

Dietary Fat, Fiber, and Carbohydrate Intake and Endogenous Hormone Levels in Premenopausal Women

Xiaohui Cui · Bernard Rosner · Walter C. Willett · Susan E. Hankinson

Published online: 23 November 2010
© Springer Science+Business Media, LLC 2010

Abstract The authors conducted a cross-sectional study to investigate the associations of fat, fiber, and carbohydrate intake with endogenous estrogen, androgen, and insulin-like growth factor (IGF) levels among 595 premenopausal women. Overall, no significant associations were found between dietary intake of these macronutrients and plasma sex steroid hormone levels. Dietary fat intake was inversely associated with IGF-I and IGF-binding protein 3 (IGFBP-3) levels. When substituting 5% of energy from total fat for the equivalent amount of energy from carbohydrate or protein intake, the plasma levels of IGF-I and IGFBP-3 were 2.8% (95% confidence interval [CI] 0.3, 5.3) and 1.6% (95% CI 0.4, 2.8) lower, respectively. Animal fat, saturated fat, and monounsaturated fat intakes also were inversely associated with IGFBP-3 levels ($P < 0.05$). Carbohydrates were positively associated with plasma IGF-I level. When substituting 5% of energy from carbohydrates for the equivalent amount of energy from fat or protein intake, the plasma IGF-I level was 2.0% (95% CI 0.1,

3.9%) higher. No independent associations between fiber intake and hormone levels were observed. The results suggest that a low-fat/high-fiber or carbohydrate diet is not associated with endogenous levels of sex steroid hormones, but it may modestly increase IGF-I and IGFBP-3 levels among premenopausal women.

Keywords Gonadal steroid hormones · Insulin-like growth factor I · Insulin-like growth factor binding protein 3 · Dietary fats · Dietary fiber · Dietary carbohydrates

Abbreviations

BMI	Body mass index
CI	Confidence interval
DHEA	Dehydroepiandrosterone
DHEAS	DHEA sulfate
FFQ	Food frequency questionnaire
IGF	Insulin-like growth factor
IGFBP	IGF-binding protein
NHS	Nurses' health study
SHBG	Sex hormone binding globulin

X. Cui (✉) · B. Rosner · W. C. Willett · S. E. Hankinson
Channing Laboratory, Department of Medicine,
Brigham and Women's Hospital and Harvard Medical School,
181 Longwood Avenue,
Boston 02115 MA, USA
e-mail: nhxxc@channing.harvard.edu

B. Rosner
Department of Biostatistics, Harvard School of Public Health,
Boston, MA, USA

W. C. Willett · S. E. Hankinson
Department of Epidemiology, Harvard School of Public Health,
Boston, MA, USA

W. C. Willett
Department of Nutrition, Harvard School of Public Health,
Boston, MA, USA

Endogenous sex hormone levels play an important role in the etiology of endometrial [1–4], breast [5–12], and ovarian [13, 14] cancers. Insulin-like growth factors (IGFs) are important in regulating cell proliferation, differentiation, apoptosis, and transformation, and IGF binding proteins (IGFBPs) can enhance or inhibit the effect of IGFs [15–18]. Circulating levels of IGF-I and IGFBP-3 have been associated with risk of prostate [19–23], breast [24–28], colorectal [29, 30], and lung cancer [22, 25, 31–33] in some but not all studies.

Although some epidemiological studies have suggested that dietary fat or fiber may be related to the risk of

endometrial [34–37], ovarian[38–41], and colon[42–44] cancers, the evidence has not been consistent. A primary mechanism by which these factors might influence risk is through the modulation of hormone levels. A low-fat/high-fiber diet is thought to reduce plasma estrogen levels by increasing the fecal excretion of estrogens [45, 46]. Energy and protein intakes appear to increase IGF levels [47], but little is known about the effect of dietary fat, fiber, or carbohydrates on the IGF axis, and results from previous studies are inconsistent[48–51].

Data from dietary fat and fiber in relation to sex hormone and IGF levels among premenopausal women are particularly limited. To investigate the relationship of dietary fat and fiber intake with endogenous hormone levels, we conducted a cross-sectional study among premenopausal participants in the Nurses' Health Study (NHS) II cohort, for whom blood samples were timed according to the menstrual cycle.

Materials and Methods

Study Population

The NHS II is a prospective cohort study established in 1989, when 116,671 registered nurses in 14 states, 25–42 years of age at that time, completed a baseline questionnaire about their medical histories and lifestyles. Subsequent questionnaires requesting updated information on risk factors and medical events have been mailed every 2 years. The follow-up rate in this cohort between 1989 and 2003 was over 95% of potential person-years.

From 1996 to 1999, the NHSII participants meeting the following criteria were invited to send us a blood sample, timed within the menstrual cycle: (a) still having menstrual periods, (b) had not used oral contraceptives or other hormones in the past 6 months, (c) had not been pregnant or lactating in the previous 6 months, and (d) had no prior cancer diagnosis. These women provided an initial 15-ml blood sample drawn on the third to fifth day of their menstrual cycle (follicular blood draw) and a second 30-ml blood sample drawn 7–9 days before the anticipated start of their next cycle (luteal blood draw). To accurately determine when in the luteal phase the second blood was collected, women were asked to mail us a follow-up postcard documenting when their next menstrual cycle began. A total of 19,092 women provided timed blood samples. The women who did not meet the criteria above or declined to give a timed blood sample were asked to provide a one-time 30-ml untimed blood sample. In total, 11,090 women provided the untimed samples. Menopausal status was ascertained via questionnaire at the time of blood collection.

Follicular samples were placed in a refrigerator for 8–24 h and then aliquoted by the participant and frozen until the luteal blood collection. Both the follicular and luteal blood samples were shipped via overnight courier and with an ice-pack to our laboratory where the luteal blood sample was processed and aliquoted into plasma, red blood cell, and white blood cell components. The untimed samples were processed and stored in a similar manner. We previously showed that hormone levels are stable in whole blood processed in this manner [52].

Women included in this analysis are the premenopausal controls of a nested case-control study of breast cancer ($n=482$) (detailed in [53–56]) and the subjects of a previously published study to assess hormone reproducibility over time in premenopausal women ($n=113$) [57]. We used the first blood sample provided in this study. Among these 595 women, 502 had both follicular and luteal samples, two had only luteal samples, and 93 had untimed samples.

