *Kidney Int* 43:1329–1332
  135. Esnault VL, Audrain MA, Sesboue R (1997) Alpha-1-antitrypsin phenotyping in ANCA-associated diseases: one of several arguments for protease/antiprotease imbalance in systemic vasculitis. *Exp Clin Immunogenet* 14:206–213
  136. Audrain MA, Sesboue R, Baranger TA, Elliott J, Testa A, Martin JP, Lockwood CM, Esnault VL (2001) Analysis of anti-neutrophil cytoplasmic antibodies (ANCA): frequency and specificity in a sample of 191 homozygous (PiZZ) alpha1-antitrypsin-deficient subjects. *Nephrol Dial Transplant* 16:39–44
  137. Segelmark M, Elzouki AN, Wieslander J, Eriksson S (1995) The PiZ gene of alpha 1-antitrypsin as a determinant of outcome in PR3-ANCA-positive vasculitis. *Kidney Int* 48:844–850
  138. Gencik M, Meller S, Borgmann S, Fricke H (2000) Proteinase 3 gene polymorphisms and Wegener's granulomatosis. *Kidney Int* 58:2473–2477
  139. Biezeveld M, Geissler J, Merkus M, Kuipers IM, Ottenkamp J, Kuijpers T (2007) The involvement of Fc gamma receptor gene polymorphisms in Kawasaki disease. *Clin Exp Immunol* 147:106–111
  140. Tse WY, Abadeh S, Jefferis R, Savage CO, Adu D (2000) Neutrophil FcgammaRIIb allelic polymorphism in anti-neutrophil cytoplasmic antibody (ANCA)-positive systemic vasculitis. *Clin Exp Immunol* 119:574–577
  141. Dijkstra-Hoogkampoorer HM, Scheepers RH, Oost WW, Stegeman CA, van der Pol WL, Sluiter WJ, Kallenberg CG, van de Winkel JG, Tervaert JW (1999) Fc gamma receptor polymorphisms in Wegener's granulomatosis: risk factors for disease relapse. *Arthritis Rheum* 42:1823–1827