



# Treatments of osteoporosis increase bone material strength index in patients with low bone mass

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## Abstract

**Summary** Effects on bone material properties of two-year antiosteoporotic treatment were assessed using in vivo impact microindentation (IMI) in patients with low bone mineral density (BMD) values. Antiresorptive treatment, in contrast to vitamin D ± calcium treatment alone, induced BMD-independent increases in bone material strength index, measured by IMI, the magnitude of which depended on pretreatment values.

**Introduction** Bone material strength index (BMSi), measured by IMI in vivo, is reduced in patients with fragility fractures, but there is no information about changes in values during long-term therapy. In the present study, we assessed changes in BMSi in patients receiving antiosteoporotic treatments for periods longer than 12 months.

**Methods** We included treatment-naive patients with low bone mass who had a BMSi measurement with OsteoProbe® at presentation and consented to a repeat measurement after treatment.

**Results** We studied 54 patients (34 women), median age 58 years, of whom 30 were treated with bisphosphonates or denosumab (treatment group) and 24 with vitamin D ± calcium alone (control group). There were no differences in clinical characteristics between the two groups with the exception of a higher number of previous fragility fractures in the treatment group. Baseline hip BMD and BMSi values were lower in the treatment group. After  $23.1 \pm 6.6$  months, BMSi increased significantly in the treatment group ( $82.4 \pm 4.3$  vs  $79.3 \pm 4.1$ ;  $p < 0.001$ ), but did not change in the control group ( $81.5 \pm 5.2$  vs  $82.2 \pm 4.1$ ;  $p = 0.35$ ). Changes in BMSi with antiresorptives were inversely related with baseline values ( $r = -0.43$ ;  $p = 0.02$ ) but not with changes in BMD. Two patients in the control group with large decreases in BMSi values sustained incident fractures.

**Conclusion** In patients at increased fracture risk, antiresorptive treatments induced BMD-independent increases in BMSi values, the magnitude of which depended on pretreatment values.

**Keywords** Bisphosphonates · Bone material strength index (BMSi) · Denosumab · Fragility fracture · Impact microindentation · Reference point indentation

## Introduction

Impact microindentation (IMI), a method to assess tissue-level properties of cortical bone in vivo [1, 2], is being increasingly used in studies evaluating the contribution of such properties to bone fragility in humans [3–6]. The resistance of cortical bone to indentation, measured as bone material strength index (BMSi), was decreased in individuals with fragility fractures

compared with appropriate controls in several studies [7–10]. It is not related to bone mineral density (BMD) values [7–10], but, as recently shown, BMSi most likely assesses subperiosteal bone material properties [11].

Short-term effects of antiosteoporotic treatments on BMSi have been examined in patients initiating glucocorticoid therapy [12]. Glucocorticoids induced rapid decreases of BMSi, while treatments prevented this decline or even increased BMSi values within 7 weeks, up to 20 weeks, with no concurrent changes in BMD values. These results strongly suggested that BMSi could detect early, treatment-induced changes in bone material properties in glucocorticoid-treated patients. In addition, increases in BMSi have been reported in postmenopausal women 3 months after a high impact exercise program [13], and in HIV-infected patients after 1 year

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treatment with antiretroviral agents [14] or following gastric bypass surgery in obese subjects with and without type 2 diabetes [15]. However, the effect of antiosteoporotic treatments on BMSi in patients at increased risk of fractures not receiving glucocorticoids has not been investigated. Moreover, osteoporosis is a chronic disease requiring long-term treatment, and it is currently unknown whether changes in BMSi over time might provide information, additional to those obtained by BMD measurements, in treated individuals.

In the present study, we addressed this question in individuals at increased risk of fractures by measuring BMSi with the handheld IMI device OsteoProbe® before and after treatments given for periods longer than 12 months.

## Patients and methods

### Study design

Observational study evaluating longitudinal changes in BMSi with antiresorptive agents or vitamin D ± calcium alone in women and men with low bone mass attending the outpatient clinic of the Center for Bone Quality of the Leiden University Medical Center (LUMC) between March 2015 and September 2018.

### Patients

Included in the study were treatment-naïve subjects  $\geq 18$  years with osteopenia or osteoporosis who had an IMI measurement at presentation, were followed by one of the authors (NAD) for at least 1 year and consented to a repeat measurement. Exclusion criteria included any treatment affecting bone metabolism during follow-up—except calcium, vitamin D, bisphosphonates or denosumab—a metabolic bone disease other than osteoporosis, immobilization and local pathologies of the tibia or skin at the site of examination.