Assessment of Intake of Fat, Fiber, and Carbohydrates

A semi-quantitative food frequency questionnaire (FFQ) was collected in 1991 and every 4 years since (1995, 1999, and 2003). The questionnaire queried average intake over the last year of 133 foods, with frequency of intake ranging from never or almost never to ≥ 6 times per day. The validity of FFQs similar to those used in the NHS II cohort has been described elsewhere [58–60]. In the NHS cohort, the correlation coefficients between energy-adjusted total fat, fiber, and carbohydrates from FFQs and the average of four 1-week diet records at 3-month intervals were 0.53, 0.58, and 0.61, respectively. For saturated and polyunsaturated fats, the coefficients were 0.59 and 0.48 [59]. A previous study done in the NHS cohort also showed that plasma triglyceride levels across the categories of energy intake were comparable to those found in an intervention study, providing objective evidence that the FFQs used in this study are sensitive to dietary fat [61].

The calculation of dietary intake was based on the USDA database [62]. We evaluated total fat, animal fat, vegetable fat, specific fatty acids (saturated fat, monounsaturated fat, polyunsaturated fat, and trans fat), total fiber, fiber from cereals, vegetables, fruits, cruciferous vegetables, and legumes, and carbohydrates. Average dietary intake, calculated using data from the 1995 and 1999 FFQs (the dietary questionnaires collected closest in time to the blood draw), was used in the analyses to reduce measurement error.

Laboratory Assays

Samples were assayed in three batches. Estradiol, estrone, estrone sulfate, and testosterone were measured at Quest

Diagnostics (San Juan Capistrano, CA) by radioimmunoassay following extraction and celite column chromatography. Progesterone, androstenedione, sex hormone binding globulin (SHBG), dehydroepiandrosterone (DHEA), and DHEA sulfate (DHEAS) were measured at the Royal Marsden Hospital (London, UK) by radioimmunoassay (SHBG was measured using IMMULITE Diagnostic Products), and IGF-I and IGFBP-3 were measured at the Lady Davis Research Institute, M. Pollak's Laboratory (Montreal, CA) by ELISA (Diagnostic Systems Laboratory, TX). Different hormones were measured in follicular, luteal, and untimed samples. (Appendix 1). In each batch, we included replicate plasma samples to assess laboratory precision. All between-batch coefficients of variation were less than 10%, except for follicular estradiol (12%), SHBG (13%), and progesterone (14%).

Data Analyses

Since estrogen levels fluctuate during the menstrual cycle, follicular and luteal hormone levels were analyzed separately, and only women with timed samples were included in these analyses. Because levels of SHBG, androgens, and IGFs fluctuate only modestly [57], we used both timed and untimed samples in these analyses. For women with both follicular and luteal hormone values (i.e., for SHBG, androstenedione, and testosterone), we used their average hormone level. We identified statistical outliers based on the generalized extreme studentized deviate many-outlier detection approach [63]. Two to four observations were deleted for luteal estradiol, luteal free estradiol, luteal estrone, and luteal progesterone. Also, in the analyses of luteal estrogens and progesterone, anovulatory women (luteal progesterone <400 ng/dl) were excluded ($n=53$).

The associations between dietary fiber and percentage of energy from fat and carbohydrate intakes and hormone levels were assessed by linear regression. Statistical analyses were performed with SAS software (SAS Institute, Cary, NC) using PROC MIXED procedure. The independent variables, types of dietary fat, fiber, and carbohydrates, were continuous. The robust variance was used to ensure valid inference even if the regression residuals were not normally distributed. The difference in plasma hormone levels was modeled on the natural logarithm scale. Solving $(1 - e^{-\beta\Delta}) \times 100$, where β is the estimated regression slope and Δ is the specified incremental difference in macronutrient intake, represents the percentage difference in that hormone level. The mean hormone levels across the quintiles of nutrients intake were computed to show the absolute difference in plasma hormone levels.

The covariates included in the regression model were age, fasting status, laboratory batch, luteal day (for luteal

levels), time of day of blood draw, body mass index (BMI) at age 18, BMI at blood collection, smoking status, parity, and level of physical activity in 1997, total energy and alcohol intakes (average of 1995 and 1999 FFQs), age at menarche, and height (see Table 3 for variable definitions). In the multivariate models, the coefficients for fat and carbohydrates can be interpreted as substitution of a percentage of energy from fat/carbohydrates for an equal percentage from other sources of energy.

Results

After exclusions, 595 premenopausal women were included in the analysis. The mean age at blood draw was 43.4, and mean BMI was 25.2 kg/m² in 1997 (see Table 1). Of the women included in the analysis, 69.9% never smoked and ever smokers smoked an average of 10.1 pack-years. The average intake of total energy per day was 1,842 kcal, 29.1% of which was from fat, 51.3% from carbohydrates, and 18.1% from protein. On average, these women consumed 20 g of dietary fiber and 4.2 g of alcohol. Median plasma hormone levels and their 10th–90th ranges are shown in Table 2.

In Table 3, we present the geometric means of hormones across categories of percentage energy from fat adjusted to the mean levels of the covariates, as well as the estimated percentage difference in plasma hormone levels from substituting 5% of energy from total fat for the equivalent amount of energy from carbohydrate or protein. Unadjusted and multivariable adjusted models had similar results, so only results of multivariable models are shown. Overall, we found no significant association between

Table 1 Characteristics of women included in the study

Characteristic	Mean (SD)
Age at blood draw, years	43.4 (3.9)
BMI at 18, kg/m ²	21.0 (2.7)
BMI in 1997, kg/m ²	25.2 (5.7)
Parity among parous women	2.3 (0.9)
Ever-smoked,%	30.1 (45.9)
Pack-years among smokers	10.1 (7.7)
Energy intake, kcal/day ^a	1842 (496)
Alcohol intake, g/day ^a	4.2 (6.6)
Fiber intake energy-adjusted, g/day ^a	20.2 (5.8)
Percentage of energy from protein,% ^a	18.1 (2.8)
Percentage of energy from fat,% ^a	29.1 (5.6)
Percentage of energy from carbohydrate,% ^a	51.3 (6.8)

