INVITED EDITORIAL

'Going out on a limb': SDF-1a/CXCR4 signaling as a mechanism of remote ischemic preconditioning?

Karin Przyklenk

Received: 23 August 2013/Accepted: 23 August 2013/Published online: 4 September 2013 © Springer-Verlag Berlin Heidelberg 2013

Communication: the cornerstone of RIPC

Remote ischemic preconditioning (RIPC) is the intriguing phenomenon whereby brief, non-lethal episodes of ischemia in one organ or vascular bed render remote tissue resistant to a subsequent, sustained period of ischemia [23, 32]. While initially regarded as a laboratory curiosity [24], interest in RIPC was piqued by the observation that limb ischemia, achieved noninvasively by simple inflation of a blood pressure cuff, significantly reduced myocardial infarct size in the acute swine model of coronary artery occlusion-reperfusion [17]. In the ensuing years, since these first reports, progress has been made in defining the characteristics of RIPC-induced cardioprotection (including as-yet limited insights into cellular mechanisms), expansion of the concept beyond heart to encompass protection of other organs (including brain, kidney, liver and mesentery), and the investigation of RIPC in Phase II and III trials seeking to establish clinical efficacy in patients undergoing cardiac surgery or percutaneous intervention [2, 3, 6, 10, 14, 19, 20, 22, 24, 25, 31]. However, resolution of the distinguishing feature of RIPC has remained elusive [22, 24, 25]: how is the protective stimulus transferred or

This comment refers to the article available at doi:10.1007/s00395-013-0377-6.

K. Przyklenk (🖂)

Cardiovascular Research Institute, Wayne State University School of Medicine, Elliman Building, Room 1107, 421 E Canfield, Detroit, MI 48201, USA e-mail: kprzykle@med.wayne.edu

K. Przyklenk

communicated from the site of the RIPC stimulus to the heart?

Among the theories that have been proposed, considerable attention has focused on the concept that: (i) brief ischemia-reperfusion at the remote site triggers the release of one or more protective humoral factors, either directly or as a secondary consequence of neuronal stimulation; and (ii) the humoral factor(s) are then conveyed via the circulation to the myocardium [8, 22–25, 32]. Despite the emerging consensus that the factor(s) of interest are small (~3.5–15 kDa), presumably peptide(s), and hydrophobic [7, 28, 29], precise identification of the endogenous peptides transferred to the heart and capable of conferring cardioprotection has been problematic.

SDF-1a: the sought-after humoral protective factor?

In a recent issue of Basic Research in Cardiology, Davidson and colleagues posit that the circulating humoral peptide underlying the infarct-sparing effect of RIPC may be stromal cell derived factor (SDF)-1 α , and that SDF-1 α may activate cardioprotective signaling pathways by binding to its receptor, CXC chemokine receptor 4 (CXCR4) in heart [5]. To develop and test this novel hypothesis, a rat model was used in which the RIPC stimulus (three 5-min episodes of ischemia) was administered in vivo by tightening a tourniquet on one hindlimb; the heart was then excised and buffer-perfused, and, after 40 min of stabilization, a sustained, 35-min period of coronary artery occlusion was applied in vitro. Support for the authors' hypothesis was provided by three key pieces of evidence: (i) a significant 50 % increase in the plasma concentration of SDF-1a, assessed from samples collected immediately after RIPC, when compared with

Departments of Physiology and Emergency Medicine, Wayne State University School of Medicine, Detroit, MI, USA

time-matched sham-controls; (ii) documentation of CXCR4 protein expression in rat heart homogenates and isolated adult rat cardiomyocytes, and, of particular importance (iii) inhibition of RIPC-induced reduction of infarct size in rats that received AMD3100, the canonical CXCR4 antagonist, 10 min before imposing the first brief episode of hindlimb ischemia [5].

Strengths and limitations of the hypothesis

SDF-1 α contributes to the trafficking, homing and tissue retention of progenitor cells, and has garnered interest as a potential therapeutic strategy to enhance the efficacy of stem cell-based cardiac regenerative therapies [1, 15, 18, 27, 33]. The concept that this chemokine may have an asyet unappreciated role as the circulating, protective peptide that triggers infarct size reduction by RIPC is, for many reasons, logical and appealing. For example, SDF1- α is a small (10 kDa) molecule that displays an increase in expression under conditions of hypoxia and ischemia [4, 15]. In addition, there is evidence that SDF-1 α has a direct, cardioprotective effect when administered before coronary artery occlusion-reperfusion or permanent coronary artery ligation [15, 26], reportedly mediated via SDF-1a/CXCR4 binding and up-regulation of classic 'survival' kinases (including components of the Reperfusion Injury Salvage Kinase [RISK] and Survivor Activating Factor Enhancement [SAFE] pathways) involved in myocardial pre-, postand remote conditioning [11, 12, 15, 22, 24–26].