Treatment with antiresorptive agents (with vitamin D ± calcium) or vitamin D ± calcium alone was given according to Dutch national guidelines for the management of osteoporosis [16]. Treatment was initiated with oral bisphosphonates (BP) unless patients had epigastric complaints or did not want to receive oral BP; in these patients, iv BP or denosumab (DMAb) were given according to patients' preference and physician's judgement. All patients were followed at regular intervals in the outpatient clinic according to protocols of the Center for Bone Quality for the different treatment regimens.

The study was approved by the Medical Ethical Committee of the LUMC, and written informed consent was obtained from all individuals included in the study.

## Methods

Detailed medical history, clinical risk factors for fracture, fracture history with documentation of sites and dates of occurrence and information about use of medication were collected at baseline and follow-up visits. A fragility fracture was defined as any low-energy fracture, excluding those of the hands, feet and skull. Reasons and time of discontinuation or change to another antiresorptive agent during follow-up were recorded.

### Laboratory measurements

Serum calcium (albumin-corrected), creatinine and alkaline phosphatase were measured by semiautomated techniques. Plasma intact PTH was measured by immulite 2500 (Siemens Diagnostics, Breda, The Netherlands) and serum 25-hydroxyvitamin D (25-OH D) by the 25-OH-vitamin D TOTAL assay (DiaSorin D.A./N.V., Brussels, Belgium).

### Bone mineral density

Areal BMD was measured at the lumbar spine (L1–L4) and the left and right hip by dual-energy X-ray absorptiometry (DXA) with the Hologic QDR 4500 (Hologic Inc., Waltham, MA, USA). Average BMD values of the hip were used in the analysis. NHANES III reference values compatible with reference values of the Dutch population were used to calculate T-scores. Osteopenia and osteoporosis were diagnosed according to WHO criteria.

### Vertebral fracture assessment

Spine radiographs for the detection of vertebral deformities were performed at baseline in all patients. During follow-up, radiographs were only performed if there was loss of height  $> 3$  cm or clinical complaints. Radiographs were independently evaluated by two of the authors (NAD and MS) using the semiquantitative method of Genant et al. [17], and only fractures grade 2 or higher were considered in the analysis.

### Bone material strength index

BMSi was measured in all patients by IMI using the handheld microindenter device (OsteoProbe® RUO, Active Life Scientific, CA, USA) at the midshaft of the tibia. Measurements were performed by two experienced operators (NAD and FM) according to our previously published protocol [7, 8, 18, 19]. The patient was placed in a decubitus supine position with the tibia in external rotation. The measurement site was defined as the mean distance between the medial malleolus and the distal apex of the patella. After disinfection and local anaesthesia of the skin and periosteum with

lidocaine 1%, the test probe was gently inserted in the skin until the bone surface was reached. It was ensured that the test probe was always perpendicular to the bone surface during measurements. The operator was not allowed to check the measurements on the computer screen before these were classified as “well performed”, “adequate” or “poorly performed”. Measurements were classified as “well performed” when the test probe was exactly perpendicular to the bone surface, as “adequate” when the test probe was within acceptable deviation from the bone surface [2] and as “poorly performed” when the operator judged that the test probe was not appropriately placed. “Poorly performed” measurements are usually due to slipping of the test probe, moving of the subject’s leg, indenting the same spot twice, or failure to place the device perpendicularly to the bone surface and were manually discarded. In addition, the first measurement was systematically discarded since there is often inadequate penetration of the probe through the periosteum. After at least five adequate measurements (range 6 to 15, mean  $10.0 \pm 1.8$  measurements), five additional measurements were performed on a polymethylmethacrylate (PMMA) calibration phantom. BMSi was calculated by the computer software. The operator was not blinded to the patient’s treatment, but was blinded to baseline BMSi results, and classification of measurements was done before checking the results on the computer screen. The intraobserver coefficient of variation (CV) was 2.2%, and the interobserver CV was 1.6%.

