

John Arcaroli  
Jeff Sankoff  
Nianjun Liu  
David B. Allison  
James Maloney  
Edward Abraham

## Association between urokinase haplotypes and outcome from infection-associated acute lung injury

Received: 6 July 2007  
Accepted: 12 October 2007  
Published online: 10 November 2007  
© Springer-Verlag 2007

**Electronic supplementary material**  
The online version of this article  
(doi:10.1007/s00134-007-0930-6) contains  
supplementary material, which is available  
to authorized users.

Supported by NIH grant R01 HL76206.

J. Arcaroli · J. Maloney  
University of Colorado at Denver and  
Health Sciences Center, Division of  
Pulmonary Sciences and Critical Care  
Medicine, Department of Surgery,  
Denver Colorado, USA

J. Sankoff  
University of Colorado at Denver and  
Health Sciences Center, Division of  
Emergency Medicine, Department of  
Surgery,  
Denver Colorado, USA

N. Liu · D. B. Allison  
University of Alabama at Birmingham,  
Section on Statistical Genetics, Department  
of Biostatistics,  
Birmingham Alabama, USA

E. Abraham (✉)  
University of Alabama at Birmingham,  
Department of Medicine, Chairman's  
Office,  
1530 3rd Avenue South, BDB 420,  
Birmingham 35294-0012, Alabama, USA  
e-mail: eabraham@uab.edu

**Abstract** *Objective:* Alterations in coagulation, including elevated pulmonary and systemic concentrations of urokinase, are frequent in patients with acute lung injury (ALI). Urokinase potentiates neutrophil activation and contributes to the severity of pulmonary injury in preclinical models of ALI. The objective of this study was to examine associations between polymorphisms and haplotypes of urokinase with risk for and outcomes from ALI. *Design:* Prospective cohorts of healthy European-American adults and those with infection-associated ALI. *Setting:* Academic medical centers participating in NIH funded studies of low tidal volume ventilation for ALI. *Patients:* Controls were 175 healthy European-American subjects. Patients were 252 individuals with infection-associated ALI, prospectively followed for 60

days for mortality. *Interventions:* Genetic polymorphisms and haplotypes in the urokinase gene were determined. *Measurements and main results:* Six polymorphisms, rs1916341, rs2227562, rs2227564, rs2227566, rs2227571, and rs4065, defining 98% of all urokinase haplotypes, were analyzed. There were no statistically significant associations between any single urokinase polymorphism or haplotype and risk for developing ALI. In contrast, there was a statistically significant relationship between the CGCCCC haplotype and both 60-day mortality and ventilator-free days that remained present in a multivariate analysis controlling for age and sex ( $p = 0.033$  for 60-day mortality and  $< 0.001$  for ventilator-free days). *Conclusions:* These results identify a specific urokinase haplotype as a genetic risk factor for higher mortality and more severe clinical outcome in patients with infection-associated ALI.

**Keywords** Acute lung injury · Genetics · Urokinase · Sepsis · Polymorphisms · Haplotypes

## Introduction

Acute lung injury (ALI) is clinically defined by hypoxemia and bilateral infiltrates on chest radiographs without evidence of left-sided cardiac failure [1]. Histopathologic examination of the lungs obtained from experimental models or patients with ALI demonstrates interstitial edema, microvascular thrombi, neutrophil accumulation, and alveolar fibrin formation [2, 3]. Alterations in pathways related to coagulation and fibrinolysis are present in patients with ALI and are postulated to play an important role in mediating the microvascular and inflammatory changes in the lungs that accompany ALI [3, 4].

Urokinase is a serine protease that cleaves plasminogen to generate plasmin, a potent mediator of fibrinolysis [5, 6]. In addition to its fibrinolytic properties, urokinase has recently been shown to enhance neutrophil activation and participate in the development of experimental ALI [7]. Bronchoalveolar lavage concentrations of urokinase are increased in human volunteers after local instillation of LPS into the lung [8]. In the setting of pneumonia, urokinase levels are elevated in the involved segments of the lungs [9, 10]. Both serum and bronchoalveolar lavage concentrations of urokinase are increased in patients with ALI [9, 11]. In patients with ALI or severe sepsis, levels of plasminogen activator inhibitor 1 (PAI-1), which inhibits the fibrinolytic properties of urokinase through binding to its protease domain, are usually increased to a greater degree than are those of urokinase, resulting in a procoagulant state that is present both systemically and in the lungs [12–15]. However, portions of the urokinase molecule that are not involved in fibrinolysis or interaction with PAI-1, such as its kringle sequence, also have proinflammatory effects and participate in the pathogenesis of experimental ALI [16, 17].