However, to definitively establish that release of SDF- 1α from the ischemic limb and subsequent SDF- 1α / CXCR4 binding in heart plays a mechanistic role in RIPC, compelling evidence of cause-and-effect is required. In this regard, the pivotal experiment, administration of AMD3100 in an effort to block infarct size reduction with RIPC, yielded intermediate results: infarct size averaged 53 ± 3 , 27 ± 3 and 40 ± 4 % of the at-risk myocardium in control, RIPC and AMD3100-pretreated RIPC groups, respectively [5]. Indeed, if the sample sizes (and thus the statistical power) were increased from the current value of n = 6 to n = 8 per group with no change in variance, the intermediate infarct size in the AMD3100-treated RIPC cohort would differ significantly from controls at the p < 0.05 level.

This partial inhibition achieved with AMD3100 may, as discussed by Davidson and colleagues, reflect the involvement of multiple circulating factors in RIPC-induced cardioprotection [5]. Alternatively, as only one dose of AMD3100 was evaluated, the outcome may also be explained by a suboptimal dose of the antagonist. There is an additional and potentially confounding issue that also warrants consideration: neither SDF-1 α nor AMD3100

bind exclusively to CXCR4. SDF-1 α is a ligand for both CXCR4 and CXCR7 [30], while AMD3100 also binds to and is an *agonist* (rather than antagonist) for—CXCR7 [16]. There is evidence that CXCR7 is expressed in heart [9, 30], but its potential contribution to cardioprotection is unexplored.

Future directions

In addition to resolution of the aforementioned uncertainties and limitations regarding selectivity that plague all studies using pharmacologic antagonists, definitive conclusions regarding the involvement of the SDF-1 α / CXCR4 axis in RIPC will require confirmation in multiple models and species, including more 'standard' in vivo protocols with no sustained delay (as in the Davidson study [5]) between the RIPC stimulus and the onset of myocardial ischemia. Of particular importance, clinical evidence of increased plasma concentrations of SDF-1a following brief limb ischemia will be required. Interestingly, in a recent, comprehensive proteomic analysis of human plasma samples, SDF-1 α was not among the candidates identified as being up-regulated after an RIPC stimulus [13]. Finally, future studies—and future attempts to exploit SDF-1 α as either an 'RIPC-mimetic' or an assay to guide in the optimization of RIPC-must take into consideration the apparent complexities of SDF-1 α / CXCR4 signaling, including reports that stimulation of chemokine receptors may up-regulate both pro-survival and pro-apoptotic signaling [18, 21]. Identification of SDF-1 α as a protective humoral factor has the potential to represent a breakthrough in our understanding of RIPC, but much work remains before we can conclude with certainty that the SDF-1a/CXCR4 axis plays a mechanistic role in the cardioprotection conferred by remote ischemia.

References

- Abbott JD, Huang Y, Liu D, Hickey R, Krause DS, Giordano FJ (2004) Stromal cell-derived factor-lalpha plays a critical role in stem cell recruitment to the heart after myocardial infarction but is not sufficient to induce homing in the absence of injury. Circulation 110:3300–3305. doi:10.1161/01.CIR.0000147780.30124.CF
- Botker HE, Kharbanda R, Schmidt MR, Bottcher M, Kaltoft AK, Terkelsen CJ, Munk K, Andersen NH, Hansen TM, Trautner S, Lassen JF, Christiansen EH, Krusell LR, Kristensen SD, Thuesen L, Nielsen SS, Rehling M, Sorensen HT, Redington AN, Nielsen TT (2010) Remote ischaemic conditioning before hospital admission, as a complement to angioplasty, and effect on myocardial salvage in patients with acute myocardial infarction: a randomised trial. Lancet 375:727–734. doi:10.1016/S0140-6736(09)62001-8