^a Average of 1995 and 1999 reported intake

Table 2 Plasma hormone levels

	Number	Median	10th–90th Percentile
Estradiol (pg/ml)			
Follicular	460	46.0	21.0, 98.0
Luteal	408	125	79.0, 203
Free estradiol (pg/ml)			
Follicular	436	0.6	0.3, 1.1
Luteal	399	1.6	0.9, 2.6
Estrone (pg/ml)			
Follicular	464	40.0	25.0, 63.0
Luteal	442	80.0	50.0, 124
Estrone sulfate (pg/ml)			
Follicular	449	667	291, 1514
Luteal	402	1478	573, 3,235
Progesterone (ng/ml)	447	1509	740, 2,639
SHBG ^a (nmol/l)	593	63.1	31.5, 109
Androstenedione ^a (ng/ml)	593	106	63.0, 176
Testosterone ^a (ng/ml)	588	24.0	14.5, 37.0
Free testosterone ^a (ng/ml)	588	0.2	0.1, 0.4
DHEA ^b (ng/dl)	477	642	344, 1133
DHEA sulfate ^b (μg/dl)	477	79.5	39.1, 142
IGF-I ^b (ng/ml)	595	245	147, 349
IGFBP-3 ^b (ng/ml)	595	4,769	3,297, 5,911

^a Average of follicular and luteal timed values for women with both

^b Concatenation of luteal values and untimed values

percentage energy from total fat and plasma levels of estrogens, progesterone, SHBG, or androgens. Although there was no linear relationship between total fat and plasma follicular levels, levels of follicular estradiol were significantly higher among the first category (percentage of energy from total fat $\leq 20\%$) versus all other categories ($P=0.01$). A higher percentage of energy from total fat was related to modestly lower levels of IGF-I and IGFBP-3, but not the IGF-I/IGFBP-3 ratio. For a 5% increase in energy from total fat intake, plasma IGF-I levels were 2.8% lower (95% confidence interval [CI] 0.3%, 5.3%) and plasma IGFBP-3 levels 1.6% lower (95% CI 0.4%, 2.8%). We also found that IGF-I levels among those with $\leq 20\%$ of energy from total fat were significantly higher than levels in all other categories ($P<0.01$).

We also estimated the percentage difference in plasma hormone levels from substituting 1% of energy from specific types of fat for the equivalent amount of energy from other sources as replacement (Table 4). As observed with total fat, most hormone levels were not related with any fat type. Saturated fat was inversely associated with IGF-I levels (1.2% decrease; 95% CI 0, 2.4%). IGFBP-3 also was inversely associated with intakes of animal fat,

saturated fat, and monounsaturated fat. For a 1% increase in energy from animal fat, saturated fat, and monounsaturated fat, plasma IGFBP-3 levels decreased 0.4% (95% CI 0.1, 0.7%), 0.8% (95% CI 0.2%, 1.4%), and 0.6% (95% CI 0.1%, 1.2%), respectively. Similarly, we did not find significant associations between intake of fat types and the IGF-I/IGFBP-3 ratio.

Overall, the percentage of energy from carbohydrates was not related to estrogen, progesterone, androgen, or SHBG levels (Table 5). We observed a significant positive relationship between percentage of energy from carbohydrates and IGF-I levels. With substitution of 5% of energy from carbohydrates for the equivalent amount of energy from fat or protein, plasma IGF-I levels increased by 2.0% (95% CI 0.1, 3.9%).

We also observed no substantial difference in mean hormone values across categories of total fiber intake (Table 6). Similarly, we did not find significant relationships between levels of any hormone and total fiber intake, except for androstenedione. A 5-g increment of total fiber intake was related to a 3.0% decrease in androstenedione levels (95% CI 0.1%, 5.9%). Also, we did not find significant associations between plasma hormone levels

Table 3 Geometric means across categories of percentage of energy from total fat and estimated percentage difference in plasma hormone levels from substituting 5% of energy from total fat for the equivalent amount of energy from carbohydrate or protein intake

N (range)	% Energy from total fat					% Difference in hormone levels	
	≤20%	20.1–25%	25.1–30%	30.1–35%	>35%	Estimate	95% CI
	15–25	70–114	143–215	100–148	63–90		
	Geometric means						
Estradiol (pg/ml)							
Follicular	61.3	40.4	50.1	46.6	43.1	−1.1	−5.7, 3.2
Luteal	128	122	116	124	118	−0.1	−0.8, 0.5
Free estradiol (pg/ml)							
Follicular	0.7	0.5	0.6	0.6	0.5	−1.0	−5.5, 3.3
Luteal	1.6	1.5	1.5	1.5	1.4	−0.3	−1.0, 0.3
Estrone (pg/ml)							
Follicular	39.0	37.5	41.8	38.9	41.7	1.3	−1.9, 4.5
Luteal	75.4	75.8	74.6	75.2	75.4	−0.1	−0.7, 0.5
Estrone sulfate (pg/ml)							
Follicular	752	619	679	697	666	0.8	−4.5, 5.8
Luteal	1,336	1,401	1,332	1,322	1,368	−0.0	−1.2, 1.1
Progesterone (ng/ml)	1,229	1,369	1,350	1,484	1,176	−0.3	−1.0, 0.5
SHBG (nmol/l)	65.1	57.0	60.3	62.2	64.7	2.2	−1.0, 5.3
Androstenedione (ng/ml)	105.1	97.5	103.3	106.5	109.6	2.6	−0.4, 5.4
Testosterone (ng/ml)	24.1	21.8	23.8	23.4	24.6	1.6	−1.2, 4.3
Free testosterone (ng/ml)	0.2	0.2	0.2	0.2	0.2	0.1	−3.0, 3.1
DHEA (ng/dl)	587	626	645	650	634	1.5	−2.1, 4.9
DHEA sulfate (μg/dl)	66.1	75.4	77.9	78.0	77.2	1.3	−2.9, 5.3
IGF-I (ng/ml)	275	229	237	229	220	−2.8	−5.3, −0.3*
IGFBP-3 (ng/ml)	4,655	4,622	4,644	4,585	4,399	−1.6	−2.8, −0.4*
IGF-I/IGFBP-3	0.06	0.05	0.05	0.05	0.05	−1.1	−3.2, 0.9

^a Adjusted for age at blood draw (continuous), fasting status [<8 , ≥ 8 h], laboratory batch, day of luteal blood draw (for luteal hormones, 3–7, 8–28 days, missing/untimed sample), smoking status (pack years, never, 1–10, 11–20, 20+years), BMI in 1997 (continuous), BMI at age 18 (continuous), total energy intake (average of 1995 and 1999, continuous), alcohol intake (average of 1995 and 1999, continuous), parity (nulliparity, 1 and age at first birth <25 , 1 and age at first birth ≥ 25 , 2 and age at first birth <25 , 2 and age at first birth ≥ 25 , 3+ and age at first birth <25 , 3+ and age at first birth ≥ 25), age at menarche [<12 , 12, >12], and height (continuous).