The urokinase gene is located in the 10q24 chromosomal region [18]. Polymorphisms in urokinase appear to be functionally significant and are associated with diseases involving altered immune regulation, including asthma, rheumatoid arthritis, type 1 diabetes, colorectal cancer, and late-onset Alzheimer's disease [19–24]. Therefore, given the importance of alterations in coagulation, neutrophil activation, and inflammation in ALI, and the ability of urokinase to participate in all of these processes, we hypothesized that genetic variants in urokinase might be

associated with susceptibility to or outcome from ALI. To explore this hypothesis, we examined outcome in a well-characterized cohort of patients with infection-associated ALI, and found that a specific urokinase haplotype was associated with survival and need for prolonged ventilatory support in ALI.

## Materials and methods

### Volunteer selection

Healthy white volunteers, 19–89 years of age, were recruited to determine urokinase polymorphisms. The sample included 98 males (mean age  $30.0 \pm 6.7$  years, range 19–57 years) and 77 females (mean age  $32.0 \pm 10.0$  years, range 19–89 years). The study was approved by the institutional review board, and each volunteer signed an informed consent document.

### Patients with acute lung injury

All patients included in this study had been enrolled in NIH ARDS Network studies and had received low tidal volume ventilation using previously published protocols [25]. All patients were European-American and had infection-associated ALI, as characterized by either systemic or localized pulmonary infection as the investigator-identified primary etiology for ALI. ALI was defined by standard criteria. Septic shock was defined as a systolic blood pressure of  $< 90$  mmHg for at least 30 min despite fluid replacement or the use of inotropic support to maintain blood pressure. Exclusion criteria included age below 18 years, a neurologic condition that could impair weaning from mechanical ventilation, severe chronic respiratory disease, severe chronic liver disease (defined as a Child–Pugh score of  $> 10$ ), burns covering  $> 30\%$  of total body surface area, malignancy or other irreversible condition for which the mortality at 6 months was estimated to be above 50%, use of high-dose immunosuppressive therapy, and a history of lung or bone marrow transplantation. Table 2 shows characteristics of the patients. This genetic analysis study was approved by the University of Colorado Health Sciences Center and the University of Alabama

**Table 1** Genotyped single nucleotide polymorphism (SNP) characteristics

SNP ID number	Base change	Minor allele frequency	Chromosomal location	Genomic features
rs1916341	A/C	0.41	75341168	Intron
rs2227562	G/A	0.11	75342967	Intron
rs2227564	C/T	0.34	75343107	Exon 5
rs2227566	T/C	0.43	75343737	Nonsynonymous P141L
rs2227571	T/C	0.48	75343885	Intron
rs4065	T/C	0.38	75346470	3'UTR

at Birmingham institutional review boards. Consent was obtained from all patients or their surrogates as part of their enrollment into the NIH protocols.

### Allelic discrimination

Real-time polymerase chain reaction (PCR) allelic discrimination assays were developed by the assay-by-design service offered by Applied Biosystems (Foster City, CA). Probe and primer combinations were designed for the six urokinase single nucleotide polymorphisms (SNPs), rs1916341, rs2227562, rs2227564, rs2227566, rs2227568, and rs4065, which capture all haplotypes with frequency greater than 2.2% in the European-American population as per the Seattle SNPs database (Table 1). PCR reactions were performed in a final volume of 25  $\mu$ l, which consisted of 1–25 ng of DNA diluted in dH<sub>2</sub>O, 12.5  $\mu$ l of 2  $\times$  TaqMan Universal PCR Master Mix, and 1.25  $\mu$ l of 20  $\times$  TaqMan SNP genotyping Assay Mix. PCR was performed using an Applied Biosystems 7300 Real-Time PCR system. There were no missing genotypes. An approach similar to the one proposed by Scheet and Stephens [26] was used to check for genotyping errors. There was no statistical significant evidence for the appearance of genotyping errors in the data.

### Single-marker and haplotype analysis

For single-marker analysis, two approaches were used to analyze the data. The first was based on a set of score tests for binary response, as described by Zhao et al. [27–29], without considering other covariates; the other analysis was based on a regression approach where the other covariates were included [30–32]. For haplotype analysis, two regression-based methods (for binary, continuous, and time-to-event responses) as described by Schaid et al. [33] and Lake et al. [34] and by Lin and Zeng [30–32] where the other covariates were included and excluded, were employed. For haplotype analysis, we performed both joint-effect analysis and separate-effect analysis. All significance levels reported are two-tailed.