- Candilio L, Malik A, Hausenloy DJ (2013) Protection of organs other than the heart by remote ischemic conditioning. J Cardiovasc Med (Hagerstown) 14:193–205. doi:10.2459/JCM.0b013e 328359dd7b
- Ceradini DJ, Kulkarni AR, Callaghan MJ, Tepper OM, Bastidas N, Kleinman ME, Capla JM, Galiano RD, Levine JP, Gurtner GC (2004) Progenitor cell trafficking is regulated by hypoxic gradients through HIF-1 induction of SDF-1. Nat Med 10:858–864. doi:10.1038/nm1075
- Davidson SM, Selvaraj P, He D, Boi-Doku C, Yellon RL, Vicencio JM, Yellon DM (2013) Remote ischaemic preconditioning involves signalling through the SDF-1alpha/CXCR4 signalling axis. Basic Res Cardiol 108:377. doi:10.1007/s00395-013-0377-6
- Davies WR, Brown AJ, Watson W, McCormick LM, West NE, Dutka DP, Hoole SP (2013) Remote ischemic preconditioning improves outcome at 6 years after elective percutaneous coronary intervention: the CRISP stent trial long-term follow-up. Circ Cardiovasc Interv 6:246–251. doi:10.1161/CIRCINTERVEN TIONS.112.000184
- Dickson EW, Blehar DJ, Carraway RE, Heard SO, Steinberg G, Przyklenk K (2001) Naloxone blocks transferred preconditioning in isolated rabbit hearts. J Mol Cell Cardiol 33:1751–1756. doi:10.1006/jmcc.2001.1436
- Dickson EW, Lorbar M, Porcaro WA, Fenton RA, Reinhardt CP, Gysembergh A, Przyklenk K (1999) Rabbit heart can be "preconditioned" via transfer of coronary effluent. Am J Physiol 277:H2451–H2457 n/a
- Gerrits H, van Ingen Schenau DS, Bakker NE, van Disseldorp AJ, Strik A, Hermens LS, Koenen TB, Krajnc-Franken MA, Gossen JA (2008) Early postnatal lethality and cardiovascular defects in CXCR7-deficient mice. Genesis 46:235–245. doi:10.1002/dvg. 20387
- Hausenloy DJ, Candilio L, Laing C, Kunst G, Pepper J, Kolvekar S, Evans R, Robertson S, Knight R, Ariti C, Clayton T, Yellon DM, Investigators ET (2012) Effect of remote ischemic preconditioning on clinical outcomes in patients undergoing coronary artery bypass graft surgery (ERICCA): rationale and study design of a multi-centre randomized double-blinded controlled clinical trial. Clin Res Cardiol 101:339–348. doi:10.1007/s00392-011-0397-x
- Hausenloy DJ, Lecour S, Yellon DM (2011) Reperfusion injury salvage kinase and survivor activating factor enhancement prosurvival signaling pathways in ischemic postconditioning: two sides of the same coin. Antioxid Redox Signal 14:893–907. doi:10.1089/ars.2010.3360
- Hausenloy DJ, Yellon DM (2004) New directions for protecting the heart against ischaemia-reperfusion injury: targeting the Reperfusion Injury Salvage Kinase (RISK)-pathway. Cardiovasc Res 61:448–460. doi:10.1016/j.cardiores.2003.09.024
- Hepponstall M, Ignjatovic V, Binos S, Monagle P, Jones B, Cheung MH, d'Udekem Y, Konstantinov IE (2012) Remote ischemic preconditioning (RIPC) modifies plasma proteome in humans. PLoS One 7:e48284. doi:10.1371/journal.pone.0048284
- Heusch G (2013) Cardioprotection: chances and challenges of its translation to the clinic. Lancet 381:166–175. doi:10.1016/S0140-6736(12)60916-7
- Hu X, Dai S, Wu WJ, Tan W, Zhu X, Mu J, Guo Y, Bolli R, Rokosh G (2007) Stromal cell derived factor-1 alpha confers protection against myocardial ischemia/reperfusion injury: role of the cardiac stromal cell derived factor-1 alpha CXCR4 axis. Circulation 116:654–663. doi:10.1161/CIRCULATIONAHA. 106.672451
- Kalatskaya I, Berchiche YA, Gravel S, Limberg BJ, Rosenbaum JS, Heveker N (2009) AMD3100 is a CXCR7 ligand with allosteric agonist properties. Mol Pharmacol 75:1240–1247. doi:10. 1124/mol.108.053389