### Statistical analysis

Results are presented as mean  $\pm$  SD unless otherwise stated. Descriptive statistics were used to describe clinical and laboratory parameters. Between-group differences in baseline characteristics were assessed using a Student’s *t* test or Mann-Whitney *U* test and chi-square test or Fisher’s exact test for normally and not normally distributed continuous and for categorical variables, respectively. Within-group changes in BMSi and BMD were assessed using a paired *t* test. Normality of the distribution of BMSi and BMD was checked by a Kolmogorov-Smirnov test and visually with histograms. Between-group differences in % changes from baseline to follow-up measurements were assessed by a Student’s *t* test. ANCOVA models with % change in BMSi as outcome variable, adjusted for baseline BMSi, BMD and fragility fracture, were used to compare % change in BMSi between groups. Correlations between BMSi and BMD values and between % changes in BMSi and % changes in BMD or duration of treatment were examined by a Pearson’s test. To determine independent effects of factors possibly influencing changes in BMSi, a multiple linear regression analysis was used. Age, gender, fragility fracture, baseline BMSi, treatment duration as well as examiner of the first and the second IMI measurement, respectively, were included in the model. A

power calculation was performed using a difference of 3.1% in BMSi values. This difference was previously found between patients with and patients without fragility fractures (BMSi  $79.9 \pm 0.6$  vs  $82.4 \pm 1.0$ ) and considered to be clinically relevant [7]. The sample size to detect this difference in BMSi from baseline to follow-up with a standard deviation of 5 and power of 0.8 at a significance level of 0.05 was calculated to be 24 per group. All analyses were performed using SPSS software for Windows (version 25.0; SPSS Inc., Chicago, IL, USA). A *p* value  $< 0.05$  was considered to be statistically significant. Graphs were constructed with Graphpad Prism (version 8.0; Graphpad Software Inc., La Jolla, CA, USA).

### Results

Fifty-four patients were included in the study (Fig. 1). These were 34 women and 20 men with median age 58.0 years (IQR 48.5–63.3 years). Thirty-one patients had osteoporosis (57.4%) and 23 had osteopenia (42.6%); 23 patients (7 men) had sustained one or more fragility fractures (11 vertebral, 2 hip, 14 non-hip/non-vertebral) at the time of the first IMI measurement (Table 1). Mean time to second measurement was  $23.1 \pm 6.6$  months.

Baseline characteristics of patients who were excluded from the study, either because they were referred to their general practitioners, were followed by other physicians in our centre or were lost to follow-up (Fig. 1), were not different from those of the included patients; age  $56.2 \pm 14.2$  vs  $55.3 \pm 13.0$  years, *p* = 0.637; female gender 59.5% vs 63.0%, *p* = 0.665; prevalent fragility fractures 47.7% vs 42.6%, *p* = 0.500; BMSi  $80.7 \pm 5.6$  vs  $80.6 \pm 4.3$ , *p* = 0.894.

Of the 54 patients included in the analysis, 30 started antiresorptive therapy after a median of 4 days following the first IMI measurement (treatment group); 23 patients received BPs (16 oral, 7 iv) and 7 DMAB. Twenty-four patients received vitamin D  $\pm$  calcium alone (control group) (Table 1). Patients in the treatment group had sustained significantly more fragility fractures compared with those of the control group (25 vs 4, *p* < 0.01), due mainly to the higher prevalence of vertebral fractures in the former group (14 vs 0, *p* < 0.01).

### Bone mineral density

In the treatment group, 20 patients had osteoporosis (T-score  $\leq -2.5$ ) and 10 had osteopenia (T-score between  $-2.5$  and  $-1.0$ ); in the control group, 11 patients had osteoporosis and 13 had osteopenia (*p* = 0.124 between the two groups). Lumbar spine (LS) BMD was lower in the treatment group, but the difference between the two groups was not significant ( $0.81 \pm 0.08$  vs  $0.86 \pm 0.10$  g/cm<sup>2</sup>, *p* = 0.06); femoral neck (FN) BMD and total hip (TH) BMD were significantly lower

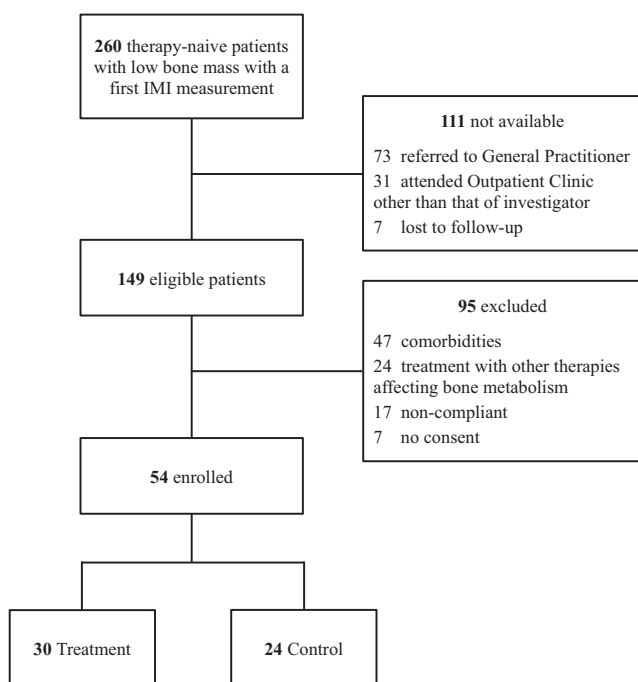


Fig. 1 Patient flowchart

in the treatment group (FN  $0.66 \pm 0.09$  vs  $0.70 \pm 0.09$  g/cm<sup>2</sup>,  $p = 0.041$ ; TH  $0.77 \pm 0.10$  vs  $0.84 \pm 0.12$  g/cm<sup>2</sup>,  $p = 0.016$ ).