## Results

### Characteristics of the patient population

A total of 252 European-American patients with infection-associated acute lung injury (ALI) were available for analysis (Table 2). The primary source of infection was the lungs, involving 55% of these patients. Men and women were equally represented in the ALI patients. An additional population of 175 healthy volunteers was recruited for comparison.

**Table 2** Characteristics of acute lung injury patients

<i>N</i>	252
Age, years (mean $\pm$ SD, range)	51.3 $\pm$ 15.6, 17–90
Male	51.9 $\pm$ 16.1, 17–90
Female	50.8 $\pm$ 15.0, 22–88
Male, %	50.0
Apache III (mean $\pm$ SD)	96.2 $\pm$ 29.7
Male	96.6 $\pm$ 31.5
Female	95.8 $\pm$ 28.0
Mechanical ventilation, %	100
60-Day mortality, %	25.0
Male	26.0
Female	24.0
Source of infection	
Lung: primary, %	55.0
Lung: secondary, %	21.0
Peritoneum, %	4.0
Vascular line, %	4.0
Skin, %	2.0
Gastrointestinal/biliary tract, %	2.0
Other, %	12.0

Case–control comparison of urokinase polymorphisms and haplotypes in healthy control and acute lung injury populations

Prior to analysis, Hardy–Weinberg equilibrium (HWE) was tested using the approaches described by Guo and Thompson [35] as implemented in Arlequin version 3.0a [<http://anthro.unige.ch/software/arlequin/>]. SNP 5 showed marginal departure from HWE (unadjusted *p*-value 0.046) on the pooled cases and controls. When tested on cases and controls separately, HWE held at all SNPs (unadjusted *p*-values > 0.092). For this analysis, all patients and controls were included.

As an initial approach, single-marker analysis of the six urokinase polymorphisms was undertaken, using the methods by Zhao et al. [28, 29], as implemented in the Nonlinear program. As shown in Table 3, there were no significant differences between the frequency of polymorphisms in the control population and patients with ALI. Analysis, with age and sex included, was also performed, using the approach described by Lin and Zeng [30–32], as implemented in the HAPSTAT program. No significant evidence of association between any polymorphism and risk for ALI was found with this approach.

Six haplotypes with frequencies larger than 0.01 for urokinase were identified (Table 4). Haplotype analyses were conducted using both a joint-effects model [30–34], in which all haplotypes except the most frequent one are compared to the most frequent haplotype simultaneously, and a separate-effect model [30–32] in which each haplotype is compared to all of the other haplotypes (i.e., all other haplotypes pooled together as baseline), one at a time, after controlling for age and sex. The HAPSTAT [27–29] and HAPLO.STATS [33, 34] programs

**Table 3** Score tests, presenting *p*-values, for urokinase polymorphisms in healthy controls and patients with ALI

Test	2065 rs1916341	3855 rs2227562	3955 rs2227564	4625 rs2227566	5634 rs2227571	7358 rs4065
Chi-square	0.447	0.622	0.405	0.727	0.460	0.597
Entropy <sup>a</sup>	0.439	0.616	0.395	0.723	0.452	0.590
Exponential <sup>a</sup>	0.439	0.614	0.394	0.723	0.453	0.590
Polynomial <sup>a</sup>	0.439	0.614	0.394	0.723	0.453	0.590
Sigmoid <sup>a</sup>	0.439	0.614	0.393	0.723	0.452	0.590
Gaussian <sup>a</sup>	0.439	0.614	0.393	0.723	0.452	0.590
Reciprocal <sup>a</sup>	0.440	0.617	0.398	0.723	0.453	0.591

<sup>a</sup> Score tests are presented as proposed in [28, 29]; The *p*-values presented in the table are not adjusted for multiple comparisons

**Table 4** Urokinase haplotypes and their frequencies in patients with ALI and healthy controls. Haplotypes with frequencies greater than 0.01 are shown

2065 rs1916341	3855 rs2227562	3955 rs2227564	4625 rs2227566	5634 rs2227571	7358 rs4065	Haplotype frequency: ALI	Haplotype frequency: controls
A	G	C	T	C	T	0.020	0.020
C	A	C	C	C	C	0.144	0.148
C	G	C	C	C	C	0.042	0.029
C	G	C	C	C	T	0.010	0.011
C	G	T	C	C	C	0.241	0.254
A	G	C	T	T	T	0.531	0.514

were used for haplotype analysis. Neither analysis found significant association between any haplotype and risk for ALI.