- Kharbanda RK, Mortensen UM, White PA, Kristiansen SB, Schmidt MR, Hoschtitzky JA, Vogel M, Sorensen K, Redington AN, MacAllister R (2002) Transient limb ischemia induces remote ischemic preconditioning in vivo. Circulation 106:2881–2883. doi:10.1161/01.CIR.0000043806.51912.9B
- Liehn EA, Tuchscheerer N, Kanzler I, Drechsler M, Fraemohs L, Schuh A, Koenen RR, Zander S, Soehnlein O, Hristov M, Grigorescu G, Urs AO, Leabu M, Bucur I, Merx MW, Zernecke A, Ehling J, Gremse F, Lammers T, Kiessling F, Bernhagen J, Schober A, Weber C (2011) Double-edged role of the CXCL12/ CXCR4 axis in experimental myocardial infarction. J Am Coll Cardiol 58:2415–2423. doi:10.1016/j.jacc.2011.08.033
- 19. Meybohm P, Zacharowski K, Cremer J, Roesner J, Kletzin F, Schaelte G, Felzen M, Strouhal U, Reyher C, Heringlake M, Schon J, Brandes I, Bauer M, Knuefermann P, Wittmann M, Hachenberg T, Schilling T, Smul T, Maisch S, Sander M, Moormann T, Boening A, Weigand MA, Laufenberg R, Werner C, Winterhalter M, Treschan T, Stehr SN, Reinhart K, Hasenclever D, Brosteanu O, Bein B (2012) Remote ischaemic preconditioning for heart surgery. The study design for a multicenter randomized double-blinded controlled clinical trial-the RIPHeart-Study. Eur Heart J 33:1423–1426 n/a
- Ovize M, Thibault H, Przyklenk K (2013) Myocardial conditioning: opportunities for clinical translation. Circ Res 113:439–450. doi:10.1161/CIRCRESAHA.113.300764
- Proulx C, El-Helou V, Gosselin H, Clement R, Gillis MA, Villeneuve L, Calderone A (2007) Antagonism of stromal cellderived factor-1alpha reduces infarct size and improves ventricular function after myocardial infarction. Pflugers Arch 455:241–250. doi:10.1007/s00424-007-0284-5
- Przyklenk K (2013) Reduction of myocardial infarct size with ischemic "conditioning": physiologic and technical considerations. Anesth Analg doi. doi:10.1213/ANE.0b013e318294fc63
- Przyklenk K, Bauer B, Ovize M, Kloner RA, Whittaker P (1993) Regional ischemic 'preconditioning' protects remote virgin myocardium from subsequent sustained coronary occlusion. Circulation 87:893–899. doi:10.1161/01.CIR.87.3.893
- Przyklenk K, Whittaker P (2013) Genesis of remote conditioning: action at a distance–'hypotheses non fingo'? J Cardiovasc Med (Hagerstown) 14:180–186. doi:10.2459/JCM.0b013e328358c8eb
- Przyklenk K, Whittaker P (2011) Remote ischemic preconditioning: current knowledge, unresolved questions, and future priorities. J Cardiovasc Pharmacol Ther 16:255–259. doi:10. 1177/1074248411409040
- Saxena A, Fish JE, White MD, Yu S, Smyth JW, Shaw RM, DiMaio JM, Srivastava D (2008) Stromal cell-derived factorlalpha is cardioprotective after myocardial infarction. Circulation 117:2224–2231. doi:10.1161/CIRCULATIONAHA.107.694992
- Schober A, Karshovska E, Zernecke A, Weber C (2006) SDFlalpha-mediated tissue repair by stem cells: a promising tool in cardiovascular medicine? Trends Cardiovasc Med 16:103–108. doi:10.1016/j.tcm.2006.01.006
- Serejo FC, Rodrigues LF Jr, da Silva Tavares KC, de Carvalho AC, Nascimento JH (2007) Cardioprotective properties of humoral factors released from rat hearts subject to ischemic preconditioning. J Cardiovasc Pharmacol 49:214–220. doi:10. 1097/FJC.0b013e3180325ad9
- 29. Shimizu M, Tropak M, Diaz RJ, Suto F, Surendra H, Kuzmin E, Li J, Gross G, Wilson GJ, Callahan J, Redington AN (2009) Transient limb ischaemia remotely preconditions through a humoral mechanism acting directly on the myocardium: evidence suggesting cross-species protection. Clin Sci (Lond) 117:191– 200. doi:10.1042/CS20080523
- Sierro F, Biben C, Martinez-Munoz L, Mellado M, Ransohoff RM, Li M, Woehl B, Leung H, Groom J, Batten M, Harvey RP, Martinez AC, Mackay CR, Mackay F (2007) Disrupted cardiac

development but normal hematopoiesis in mice deficient in the second CXCL12/SDF-1 receptor, CXCR7. Proc Natl Acad Sci USA 104:14759–14764. doi:10.1073/pnas.0702229104

- 31. Thielmann M, Kottenberg E, Kleinbongard P, Wendt D, Gedik N, Pasa S, Price V, Tsagakis K, Neuhauser M, Peters J, Jakob H, Heusch G (2013) Cardioprotective and prognostic effects of remote ischaemic preconditioning in patients undergoing coronary artery bypass surgery: a single-centre randomised, doubleblind, controlled trial. Lancet 382:597–604. doi:10.1016/S0140-6736(13)61450-6
- 32. Whittaker P, Przyklenk K (1994) Reduction of infarct size in vivo with ischemic preconditioning: mathematical evidence for protection via non-ischemic tissue. Basic Res Cardiol 89:6–15 n/a
- 33. Yan F, Yao Y, Chen L, Li Y, Sheng Z, Ma G (2012) Hypoxic preconditioning improves survival of cardiac progenitor cells: role of stromal cell derived factor-1alpha-CXCR4 axis. PLoS One 7:e37948. doi:10.1371/journal.pone.0037948