* $P < 0.05$

and fiber from cereals, fiber from vegetables, fiber from fruits, fiber from cruciferous vegetables, and fiber from legumes (data not shown).

Discussion

In this cross-sectional study among premenopausal women, we did not find significant associations between dietary fat, fiber, or carbohydrates and plasma levels of estrogens, progesterone, androgens, or SHBG. However, we observed modest but significant inverse associations between the percentage of energy from dietary fat and

plasma levels of both IGF-I and IGFBP-3 and a positive association between the percentage of energy from carbohydrates and plasma IGF-I levels.

Results from the few prior studies of premenopausal women have been inconsistent, although differences in study design (e.g., the intervention used, hormones measured) complicate comparisons. In a meta-analysis of ten intervention studies conducted among premenopausal women, subjects changed from a high-fat (29–46% of fat in calories) to a low-fat (12–25% of fat calories) diet typically for 2 or 3 months, and overall a statistically significant 7.4% reduction in serum estradiol level was observed [64]. However, only one of the ten

Table 4 Estimated percentage difference in plasma hormone levels from substituting 1% of energy from types of fat for the equivalent amount of energy from other sources as replacement

	% Difference in hormone levels												
	Animal fat		Vegetable fat		Saturated fat		Monounsaturated fat		Polyunsaturated fat				
	Estimate	95% CI	Estimate	95% CI	Estimate	95% CI	Estimate	95% CI	Estimate	95% CI			
Estradiol (pg/ml)													
Follicular	-0.4	-1.5, 0.7	0.1	-1.4, 1.5	-0.1	-2.3, 2.0	-0.5	-2.6, 1.6	-1.8	-5.9, 2.3			
Luteal	0.1	-0.8, 0.9	-0.5	-1.5, 0.5	0.1	-1.7, 1.7	-0.4	-2.0, 1.1	-1.0	-4.0, 1.8			
Free estradiol (pg/ml)													
Follicular	0.1	-1.2, 1.0	-0.4	-1.8, 1.1	-0.3	-2.4, 1.7	-0.4	-2.6, 1.7	-1.1	-5.0, 2.7			
Luteal	-0.1	-1.0, 0.7	-0.8	-1.8, 0.3	-0.6	-2.3, 1.0	-0.9	-2.5, 0.6	-1.0	-4.1, 2.1			
Estrone (pg/ml)													
Follicular	0.2	-0.6, 1.0	0.4	-0.6, 1.4	0.4	-1.2, 2.0	0.6	-0.9, 2.1	1.1	-1.7, 3.8			
Luteal	-0.1	-0.8, 0.7	-0.1	-1.0, 0.8	-0.3	-1.8, 1.1	-0.0	-1.3, 1.2	-0.4	-3.2, 2.4			
Estrone sulfate (pg/ml)													
Follicular	0.1	-1.2, 1.3	0.3	-1.4, 2.0	-0.1	-2.5, 2.3	0.7	-1.7, 3.1	0.6	-4.2, 5.2			
Luteal	0.0	-1.5, 1.4	-0.0	-2.1, 2.1	-0.8	-3.6, 2.0	0.2	-2.5, 2.9	1.8	-4.0, 7.2			
Progesterone (ng/ml)	0.1	-0.8, 1.0	-0.9	-2.2, 0.4	-0.2	-2.0, 1.6	-0.8	-2.5, 0.9	-1.0	-4.5, 2.4			
SHBG (nmol/l)	0.2	-0.6, 1.0	0.8	-0.2, 1.7	1.1	-0.5, 2.6	1.2	-0.2, 2.7	-0.0	-2.9, 2.7			
Androstenedione (ng/ml)	0.5	-0.2, 1.2	0.5	-0.4, 1.4	1.3	-0.2, 2.7	0.7	-0.6, 2.1	1.9	-0.6, 4.4			
Testosterone (ng/ml)	0.4	-0.3, 1.1	0.1	-0.8, 1.0	1.1	-0.3, 2.4	0.4	-1.0, 1.6	0.4	-2.0, 2.7			
Free testosterone (ng/ml)	0.3	-0.5, 1.0	-0.4	-1.4, 0.6	0.4	-1.4, 1.8	-0.5	-2.0, 0.9	0.5	-2.2, 3.2			
DHEA (ng/dl)	0.5	-0.4, 1.3	-0.0	-1.1, 1.0	0.8	-0.9, 2.5	0.3	-1.3, 1.9	1.0	-2.5, 4.4			
DHEA sulfate (μg/dl)	0.6	-0.4, 1.6	-0.4	-1.8, 0.9	0.5	-1.5, 2.4	0.5	-1.4, 2.4	0.6	-3.8, 4.7			
IGF-1 (ng/ml)	-0.5	-1.1, 0.1	-0.5	-1.3, 0.2	-1.2	-2.4, 0.0*	-1.0	-2.1, 0.0	-1.9	-4.1, 0.4			
IGFBP-3 (ng/ml)	-0.4	-0.7, -0.1*	-0.2	-0.5, 0.2	-0.8	-1.4, -0.2*	-0.6	-1.2, -0.1*	-0.8	-1.9, 0.3			
IGF-I/IGFBP-3	-0.1	-0.6, 0.4	0.4	-1.0, 0.3	-0.4	-1.4, 0.6	-0.4	-1.3, 0.5	-1.0	-3.0, 0.9			