#### Associations of urokinase polymorphisms and haplotypes with outcome from acute lung injury

In order to test the association of genetic markers with outcome from ALI, an analysis of the patients with infection-induced ALI was undertaken. Of note, SNPs 1 (rs1916341) and 4 (rs2227566) were found to be in complete linkage disequilibrium (LD), and were in very strong LD with SNP 5 (rs2227571). Therefore, the results for SNPs 1 and 4 are identical in all single-marker analyses (Table 5). Unlike age, which was significantly associated with 60-day survival among patients with ALI, as well as ventilator-free days and the presence of fluid-unresponsive shock, there was no statistically significant association

between any of the six single polymorphisms examined and 60-day survival from ALI, using either 60-day mortality as a binary response or as a time-to-event survival response. This lack of association was present with or without the inclusion of other covariates in the analysis. In contrast, when haplotype analyses were performed using a dominant-effect model, the haplotype CGCCCC was found to be significantly associated with 60-day mortality, either compared to the most frequent haplotype (AGCTTT) in a joint-effect model ( $p = 0.033$ ) (Table 6), or when compared to all other haplotypes in a separate-effect model ( $p = 0.021$ ), after controlling for age and sex. The association between the CGCCCC haplotype and mortality at 60 days was also weakly supported by data from a survival analysis ( $p = 0.077$ ).

Measurement of ventilator-free days is a clinically relevant parameter in patients with ALI, providing an assessment of the duration of requirement for mechanical ventilation. As was the case for 60-day mortality, no

**Table 5** Score tests, presenting *p*-values, for urokinase polymorphisms and 60-day mortality in patients with ALI

	2065 rs1916341	3855 rs2227562	3955 rs2227564	4625 rs2227566	5634 rs2227571	7358 rs4065
Chi-square	0.679	0.709	0.576	0.679	0.571	0.917
Entropy <sup>a</sup>	0.632	0.670	0.522	0.632	0.512	0.904
Exponential <sup>a</sup>	0.632	0.673	0.526	0.632	0.512	0.904
Polynomial <sup>a</sup>	0.632	0.673	0.526	0.632	0.512	0.904
Sigmoid <sup>a</sup>	0.632	0.673	0.527	0.632	0.512	0.904
Gaussian <sup>a</sup>	0.632	0.673	0.526	0.632	0.512	0.904
Reciprocal <sup>a</sup>	0.633	0.667	0.520	0.633	0.513	0.904

<sup>a</sup> Score tests are presented as proposed in [28, 29]; The *p*-values presented in the table are not adjusted for multiple comparisons

**Table 6** Analysis of urokinase haplotypes and 60-day mortality in patients with ALI

Haplotype	Estimate	Standard error	<i>t</i> Statistic	<i>p</i> -value
AGCTCT	0.243	0.740	0.328	0.743
CACCCC	-0.105	0.359	-0.292	0.770
CGCCCC	1.004	0.467	2.151	0.033
CGTCCC	-0.082	0.318	-0.259	0.796
Rare haplotypes <sup>a</sup>	0.236	0.724	0.326	0.745
Age	0.030	0.010	3.01	0.0029

<sup>a</sup> Haplotypes with frequencies less than 0.01 were grouped together; Values presented are derived from the HAPLO.STATS program under a dominant effects model; Haplotype AGCTTT, which has the largest frequency, was used as baseline

individual urokinase polymorphism was significantly associated with ventilator-free days. In contrast, there was a statistically significant relationship between the CGCCCC haplotype and ventilator-free days, either compared to the most frequent haplotype (AGCTTT) in the joint-effect model ( $p < 0.001$ ), or compared to all other haplotypes in the separate-effect model ( $p = 0.047$ ), after controlling for age and sex.

Days of shock were assessed both by sequential daily measurement of vital signs, with a systolic blood pressure of less than 90 despite adequate fluid administration, being considered as shock, or by the Brussels system, with requirement for vasopressor support to maintain normotension being required for scoring as shock. However, using either of these criteria, there were no statistically significant relationships between shock and any single polymorphism or haplotype. In particular, for the CGCCCC haplotype, which was significantly associated with 60-day mortality and ventilator-free days, the *p*-values for association with shock were  $> 0.11$  using the Brussels scoring system and  $> 0.26$  using vital signs, in all the analyses performed, even after controlling for the covariates of sex and age.