Antiresorptive treatment was associated with significant increases in LS-BMD ( $6.3 \pm 5.3\%$ ) and TH-BMD ( $1.1 \pm 2.9\%$ ) but not in FN-BMD ( $1.2 \pm 3.6\%$ ) (Fig. 2). In the control group, LS-BMD did not change significantly during follow-up ( $-1.3 \pm 3.6\%$ ), while hip BMD decreased significantly at both measured sites (FN-BMD  $-2.7 \pm 4.6\%$ , TH-BMD  $-2.4 \pm 2.9\%$ ) (Fig. 2).

### Bone material strength index

Consistent with our previous studies, baseline BMSi values of all studied patients were not associated with baseline BMD values at any site (LS  $r = -0.072$ ,  $p = 0.617$ ; FN  $r = 0.067$ ,  $p = 0.631$ ; TH  $r = 0.048$ ,  $p = 0.728$ ) and were significantly lower in patients with fragility fractures compared with those without ( $78.5 \pm 3.0$  vs  $82.1 \pm 4.5$ ,  $p = 0.002$ ).

Table 1 Baseline characteristics of 54 patients

Characteristics	All patients	Treatment	Control	<i>p</i> value
<i>n</i>	54	30	24	
Age, years	$58.0 \pm 1.8$	$59.0 \pm 1.8$	$55.0 \pm 3.2$	0.16
Male/female	20/34	8/22	12/12	0.11
BMI, kg/m <sup>2</sup>	$24.2 \pm 3.6$	$24.0 \pm 3.4$	$24.5 \pm 4.0$	0.67
Smoking, <i>n</i>	7	6	1	0.12
Alcohol use $\geq 3$ U/d, <i>n</i>	5	2	3	0.65
Previous fragility fracture, <i>n</i> (%)	23 (42.6)	19 (63.3)	4 (16.7)	0.001
Hip fracture, <i>n</i> (%)	2 (3.7)	2 (6.7)	0 (0.0)	0.50
NHNV fracture, <i>n</i> (%)	14 (25.9)	10 (33.3)	4 (16.7)	0.17
Vertebral fracture, <i>n</i> (%)	11 (20.4)	11 (40.0)	0 (0.0)	0.001
Calcium <sup>a</sup> , mmol/L	$2.32 \pm 0.08$	$2.34 \pm 0.08$	$2.31 \pm 0.08$	0.11
Creatinine <sup>b</sup> , umol/L	$76.1 \pm 15.0$	$74.0 \pm 13.8$	$78.8 \pm 16.2$	0.24
25-OH D <sup>c</sup> , nmol/L	$80.9 \pm 27.7$	$78.7 \pm 29.0$	$83.6 \pm 26.4$	0.52
PTH <sup>d</sup> , pmol/L	$3.5 \pm 1.3$	$3.6 \pm 1.6$	$3.2 \pm 0.9$	0.13
LS-BMD T-score	$-2.1 \pm 0.8$	$-2.3 \pm 0.7$	$-1.9 \pm 0.9$	0.10
FN-BMD T-score	$-1.7 \pm 0.7$	$-1.9 \pm 0.7$	$-1.5 \pm 0.6$	0.08
TH-BMD T-score	$-1.3 \pm 0.7$	$-1.5 \pm 0.6$	$-1.1 \pm 0.8$	0.026
BMSi	$80.6 \pm 4.3$	$79.3 \pm 4.1$	$82.2 \pm 4.1$	0.014

NHNV, Non-hip/non-vertebral; BMD, bone mineral density; LS, lumbar spine; FN, femoral neck; TH, total hip; BMSi, bone material strength index. Values are expressed as mean  $\pm$  SD. Age is expressed as median  $\pm$  SEM. *p* values are displayed for treatment vs control