^a Adjusted for age at blood draw (continuous), fasting status (<8, ≥8 h), laboratory batch, day of luteal blood draw (for luteal hormones, 3–7, 8–28 days, missing/unintended sample), smoking status (pack years, never, 1–10, 11–20, 20+ years), BMI in 1997 (continuous), BMI at age 18 (continuous), total energy intake (average of 1995 and 1999, continuous), alcohol intake (average of 1995 and 1999, continuous), parity (nulliparity, 1 and age at first birth <25, 1 and age at first birth ≥25, 2 and age at first birth <25, 2 and age at first birth ≥25, 3+ and age at first birth <25, 3+ and age at first birth ≥25), age at menarche [<12, 12, >12], and height (continuous)

* $P < 0.05$

Table 5 Geometric means across categories of percentage of energy from carbohydrates and estimated percentage difference in plasma hormone levels from substituting 5% of energy from carbohydrates for the equivalent amount of energy from fat or protein intake

N (range)	% Energy from total carbohydrate					% Difference in hormone levels	
	≤40%	40.1–45%	45.1–50%	50.1–55%	>55%	Estimate	95%CI
	21–27	44–65	93–139	128–195	109–166		
	Geometric means						
Estradiol (pg/ml)							
Follicular	44.1	40.8	48.0	47.6	46.4	1.7	–2.1, 5.3
Luteal	124	128	113	116	124	0.6	–2.1, 3.2
Free estradiol (pg/ml)							
Follicular	0.6	0.5	0.6	0.6	0.6	0.7	–3.1, 4.3
Luteal	1.6	1.5	1.4	1.5	1.6	1.1	–1.5, 3.7
Estrone (pg/ml)							
Follicular	39.2	38.6	41.3	41.1	38.7	–0.5	–3.3, 2.3
Luteal	72.7	77.5	72.2	77.0	74.6	0.8	–1.6, 3.1
Estrone sulfate (pg/ml)							
Follicular	644	614	696	692	659	0.3	–4.3, 4.7
Luteal	1,345	1,332	1,198	1,419	1,393	2.0	–2.9, 6.6
Progesterone (ng/ml)	1,174	1,317	1,361	1,343	1,330	0.4	–2.8, 3.4
SHBG (nmol/l)	56.3	69.2	60.8	60.1	60.0	–0.9	–3.6, 1.7
Androstenedione (ng/ml)	101	108	106	106	99	–1.6	–4.1, 0.8
Testosterone (ng/ml)	20.6	25.8	23.6	23.4	22.9	–0.6	–2.9, 1.7
Free testosterone (ng/ml)	0.2	0.2	0.2	0.2	0.2	0.0	–2.6, 2.6
DHEA (ng/dl)	612	659	635	665	606	–1.1	–4.2, 1.9
DHEA sulfate (μg/dl)	65.6	80.5	77.9	79.2	73.6	–0.7	–4.4, 2.9
IGF-I (ng/ml)	211	230	225	238	237	2.0	0.1, 3.9*
IGFBP-3 (ng/ml)	4,456	4,484	4,533	4,668	4,599	1.0	–0.0, 1.9
IGF-I/IGFBP-3	0.05	0.05	0.05	0.05	0.05	1.1	–0.7, 2.8

Adjusted for age at blood draw (continuous), fasting status (<8, ≥8 h), laboratory batch, day of luteal blood draw (for luteal hormones, 3–7, 8–28 days, missing/untimed sample), smoking status (pack years, never, 1–10, 11–20, 20+years), BMI in 1997 (continuous), BMI at age 18 (continuous), total energy intake (average of 1995 and 1999, continuous), alcohol intake (average of 1995 and 1999, continuous), parity (nulliparity, 1 and age at first birth <25, 1 and age at first birth ≥25, 2 and age at first birth <25, 2 and age at first birth ≥25, 3+ and age at first birth <25, 3+ and age at first birth ≥25), age at menarche (<12, 12, >12), and height (continuous)

* $P < 0.05$

studies had a simultaneous control group [65]. In one 2-month intervention study with 62 women, a significant reduction was observed in luteal estradiol and estrone levels among wheat-bran but not oat- or corn-bran supplements groups [66]. However, two other dietary intervention studies (using a 12-month low-fat/high-fiber intervention among 213 women [67] or a 2-month replacement of saturated fat with polyunsaturated fat among 14 women [68]) found no influence of diet change on blood estrogen levels in the luteal [67, 68] or follicular [68] phases. Several cross-sectional studies found significant positive associations between total and monounsaturated fat intakes and follicular estrone levels [69] or

inverse associations between the ratio of polyunsaturated to saturated fat (P/S) and estradiol and estrone during the luteal phase [70] or higher follicular plasma estrogen levels among high-fat/low-fiber group [71]. In other studies, no significant associations were observed between dietary fiber intakes and estrogen levels during the follicular [69, 70] or luteal [68, 70] phases. Cumulatively, no strong consistent associations have been observed, and weak to moderate effects remain uncertain; our study, which is substantially larger than previous cross-sectional studies, suggests there is little, if any, association between these macronutrients and premenopausal estrogens. The significantly higher levels of follicular estradiol among

Table 6 Geometric means across categories of total fiber and estimated percentage difference in plasma hormone levels from 5-g increment of total fiber