In our haplotype analyses, we used the joint-effect model as our primary analysis and the separate-effect model as confirmation. Since SNP 5 showed marginal departure from HWE in the pooled data, to be prudent, we also conducted analyses with SNP5 excluded. The results were similar to these presented above (data not shown).

## Discussion

In this study, a significant relationship was demonstrated between a specific urokinase haplotype, CGCCCC, and increased 60-day mortality as well as requirement for prolonged mechanical ventilation among patients with ALI. This association between the CGCCCC haplotype and worse outcome from ALI was present both when the haplotype was considered alone and also in multivariate analysis, when both age and sex were included as covariates. While the CGCCCC urokinase haplotype was associated with outcome from ALI, there was no apparent relationship between this haplotype and susceptibility to ALI,

with no significant differences in the frequencies of the haplotype in healthy controls and patients with ALI. Such findings indicate that while genetic alterations in urokinase contribute to outcome in ALI, urokinase haplotypes do not constitute a risk factor for this life-threatening condition.

There is only limited information about urokinase polymorphisms and inflammatory conditions. In the present study, we examined a series of tagged and common polymorphisms that were previously found in the Seattle SNPs database to characterize all haplotypes present in more than 2.2% of the European-American population. The six haplotypes identified in the present study encompass more than 98% of the subjects included in the control and ALI populations examined, providing a comprehensive assessment of the relationships between genetic variation in urokinase and ALI.

While there have been reports of associations between the nonsynonymous C → T polymorphism rs2227564 within intron 7 and Alzheimer's disease, type 1 diabetes, and colonic cancer, this has not been a universal finding [19, 20, 22, 23, 36]. Although previous studies have shown associations between polymorphisms in proteins related to coagulation and mortality associated with sepsis [37, 38], there are no data on the relationship between urokinase polymorphisms and outcome from critical illness. In the present study, no association was found between specific urokinase polymorphisms and either the incidence or the outcome of ALI.

In addition to its well-characterized fibrinolytic activity, urokinase has additional properties that are likely to make it a relevant mediator of acute inflammatory processes, such as ALI. In particular, the high concentrations of urokinase present in the lungs after endotoxin exposure [8, 39], during pneumonia [9, 40, 41], and with ALI [3, 12, 14, 42–45], even if lacking fibrinolytic activity because of association with PAI-1, may still contribute to the accumulation of large numbers of activated neutrophils which appear to play an important role in the pathophysiology of ALI. Urokinase, through interactions of its growth factor domain with the classic uPA receptor (uPAR), has potent chemotactic effects on neutrophils [46–49]. The kringle domain of urokinase has recently been shown to have proinflammatory activity, enhancing neutrophil release of cytokines relevant to ALI,

such as TNF- $\alpha$  and IL-1 $\alpha$  [17]. In murine models, antibodies to the urokinase kringle domain diminish the severity of LPS-induced ALI [16]. Even though circulating and pulmonary levels of PAI-1 are elevated in ALI, resulting in inhibition of the fibrinolytic properties of urokinase, PAI-1 binds to the urokinase protease domain, without apparent interaction with either the kringle or growth factor domains [14, 50, 51]. Therefore, interactions between PAI-1 and urokinase would not be expected to modify effects leading to neutrophil activation or accumulation in the lungs that are modulated by urokinase domains distinct from the protease domain.

To minimize confounding variables, such as racial admixture and heterogeneous etiologies for ALI, the present study focused on European-American patients whose primary etiology for ALI was infection. Management of mechanical ventilation was standardized for these patients as a result of being enrolled in studies in which protocols were used with levels of compliance exceeding 98%. In particular, all patients included in this study had been enrolled in NIH ARDS Network studies and received low tidal volume ventilation, presently considered the optimal ventilatory therapy for ALI [1]. However, because of the

restricted patient population included in this analysis, it is presently unknown whether the association between the CGCCCC urokinase haplotype and outcome from ALI is present in other groups of patients with ALI or is limited only to European-Americans with infection-associated ALI. An additional limitation of the present study is that neither urokinase levels nor activities were determined, so there is no information concerning the functional significance of the urokinase haplotype identified as being associated with worse outcome from ALI.