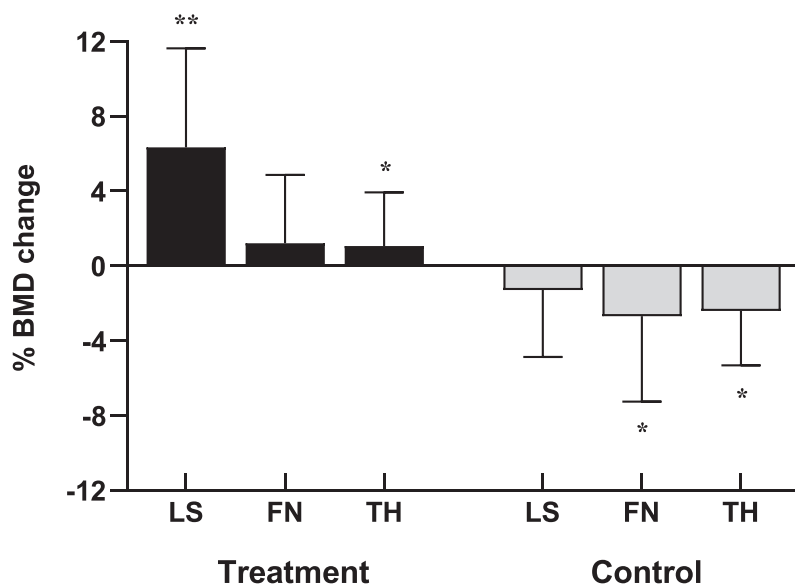
<sup>a</sup> Calcium (albumin-corrected) reference range, 2.15–2.55 mmol/L

<sup>b</sup> Creatinine reference range, 64–104 umol/L for males; 49–90 umol/L for females

<sup>c</sup> 25-OH vitamin D reference range, 50–250 nmol/L

<sup>d</sup> PTH reference range, 0.7–8.0 pmol/L

**Fig. 2** Mean ( $\pm$ SD) percentage changes in bone mineral density (BMD) at follow-up compared with baseline in the treatment (black bars) and control group (grey bars) at the lumbar spine (LS), femoral neck (FN) and total hip (TH). \*Significantly different from baseline,  $p < 0.05$  and \*\* $p < 0.001$



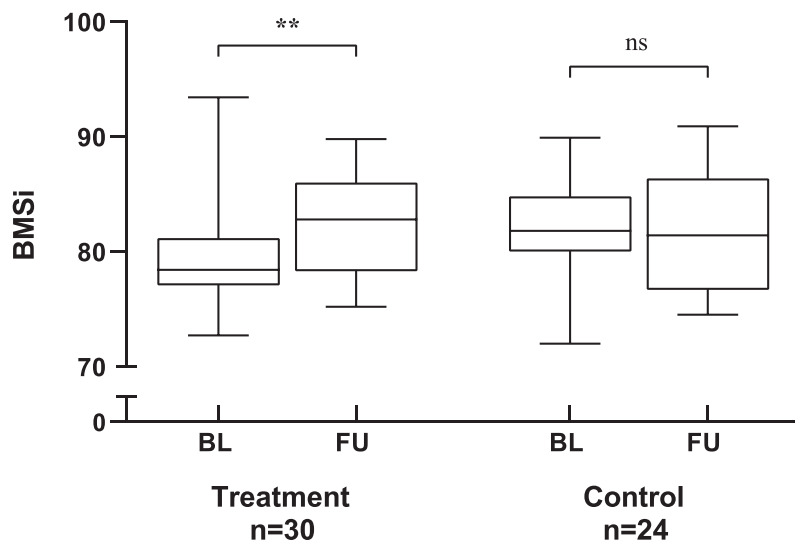
**Changes during follow-up**

Baseline BMSi values of the treatment group were significantly lower than those of the control group (Table 1) and increased significantly by  $4.0 \pm 5.2\%$  to values similar to those of the control group at baseline (from  $79.3 \pm 4.1$  to  $82.4 \pm 4.3$ ,  $p < 0.001$ ) (Fig. 3). The increase in BMSi with antiresorptive treatment was inversely related with baseline BMSi ( $r = -0.432$ ,  $p = 0.017$ ) (Fig. 4) but not with the duration of treatment ( $r = 0.089$ ,  $p = 0.639$ ) or with the BMD changes (LS  $r = -0.076$ ,  $p = 0.712$ ; FN  $r = 0.266$ ,  $p = 0.155$ ; TH  $r = -0.031$ ,  $p = 0.870$ ). In contrast, in the control group, BMSi values decreased during the period of observation but not significantly (from  $82.2 \pm 4.1$

to  $81.5 \pm 5.2$ ,  $p = 0.353$ ;  $-0.7 \pm 3.9\%$ ) (Fig. 3). The difference in % BMSi changes between the two groups was significant,  $p < 0.001$  (also after adjusting for baseline BMSi ( $p = 0.004$ ), baseline BMD ( $p = 0.007$ ) and prevalent fragility fractures ( $p = 0.016$ )).