N (range)	Total fiber				% Difference in hormone levels	
	≤15 g	15.1–20 g	20.1–25 g	>25.1 g	Estimate	95%CI
	73–97	151–230	105–164	64–101		
	Geometric means				Estimate	95%CI
Estradiol (pg/ml)						
Follicular	46.1	47.6	45.6	45.3	−1.7	−6.3, 2.7
Luteal	117	124	119	118	0.2	−3.4, 3.7
Free estradiol (pg/ml)						
Follicular	0.6	0.6	0.6	0.6	−0.6	−4.6, 3.2
Luteal	1.4	1.6	1.5	1.4	0.4	−3.2, 3.8
Estrone (pg/ml)						
Follicular	39.8	41.7	39.8	37.3	−2.0	−4.8, 0.8
Luteal	72.5	74.8	79.6	74.0	1.4	−1.5, 4.2
Estrone sulfate (pg/ml)						
Follicular	682	705	669	597	−1.5	−6.8, 3.6
Luteal	1,340	1,319	1,465	1,312	2.5	−2.8, 7.6
Progesterone (ng/ml)	1,289	1,375	1,324	1,291	−1.0	−5.3, 3.0
SHBG (nmol/l)	62.7	61.7	59.3	60.7	−1.4	−4.6, 1.7
Androstenedione (ng/ml)	105	108	103	97.2	−3.0	−5.9, −0.1*
Testosterone (ng/ml)	23.1	24.5	22.6	22.7	−1.4	−4.2, 1.3
Free testosterone (ng/ml)	0.2	0.2	0.2	0.2	−0.5	−3.8, 2.6
DHEA (ng/dl)	594	684	631	594	−1.6	−5.8, 2.4
DHEA sulfate (μg/dl)	72.3	81.2	76.9	71.4	−0.9	−5.2, 3.3
IGF-I (ng/ml)	223	232	233	243	1.9	−0.4, 4.2
IGFBP-3 (ng/ml)	4,416	4,570	4,702	4,610	1.3	0.0, 2.5
IGF-I/IGFBP-3	0.05	0.05	0.05	0.05	0.7	−1.3, 2.6

Adjusted for age at blood draw (continuous), fasting status (<8, ≥8 h], laboratory batch, day of luteal blood draw (for luteal hormones, 3–7, 8–28 days, missing/untimed sample), smoking status (pack years, never, 1–10, 11–20, 20+years], BMI in 1997 (continuous), BMI at age 18 (continuous), total energy intake (average of 1995 and 1999, continuous), alcohol intake (average of 1995 and 1999, continuous), parity (nulliparity, 1 and age at first birth <25, 1 and age at first birth ≥25, 2 and age at first birth <25, 2 and age at first birth ≥25, 3+ and age at first birth <25, 3+ and age at first birth ≥25], age at menarche [<12, 12, >12], and height (continuous)

* $P < 0.05$

those with the lowest percent energy from fat suggested a threshold effect, but we need to be careful in interpreting these data as there were only 20 participants in this category of intake.

Fewer studies have been done to investigate the associations between diet and endogenous levels of SHBG, androgens, or progesterone. A 2-month intervention study reported a significant reduction in follicular androstenedione and an increase in luteal testosterone levels after a low-fat/high-fiber intervention [72]. Previous intervention studies also found no significant change in blood SHBG after a 2-month high-fiber intervention [66, 72] or in luteal progesterone levels after either a low-fat/high-fiber [67] or

bran supplement [66] intervention. Several cross-sectional studies found higher SHBG levels with increased monounsaturated fat intake [69] or in the high-fat/low-fiber intake group [71]. Other studies also reported significant positive associations between P/S intakes with DHEAS [70] or an inverse association between dietary fibers with serum luteal levels of androstenedione among premenopausal women [73]. However, in these studies, no differences were found in plasma testosterone levels between groups [71], and no significant associations of fat and fiber intake with androgens or SHBG [70] or fiber with androgen, SHBG, or progesterone [73]. Similarly, we observed no association between fat/fiber and plasma androgens or progesterone.

We found modest but significant associations between plasma IGF-I levels and both total fat (inverse association) and carbohydrates (positive association). Since the percentage of energy from total fat and from carbohydrates is highly correlated (Spearman correlation coefficient is -0.86), it is hard to distinguish the effect of fat versus carbohydrates. The inverse association between total fat and IGF-I is consistent with a previous intervention study which found that IGF-I levels rose significantly after an intervention to reduce total fat intake [74]. However, in other intervention studies, IGF-I levels did not change [75–77] after a low-fat/high-fiber diet intervention. Previous cross-sectional studies reported positive [49] or no [51, 78] associations between fat intake and IGF-I levels. The significant but modest inverse associations between intakes of total fat, animal fat, saturated fat, and monounsaturated fat and plasma IGF-I observed in our study are supported by some previous cross-sectional studies in which IGF-I levels were inversely related to fat intakes [48–50] but not another large cross-sectional study including 2,109, women which found no significant associations with dietary fat intake [51]. Our findings did not appear to be due to total energy or to protein intake, since neither of them was significant in our multivariate models. Residual confounding by known confounders also is unlikely since the results did not change substantially after multivariable adjustment. The modest positive association between carbohydrates and plasma IGF-I is inconsistent with the few prior cross-sectional studies which found no association among premenopausal women [78] or an inverse association among healthy adults ages 30 to 84 years old [49].

The strengths of our study include its relatively large sample size, generally low laboratory CVs, and evaluation of a large number of hormones with careful timing by menstrual cycle phase. Our study also had several potential limitations. The women in our study are well-nourished, and the average intake of fiber and percent energy from fat per day were 20.2 g/day and 29.1%, which were less extreme than those in a few prior studies [66, 70]. If there are associations between diet and endogenous hormone levels and the associations are non-linear (a possibility suggested by our findings on fat intake and follicular estradiol), our study might not have a sufficient number of subjects in the effect range. But we previously reported an inverse association between fat intakes and plasma sex steroid hormone levels among postmenopausal women [61], and associations of both total fat [79] and fiber [80] with disease risk. Error in our diet assessment is also a concern, although our use of two FFQs collected 4 years apart should dampen this error. Finally, although the intraclass correlation coefficients (ICC) for IGFs and

androgens were quite high (range from 0.59 to 0.89 over up to 3 years), the ICC for estrogens and progesterone were lower [57]. Although we controlled for day of luteal blood draw, the possible associations could be attenuated due to that source of within-person variability. Finally, we made a large number of comparisons in this analysis, and we cannot rule out the possibility of observing several associations by chance.

In conclusion, our results suggest that a low-fat/high-fiber and carbohydrate diet within the range of intake generally observed in the USA is not importantly associated with endogenous levels of sex steroid hormones but may modestly increase IGF-I and IGFBP-3 levels among premenopausal women. Further large studies are needed to investigate the relationships between dietary intakes and endogenous hormone levels throughout the menstrual cycle.

Acknowledgments The work was supported by the National Institutes of Health (grant numbers CA50385 and CA67262).

Conflict of Interest The authors declare that they have no conflict of interest.