The findings of the present study, showing that genetic alterations in urokinase are associated with higher mortality and the need for prolonged mechanical ventilation in patients with ALI, provide an additional indication of the importance of coagulation and its regulation in contributing to the pathophysiology of ALI and other organ system dysfunction resulting from severe infection. In addition, these results not only may have diagnostic utility in identifying ALI patients at increased risk of worse outcomes, but also may have additional future implications since they may facilitate the targeting of therapies specific for modulating injurious effects of urokinase or related molecules to appropriate patient populations.

## References

- Bernard GR (2005) Acute respiratory distress syndrome: a historical perspective. *Am J Respir Crit Care Med* 172:798–806
- Matthay MA, Zimmerman GA (2005) Acute lung injury and the acute respiratory distress syndrome: four decades of inquiry into pathogenesis and rational management. *Am J Respir Cell Mol Biol* 33:319–327
- Ware LB, Camerer E, Welty-Wolf K, Schultz MJ, Matthay MA (2006) Bench to bedside: targeting coagulation and fibrinolysis in acute lung injury. *Am J Physiol Lung Cell Mol Physiol* 291:L307–L311
- Ware LB, Matthay MA, Parsons PE, Thompson BT, Januzzi JL, Eisner MD (2007) Pathogenetic and prognostic significance of altered coagulation and fibrinolysis in acute lung injury/acute respiratory distress syndrome. *Crit Care Med* 35:1821–1828
- Blasi F, Carmeliet P (2002) uPAR: a versatile signalling orchestrator. *Nat Rev Mol Cell Biol* 3:932–943
- Idell S (2002) Endothelium and disordered fibrin turnover in the injured lung: newly recognized pathways. *Crit Care Med* 30:S274–S280
- Abraham E, Gyetko MR, Kuhn K, Arcaroli J, Strassheim D, Park JS, Shetty S, Idell S (2003) Urokinase-type plasminogen activator potentiates lipopolysaccharide-induced neutrophil activation. *J Immunol* 170:5644–5651
- van der Poll T, Levi M, Nick JA, Abraham E (2005) Activated protein C inhibits local coagulation after intrapulmonary delivery of endotoxin in humans. *Am J Respir Crit Care Med* 171:1125–1128
- Levi M, Schultz MJ, Rijneveld AW, van der Poll T (2003) Bronchoalveolar coagulation and fibrinolysis in endotoxemia and pneumonia. *Crit Care Med* 31:S238–S242
- Choi G, Schultz MJ, van Till JW, Bresser P, van der Zee JS, Boermeester MA, Levi M, van der Poll T (2004) Disturbed alveolar fibrin turnover during pneumonia is restricted to the site of infection. *Eur Respir J* 24:786–789
- Schultz MJ, Haitsma JJ, Zhang H, Slutsky AS (2006) Pulmonary coagulopathy as a new target in therapeutic studies of acute lung injury or pneumonia: a review. *Crit Care Med* 34:871–877
- Groeneveld AB, Kindt I, Raijmakers PG, Hack CE, Thijs LG (1997) Systemic coagulation and fibrinolysis in patients with or at risk for the adult respiratory distress syndrome. *Thromb Haemost* 78:1444–1449
- Prabhakaran P, Ware LB, White KE, Cross MT, Matthay MA, Olman MA (2003) Elevated levels of plasminogen activator inhibitor-1 in pulmonary edema fluid are associated with mortality in acute lung injury. *Am J Physiol Lung Cell Mol Physiol* 285:L20–L28
- Idell S (2003) Coagulation, fibrinolysis, and fibrin deposition in acute lung injury. *Crit Care Med* 31:S213–S220
- El Solh AA, Bhora M, Pineda L, Aquilina A, Abbetessa L, Berbary E (2006) Alveolar plasminogen activator inhibitor-1 predicts ARDS in aspiration pneumonitis. *Intensive Care Med* 32:110–115
- Wang XQ, Bdeir K, Yarovoi S, Cines DB, Fang W, Abraham E (2006) Involvement of the urokinase kringle domain in lipopolysaccharide-induced acute lung injury. *J Immunol* 177:5550–5557