In both BP- and DMAB-treated patients for the whole observation period, BMSi increased significantly (BP from  $80.1 \pm 4.6$  to  $82.5 \pm 4.4$  ( $3.1 \pm 5.6\%$ ),  $p = 0.037$ ; DMAB from  $78.1 \pm 3.1$  to  $84.2 \pm 4.3$  ( $7.8 \pm 3.1\%$ ),  $p = 0.001$ ). The percentage increase in BMSi was higher in the DMAB-treated patients ( $p = 0.014$ ), but after adjusting for baseline BMSi, the difference between the two treatments was not significant anymore ( $7.1 \pm 4.8\%$  vs  $3.4 \pm 4.7\%$ ,  $p = 0.104$ ). Numbers however were small.

**Fig. 3** Bone material strength index (BMSi) in patients with antiresorptive treatment (treatment) and those without antiresorptive treatment (control) at baseline (BL) and follow-up (FU). Data are shown in box-whisker plots and statistical differences are displayed for BMSi. Boxes indicate median and interquartile range. Bars indicate minimum and maximum values. \*\* $p < 0.001$





and is directly proportional to bending stiffness and failure moment being superior to total mineral density, measured by micro-CT, in predicting bone strength [30, 31]. Bisphosphonates and hormone replacement therapy given for 1 to 3 years to animals or humans increased significantly spectroscopically measured MM [32–35]. Moreover, inhibition of receptor activator of nuclear factor kappa-B ligand (RANKL) by osteoprotegerin (OPG) significantly increased MM in genetically modified ovariectomized mice to levels of wild-type animals (*E. Paschalis* personal communication). It may, therefore, be that BMSi measurements capture the observed increases in MM by antiresorptive therapies. Notably, prolongation of treatment with bisphosphonate, as in the FLEX study with alendronate, is not associated with continuous increase in MM [36] indicating that the amount of mineral taken up by the matrix in non-pathological mineralisation is limited, or self-regulated; this finding is similar to the observed plateau of bone matrix mineralisation between 5 and 10 years treatment of patients with osteoporosis with denosumab [37] and may explain the lack of further increases in BMSi values when these reach normal levels shown in our study.

Notably, patients on antiresorptive treatments in our study sustained no fractures, whereas fractures occurred only in two patients in the control group. These two patients showed significant decreases in BMSi values (−8.5% and −8.9%) during follow-up, whereas the corresponding decreases in BMD were variable and ranged between −1.3% and −7.0%. Moreover, in a cross-sectional study of postmenopausal women on long-term treatment with bisphosphonates (4 to 14 years) BMSi values were significantly lower in women with incident fractures compared with those without fractures [28]. Thus, IMI measurements may not only be a useful, complementary investigation in the assessment of fracture risk but may also provide additional long-term information about the effects of treatments on bone material properties and their association to bone fragility. This needs, however, to be confirmed in appropriately designed studies.

A limitation of the study is the inclusion of only patients followed by one of the authors. However, baseline characteristics of patients who had a baseline measurement but were not included in the study did not differ from those of patients reported here and treatment protocols were the same. Moreover, all measurements were performed by two experienced investigators that minimized intra- and interobserver variability, a known limitation of the application of IMI technique [1, 2, 6].

In summary, our study, the first to investigate longitudinal changes in BMSi with antiosteoporotic treatments in patients with low bone mass and increased fracture risk, demonstrates that IMI can capture BMD-independent, treatment-induced

increases in BMSi, the magnitude of which depends on pre-treatment values.

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## Compliance with ethical standards

**Conflict of interest** Manuela Schoeb, Frank Malgo, Joséphine J.M. Peeters and Elizabeth M. Winter declare that they have no conflict of interest. Socrates E. Papapoulos and Natasha M. Appelman-Dijkstra are unpaid members of the Scientific Board of Active Life Scientific, manufacturer of OsteoProbe®.

**Statement of human rights** The study was approved by the Medical Ethical Committee of the LUMC. All procedures performed were in accordance with the ethical standards of the institutional research committee and with the 1964 Helsinki declaration and its later amendments or comparable ethical standards.