Appendix 1: Hormones measured during menstrual cycle

	Follicular	Luteal	Untimed
Estradiol	X	X	
Estrone	X	X	
Estrone Sulfate		X	
Progesterone		X	
SHBG	X	X	X
Androstenedione	X	X	X
Testosterone	X	X	X
DHEA		X	X
DHEAS		X	X
IGF-I		X	X
IGFBP-3		X	X

X denotes hormones measured

References

- Potischman NR, Hoover N, Brinton LA, Siiteri P, Dorgan JF, Swanson CA, Berman ML et al (1996) Case-control study of endogenous steroid hormones and endometrial cancer. *J Natl Cancer Inst* 88(16):1127–1135
- Austin H, Austin JM Jr, Partridge EE, Hatch KD, Shingleton HM (1991) Endometrial cancer, obesity, and body fat distribution. *Cancer Res* 51(2):568–572

3. Mollerstrom G, Carlstrom K, Lagrelius A, Einhorn N (1993) Is there an altered steroid profile in patients with endometrial carcinoma? *Cancer* 72(1):173–181
4. Kaaks R, Lukanova A, Kurzer MS (2002) Obesity, endogenous hormones, and endometrial cancer risk: a synthetic review. *Cancer Epidemiol Biomark Prev* 11(12):1531–1543
5. Zeleniuch-Jacquotte A, Shore RE, Koenig KL, Akhmedkhanov A, Afanasyeva Y, Kato I, Kim MY, Rinaldi S, Kaaks R, Toniolo P (2004) Postmenopausal levels of oestrogen, androgen, and SHBG and breast cancer: long-term results of a prospective study. *Br J Cancer* 90(1):153–159
6. ESHRE Capri Workshop Group (2004) Hormones and breast cancer. *Hum Reprod Update* 10(4):281–293
7. Bernstein L (2002) Epidemiology of endocrine-related risk factors for breast cancer. *J Mammary Gland Biol Neoplasia* 7(1):3–15
8. Key T, Appleby P, Barnes I, Reeves G (2002) Endogenous sex hormones and breast cancer in postmenopausal women: reanalysis of nine prospective studies. *J Natl Cancer Inst* 94(8):606–616
9. Missmer SA, Eliassen AH, Barbieri RL, Hankinson SE (2004) Endogenous estrogen, androgen, and progesterone concentrations and breast cancer risk among postmenopausal women. *J Natl Cancer Inst* 96(24):1856–1865
10. Kaaks R, Rinaldi S, Key TJ, Berrino F, Peeters PH, Biessy C, Dossus L et al (2005) Postmenopausal serum androgens, oestrogens and breast cancer risk: the European prospective investigation into cancer and nutrition. *Endocr Relat Cancer* 12(4):1071–1082
11. Eliassen AH, Missmer SA, Tworoger SS, Spiegelman D, Barbieri RL, Dowsett M, Hankinson SE (2006) Endogenous steroid hormone concentrations and risk of breast cancer among premenopausal women. *J Natl Cancer Inst* 98(19):1406–1415
12. Kaaks R, Berrino F, Key T, Rinaldi S, Dossus L, Biessy C, Secreto G et al (2005) Serum sex steroids in premenopausal women and breast cancer risk within the European Prospective Investigation into Cancer and Nutrition (EPIC). *J Natl Cancer Inst* 97(10):755–765. doi:97/10/755[pii]10.1093/jnci/dji132
13. Riman T, Nilsson S, Persson IR (2004) Review of epidemiological evidence for reproductive and hormonal factors in relation to the risk of epithelial ovarian malignancies. *Acta Obstet Gynecol Scand* 83(9):783–795
14. Risch HA (1998) Hormonal etiology of epithelial ovarian cancer, with a hypothesis concerning the role of androgens and progesterone. *J Natl Cancer Inst* 90(23):1774–1786
15. Tentori L, Graziani G (2007) Doping with growth hormone/IGF-1, anabolic steroids or erythropoietin: is there a cancer risk? *Pharmacol Res* 55(5):359–369
16. Jenkins PJ, Mukherjee A, Shalet SM (2006) Does growth hormone cause cancer? *Clin Endocrinol (Oxf)* 64(2):115–121
17. Moschos SJ, Mantzoros CS (2002) The role of the IGF system in cancer: from basic to clinical studies and clinical applications. *Oncology* 63(4):317–332
18. Furstenberger G, Senn HJ (2002) Insulin-like growth factors and cancer. *Lancet Oncol* 3(5):298–302
19. Kaaks R (2004) Nutrition, insulin, IGF-1 metabolism and cancer risk: a summary of epidemiological evidence. *Novartis Found Symp* 262:247–260, discussion 260–268
20. Chan JM, Stampfer MJ, Giovannucci E, Gann PH, Ma J, Wilkinson P, Hennekens CH, Pollak M (1998) Plasma insulin-like growth factor-I and prostate cancer risk: a prospective study. *Science* 279(5350):563–566
21. Monti S, Proietti-Pannunzi L, Sciarra A, Lolli F, Falasca P, Poggi M, Celi FS, Toscano V (2007) The IGF axis in prostate cancer. *Curr Pharm Des* 13(7):719–727
22. Giovannucci E (1999) Insulin-like growth factor-I and binding protein-3 and risk of cancer. *Horm Res* 51(Suppl 3):34–41
23. Roddam AW, Allen NE, Appleby P, Key TJ, Ferrucci L, Carter HB, Metter EJ et al (2008) Insulin-like growth factors, their binding proteins, and prostate cancer risk: analysis of individual patient data from 12 prospective studies. *Ann Intern Med* 149(7):461–471, W483–468
24. Shi R, Yu H, McLarty J, Glass J (2004) IGF-I and breast cancer: a meta-analysis. *Int J Cancer* 111(3):418–423
25. Hankinson SE, Willett WC, Colditz GA, Hunter DJ, Michaud DS, Deroo B, Rosner B, Speizer FE, Pollak M (1998) Circulating concentrations of insulin-like growth factor-I and risk of breast cancer. *Lancet* 351(9113):1393–1396
26. Renehan AG, Zwahlen M, Minder C, O'Dwyer ST, Shalet SM, Egger M (2004) Insulin-like growth factor (IGF)-I, IGF binding protein-3, and cancer risk: systematic review and meta-regression analysis. *Lancet* 363(9418):1346–1353
27. Fletcher O, Gibson L, Johnson N, Altmann DR, Holly JM, Ashworth A, Peto J, Silva Idos S (2005) Polymorphisms and circulating levels in the insulin-like growth factor system and risk of breast cancer: a systematic review. *Cancer Epidemiol Biomark Prev* 14(1):2–19
28. Lann D, LeRoith D (2008) The role of endocrine insulin-like growth factor-I and insulin in breast cancer. *J Mammary Gland Biol Neoplasia* 13(4):371–379
29. Ma J, Pollak MN, Giovannucci E, Chan JM, Tao Y, Hennekens CH, Stampfer MJ (1999) Prospective study of colorectal cancer risk in men and plasma levels of insulin-like growth factor (IGF)-I and IGF-binding protein-3. *J Natl Cancer Inst* 91(7):620–625
30. Giovannucci E, Pollak MN, Platz EA, Willett WC, Stampfer MJ, Majeed N, Colditz GA, Speizer FE, Hankinson SE (2000) A prospective study of plasma insulin-like growth factor-I and binding protein-3 and risk of colorectal neoplasia in women. *Cancer Epidemiol Biomark Prev* 9(4):345–349
31. Gunter MJ, Hoover DR, Yu H, Wassertheil-Smoller S, Rohan TE, Manson JE, Li J et al (2009) Insulin, insulin-like growth factor-I, and risk of breast cancer in postmenopausal women. *J Natl Cancer Inst* 101(1):48–60
32. Yu H, Spitz MR, Mistry J, Gu J, Hong WK, Wu X (1999) Plasma levels of insulin-like growth factor-I and lung cancer risk: a case-control analysis. *J Natl Cancer Inst* 91(2):151–156
33. Bohlke K, Cramer DW, Trichopoulos D, Mantzoros CS (1998) Insulin-like growth factor-I in relation to premenopausal ductal carcinoma in situ of the breast. *Epidemiology* 9(5):570–573
34. Lukanova A, Lundin E, Micheli A, Arslan A, Ferrari P, Rinaldi S, Krogh V et al (2004) Circulating levels of sex steroid hormones and risk of endometrial cancer in postmenopausal women. *Int J Cancer* 108(3):425–432
35. Bandera EV, Kushi LH, Moore DF, Gifkins DM, McCullough ML (2007) Association between dietary fiber and endometrial cancer: a dose-response meta-analysis. *Am J Clin Nutr* 86(6):1730–1737
36. Xu WH, Dai Q, Xiang YB, Zhao GM, Ruan ZX, Cheng JR, Zheng W, Shu XO (2007) Nutritional factors in relation to endometrial cancer: a report from a population-based case-control study in Shanghai, China. *Int J Cancer* 120(8):1776–1781
37. Littman AJ, Beresford SA, White E (2001) The association of dietary fat and plant foods with endometrial cancer (United States). *Cancer Causes Control* 12(8):691–702
38. Zhang M, Lee AH, Binns CW (2004) Reproductive and dietary risk factors for epithelial ovarian cancer in China. *Gynecol Oncol* 92(1):320–326
39. La Vecchia C (2001) Epidemiology of ovarian cancer: a summary review. *Eur J Cancer Prev* 10(2):125–129
40. Edefonti V, Decarli A, La Vecchia C, Bosetti C, Randi G, Franceschi S, Dal Maso L, Ferraroni M (2008) Nutrient dietary patterns and the risk of breast and ovarian cancers. *Int J Cancer* 122(3):609–613