17. Kwak S-H, Mitra S, Bdeir K, Strassheim D, Park JS, Kim JK, Idell S, Cines DL, Abraham E (2005) The kringle domain of urokinase-type plasminogen activator potentiates LPS-induced neutrophil activation through interaction with  $\{\alpha\}V\{\beta\}3$  integrins. *J Leukoc Biol* 78:937–945
18. Irigoyen JP, Munoz-Canoves P, Montero L, Koziczak M, Nagamine Y (1999) The plasminogen activator system: biology and regulation. *Cell Mol Life Sci* 56:104–132
19. Przybylowska K, Smolarczyk K, Kulig A, Romanowicz-Makowska H, Dziki A, Ulanska J, Pander B, Blasiak J (2002). Antigen levels of the urokinase-type plasminogen activator and its gene polymorphisms in colorectal cancer. *Cancer Lett* 181:23–30
20. Majsterek I, Przybylowska K, Mlynarski W, Drzewoski J, Blasiak J (2005) Distribution of C→T and T→C polymorphisms of the urokinase-type plasminogen activator gene in children with type 1 diabetes mellitus and insulin resistance. *Clin Exp Med* 4:202–206
21. Ertekin-Taner N, Ronald J, Feuk L, Prince J, Tucker M, Younkin L, Hella M, Jain S, Hackett A, Scanlin L, Kelly J, Kihiko-Ehman M, Neltner M, Hersh L, Kindy M, Markesbery W, Hutton M, de Andrade M, Petersen RC, Graff-Radford N, Estus S, Brookes AJ, Younkin SG (2005) Elevated amyloid beta protein (Aβ42) and late onset Alzheimer's disease are associated with single nucleotide polymorphisms in the urokinase-type plasminogen activator gene. *Hum Mol Genet* 14:447–460
22. Riemenschneider M, Konta L, Friedrich P, Schwarz S, Taddei K, Neff F, Padovani A, Kolsch H, Laws SM, Klopp N, Bickeboller H, Wagenpfeil S, Mueller JC, Rosenberger A, Diehl-Schmid J, Archetti S, Lautenschlager N, Borroni B, Muller U, Illig T, Heun R, Egensperger R, Schlegel J, Forstl H, Martins RN, Kurz A (2006) A functional polymorphism within plasminogen activator urokinase (PLAU) is associated with Alzheimer's disease. *Hum Mol Genet* 15:2446–2456
23. Huang CM, Chen CL, Tsai JJ, Tsai CH, Tsai FJ (2004) Association between urokinase gene 3'-UTR T/C polymorphism and Chinese patients with rheumatoid arthritis in Taiwan. *Clin Exp Rheumatol* 22:219–222
24. Begin P, Tremblay K, Daley D, Lemire M, Claveau S, Salesse C, Kacel S, Montpetit A, Becker A, Chan-Yeung M, Kozyrskyj AL, Hudson TJ, Laprise C (2007) Association of urokinase-type plasminogen activator with asthma and atopy. *Am J Respir Crit Care Med* 175:1109–1116
25. Brower RG, Lanken PN, MacIntyre N, Matthay MA, Morris A, Ancukiewicz M, Schoenfeld D, Thompson BT (2004) Higher versus lower positive end-expiratory pressures in patients with the acute respiratory distress syndrome. *N Engl J Med* 351:327–336
26. Scheet P, Stephens M (2006) A fast and flexible statistical model for large-scale population genotype data: applications to inferring missing genotypes and haplotypic phase. *Am J Hum Genet* 78:629–644
27. Zhao J, Jin L, Xiong M (2006) Test for interaction between two unlinked loci. *Am J Hum Genet* 79:831–845
28. Zhao J, Jin L, Xiong M (2006) Non-linear tests for genomewide association studies. *Genetics* 174:1529–1538
29. Zhao JY, Xiong MM, Huang W, Wang H, Zuo J, Wu GD, Chen Z, Qiang BQ, Zhang ML, Chen JL, Ding W, Yuan WT, Xu HY, Jin L, Li YX, Sun Q, Liu QY, Boerwinkel E, Fang FD (2005) An autosomal genomic scan for loci linked to type 2 diabetes in northern Han Chinese. *J Mol Med* 83:209–215
30. Lin DY, Zeng D, Millikan R (2005) Maximum likelihood estimation of haplotype effects and haplotype-environment interactions in association studies. *Genet Epidemiol* 29:299–312
31. Zeng D, Lin DY (2005) Estimating haplotype-disease associations with pooled genotype data. *Genet Epidemiol* 28:70–82
32. Zeng D, Lin DY, Avery CL, North KE, Bray MS (2006) Efficient semi-parametric estimation of haplotype-disease associations in case-cohort and nested case-control studies. *Biostatistics* 7:486–502
33. Schaid DJ, Rowland CM, Tines DE, Jacobson RM, Poland GA (2002) Score tests for association between traits and haplotypes when linkage phase is ambiguous. *Am J Hum Genet* 70:425–434
34. Lake SL, Lyon H, Tantisira K, Silverman EK, Weiss ST, Laird NM, Schaid DJ (2003) Estimation and tests of haplotype-environment interaction when linkage phase is ambiguous. *Hum Hered* 55:56–65
35. Guo SW, Thompson EA (1992) Performing the exact test of Hardy-Weinberg proportion for multiple alleles. *Biometrics* 48:361–372
36. Myers AJ, Marshall H, Holmans P, Compton D, Crook RJ, Mander AP, Nowotny P, Smemo S, Dunstan M, Jehu L, Wang JC, Hamshere M, Morris JC, Norton J, Chakraverty S, Tunstall N, Lovestone S, Petersen R, O'Donovan M, Jones L, Williams J, Owen MJ, Hardy J, Goate A (2004) Variation in the urokinase-plasminogen activator gene does not explain the chromosome 10 linkage signal for late onset AD. *Am J Med Genet B Neuropsychiatr Genet* 124:29–37
37. Manocha S, Russell JA, Sutherland AM, Wattanathum A, Walley KR (2007) Fibrinogen-beta gene haplotype is associated with mortality in sepsis. *J Infect* 54:572–577
38. Arcaroli J, Fessler MB, Abraham E (2005) Genetic polymorphisms and sepsis. *Shock* 24:300–312
39. Maris NA, de Vos AF, Bresser P, van der Zee JS, Meijers JC, Lijnen HR, Levi M, Jansen HM, van der Poll T (2005) Activation of coagulation and inhibition of fibrinolysis in the lung after inhalation of lipopolysaccharide by healthy volunteers. *Thromb Haemost* 93:1036–1040
40. Schultz MJ, Millo J, Levi M, Hack CE, Weverling GJ, Garrard CS, van der Poll T (2004) Local activation of coagulation and inhibition of fibrinolysis in the lung during ventilator associated pneumonia. *Thorax* 59:130–135
41. Rijneveld AW, Weijer S, Bresser P, Florquin S, Vlasuk GP, Rote WE, Spek CA, Reitsma PH, van der Zee JS, Levi M, van der Poll T (2006) Local activation of the tissue factor-factor VIIa pathway in patients with pneumonia and the effect of inhibition of this pathway in murine pneumococcal pneumonia. *Crit Care Med* 34:1725–1730
42. Idell S (1994) Extravascular coagulation and fibrin deposition in acute lung injury. *New Horiz* 2:566–574
43. Abraham E (2000) Coagulation abnormalities in acute lung injury and sepsis. *Am J Respir Cell Mol Biol* 22:401–404
44. Gunther A, Mosavi P, Heinemann S, Ruppert C, Muth H, Markart P, Griminger F, Walmrath D, Temmesfeld-Wollbrück B, Seeger W (2000) Alveolar fibrin formation caused by enhanced procoagulant and depressed fibrinolytic capacities in severe pneumonia. Comparison with the acute respiratory distress syndrome. *Am J Respir Crit Care Med* 161:454–462
45. Ware LB, Bastarache JA, Wang L (2005) Coagulation and fibrinolysis in human acute lung injury: new therapeutic targets? *Keio J Med* 54:142–149