**Informed consent** Written informed consent was obtained from all individual participants included in the study.

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## References

1. Diez-Perez A, Bouxsein ML, Eriksen EF, Khosla S, Nyman JS, Papapoulos S et al (2016) Technical note: recommendations for a standard procedure to assess cortical bone at the tissue-level in vivo using impact microindentation. *Bone Rep* 5:181–185
2. Bridges D, Randall C, Hansma PK (2012) A new device for performing reference point indentation without a reference probe. *Rev Sci Instrum* 83(4):044301
3. Herrera S, Diez-Perez A (2017) Clinical experience with microindentation in vivo in humans. *Bone*. 95:175–182
4. Allen MR, McNerny EM, Organ JM, Wallace JM (2015) True gold or pyrite: a review of reference point indentation for assessing bone mechanical properties in vivo. *J Bone Miner Res* 30(9):1539–1550
5. Diez-Perez A, Farr J (2018) Reference point indentation. In: Bilezikian JP (ed) *Primer on the metabolic bone diseases and disorders of mineral metabolism*, 9th edn. Wiley, pp 287–292

6. Schoeb M, Hamdy NAT, Malgo F, Winter EM, Appelman-Dijkstra NM (2020) Added value of impact microindentation in the evaluation of bone fragility: a systematic review of the literature. *Front Endocrinol* 11(15)
7. Malgo F, Hamdy NA, Papapoulos SE, Appelman-Dijkstra NM (2015) Bone material strength as measured by microindentation in vivo is decreased in patients with fragility fractures independently of bone mineral density. *J Clin Endocrinol Metab* 100(5):2039–2045
8. Malgo F, Hamdy NAT, Papapoulos SE, Appelman-Dijkstra NM (2017) Bone material strength index as measured by impact microindentation is low in patients with fractures irrespective of fracture site. *Osteoporos Int* 28(8):2433–2437
9. Rufus-Membere P, Holloway-Kew KL, Diez-Perez A, Kotowicz MA, Pasco JA (2019) Associations between bone impact microindentation and clinical risk factors for fracture. *Endocrinology*. en.2019–00415
10. Sosa DD, Eriksen EF (2017) Reduced bone material strength is associated with increased risk and severity of osteoporotic fractures. An impact microindentation study. *Calcif Tissue Int* 101(1):34–42
11. Rokidi S, Bravenboer N, Gamsjaeger S, Misof B, Blouin S, Chavassieux P et al (2020) Impact microindentation assesses subperiosteal bone material properties in humans. *Bone*. 131: 115110
12. Mellibovsky L, Prieto-Alhambra D, Mellibovsky F, Guerri-Fernandez R, Nogues X, Randall C et al (2015) Bone tissue properties measurement by reference point indentation in glucocorticoid-induced osteoporosis. *J Bone Miner Res* 30(9): 1651–1656
13. Sundh D, Nilsson M, Zoulakis M, Pasco C, Yilmaz M, Kazakia GJ, Hellgren M, Lorentzon M (2018) High-impact mechanical loading increases bone material strength in postmenopausal women—a 3-month intervention study. *J Bone Miner Res* 33(7):1242–1251
14. Guerri-Fernandez R, Lerma-Chippirraz E, Fernandez Marron A, Garcia-Giralt N, Villar-Garcia J, Soldado-Folgado J et al (2018) Bone density, microarchitecture, and tissue quality after 1 year of treatment with tenofovir disoproxil fumarate. *Aids*. 32(7):913–920
15. Blom-Hogestol IK, Mala T, Kristinsson JA, Brunborg C, Gulseth HL, Eriksen EF (2019) Changes in bone quality after Roux-en-Y gastric bypass: a prospective cohort study in subjects with and without type 2 diabetes. *Bone*. 130:115069
16. CBO. Richtlijn Osteoporose en Fractuurpreventie 2011. <https://www.volksgezondheidenzorg.info/bestanden/documenten/cbo-richtlijn-osteoporose-en-fractuurpreventie-2011>. Accessed 2 Dec 2019
17. Genant HK, Wu CY, van Kuijk C, Nevitt MC (1993) Vertebral fracture assessment using a semiquantitative technique. *J Bone Miner Res* 8(9):1137–1148
18. Malgo F, Hamdy NA, Papapoulos SE, Appelman-Dijkstra NM (2017) Impact microindentation: consistency of serial measurements and alterations in patients with Paget's disease of the tibia. *J Bone Miner Res* 32(12):2375–2380
19. Malgo F, Hamdy NA, Rabelink TJ, Kroon HM, Claessen KM, Pereira AM et al (2017) Bone material strength index as measured by impact microindentation is altered in patients with acromegaly. *Eur J Endocrinol* 176(3):339–347