- 
46. Gyetko MR, Sitrin RG, Fuller JA, Todd RF, Petty H, Standiford TJ (1995) Function of the urokinase receptor (CD87) in neutrophil chemotaxis. *J Leukoc Biol* 58:533–538
  47. Resnati M, Guttinger M, Valcamonica S, Sidenius N, Blasi F, Fazioli F (1996) Proteolytic cleavage of the urokinase receptor substitutes for the agonist-induced chemotactic effect. *Embo J* 15:1572–1582
  48. Mondino A, Resnati M, Blasi F (1999) Structure and function of the urokinase receptor. *Thromb Haemost [Suppl 1]* 82:19–22
  49. Sitrin RG, Pan PM, Blackwood RA, Huang J, Petty HR (2001) Cutting edge: evidence for a signaling partnership between urokinase receptors (CD87) and L-selectin (CD62L) in human polymorphonuclear neutrophils. *J Immunol* 166:4822–4825
  50. Shetty S, Bdeir K, Cines DB, Idell S (2003) Induction of plasminogen activator inhibitor-1 by urokinase in lung epithelial cells. *J Biol Chem* 278:18124–18131
  51. Reuning U, Magdolen V, Hapke S, Schmitt M (2003) Molecular and functional interdependence of the urokinase-type plasminogen activator system with integrins. *Biol Chem* 384:1119–1131