20. Furst JR, Bandeira LC, Fan WW, Agarwal S, Nishiyama KK, McMahon DJ, Dworakowski E, Jiang H, Silverberg SJ, Rubin MR (2016) Advanced glycation endproducts and bone material strength in type 2 diabetes. *J Clin Endocrinol Metab* 101(6): 2502–2510
21. Guerri-Fernandez R, Molina-Morant D, Villar-Garcia J, Herrera S, Gonzalez-Mena A, Guelar A et al (2017) Bone density, microarchitecture, and tissue quality after long-term treatment with tenofovir/emtricitabine or abacavir/lamivudine. *J Acquir Immune Defic Syndr* 75(3):322–327
22. Perez-Saez MJ, Herrera S, Prieto-Alhambra D, Vilaplana L, Nogues X, Vera M et al (2017) Bone density, microarchitecture, and material strength in chronic kidney disease patients at the time of kidney transplantation. *Osteoporos Int* 28(9):2723–2727
23. Perez-Saez MJ, Herrera S, Prieto-Alhambra D, Nogues X, Vera M, Redondo-Pachon D et al (2017) Bone density, microarchitecture, and tissue quality long-term after kidney transplant. *Transplantation*. 101(6):1290–1294
24. Duarte Sosa D, Vilaplana L, Guerri R, Nogues X, Wang-Fagerland M, Diez-Perez A et al (2015) Are the high hip fracture rates among Norwegian women explained by impaired bone material properties? *J Bone Miner Res* 30(10):1784–1789
25. Sosa DD, Eriksen EF (2016) Women with previous stress fractures show reduced bone material strength: microindentation measurements in a retrospective case-control study of 60 subjects. *Acta Orthop* 87(6):626–631
26. Rufus-Membere P, Holloway-Kew KL, Diez-Perez A, Kotowicz MA, Pasco JA (2018) Feasibility and tolerability of bone impact microindentation testing: a cross-sectional, population-based study in Australia. *BMJ Open* 8(12):e023959
27. Popp KL, Caksa S, Martinez-Betancourt A, Yuan A, Tsai J, Yu EW et al (2019) Cortical bone material strength index and bone microarchitecture in postmenopausal women with atypical femoral fractures. *J Bone Miner Res* 34(1):75–82
28. Nogues X, Prieto-Alhambra D, Guerri-Fernandez R, Garcia-Giralt N, Rodriguez-Morera J, Cos L et al (2017) Fracture during oral bisphosphonate therapy is associated with deteriorated bone material strength index. *Bone*. 103:64–69
29. Paschalis EP, Gamsjaeger S, Klaushofer K (2017) Vibrational spectroscopic techniques to assess bone quality. *Osteoporos Int* 28(8): 2275–2291
30. Boskey AL (1992) Mineral-matrix interactions in bone and cartilage. *Clin Orthop Relat Res* (281):244–74
31. Donnelly E, Chen DX, Boskey AL, Baker SP, van der Meulen MC (2010) Contribution of mineral to bone structural behavior and tissue mechanical properties. *Calcif Tissue Int* 87(5):450–460
32. Boskey AL, Spevak L, Weinstein RS (2009) Spectroscopic markers of bone quality in alendronate-treated postmenopausal women. *Osteoporos Int* 20(5):793–800
33. Gourion-Arsiquaud S, Allen MR, Burr DB, Vashishth D, Tang SY, Boskey AL (2010) Bisphosphonate treatment modifies canine bone mineral and matrix properties and their heterogeneity. *Bone*. 46(3): 666–672
34. Gamsjaeger S, Buchinger B, Zwettler E, Recker R, Black D, Gasser JA et al (2011) Bone material properties in actively bone-forming trabeculae in postmenopausal women with osteoporosis after three years of treatment with once-yearly zoledronic acid. *J Bone Miner Res* 26(1):12–18
35. Paschalis EP, Boskey AL, Kassem M, Eriksen EF (2003) Effect of hormone replacement therapy on bone quality in early postmenopausal women. *J Bone Miner Res* 18(6):955–959
36. Hassler N, Gamsjaeger S, Hofstetter B, Brozek W, Klaushofer K, Paschalis EP (2015) Effects of long-term alendronate treatment on postmenopausal osteoporosis bone material properties. *Osteoporos Int* 26(1):339–352
37. Dempster DW, Brown JP, Fahrleitner-Pammer A, Kendler D, Rizzo S, Valter I, Wagman RB, Yin X, Yue SV, Boivin G (2018) Effects of long-term denosumab on bone histomorphometry and mineralization in women with postmenopausal osteoporosis. *J Clin Endocrinol Metab* 103(7):2498–2509

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