41. Pelucchi C, La Vecchia C, Chatenoud L, Negri E, Conti E, Montella M, Calza S, Dal Maso L, Franceschi S (2001) Dietary fibres and ovarian cancer risk. *Eur J Cancer* 37(17):2235–2239
42. Trock B, Lanza E, Greenwald P (1990) Dietary fiber, vegetables, and colon cancer: critical review and meta-analyses of the epidemiologic evidence. *J Natl Cancer Inst* 82(8):650–661
43. Prentice RL, Sheppard L (1990) Dietary fat and cancer: consistency of the epidemiologic data, and disease prevention that may follow from a practical reduction in fat consumption. *Cancer Causes Control* 1(1):81–97, discussion 99–109
44. Slattery ML, Benson J, Berry TD, Duncan D, Edwards SL, Caan BJ, Potter JD (1997) Dietary sugar and colon cancer. *Cancer Epidemiol Biomark Prev* 6(9):677–685
45. Goldin BR, Adlercreutz H, Gorbach SL, Warram JH, Dwyer JT, Swenson L, Woods MN (1982) Estrogen excretion patterns and plasma levels in vegetarian and omnivorous women. *N Engl J Med* 307(25):1542–1547
46. Goldin BR, Adlercreutz H, Gorbach SL, Woods MN, Dwyer JT, Conlon T, Bohn E, Gershoff SN (1986) The relationship between estrogen levels and diets of Caucasian American and Oriental immigrant women. *Am J Clin Nutr* 44(6):945–953
47. Thissen JP, Ketelslegers JM, Underwood LE (1994) Nutritional regulation of the insulin-like growth factors. *Endocr Rev* 15(1):80–101
48. Holmes MD, Pollak MN, Willett WC, Hankinson SE (2002) Dietary correlates of plasma insulin-like growth factor I and insulin-like growth factor binding protein 3 concentrations. *Cancer Epidemiol Biomark Prev* 11(9):852–861
49. Kaklamani VG, Linos A, Kaklamani E, Markaki I, Koumantaki Y, Mantzoros CS (1999) Dietary fat and carbohydrates are independently associated with circulating insulin-like growth factor 1 and insulin-like growth factor-binding protein 3 concentrations in healthy adults. *J Clin Oncol* 17(10):3291–3298
50. Nagata C, Shimizu H, Takami R, Hayashi M, Takeda N, Yasuda K (2003) Dietary soy and fats in relation to serum insulin-like growth factor-1 and insulin-like growth factor-binding protein-3 levels in premenopausal Japanese women. *Nutr Cancer* 45(2):185–189
51. Norat T, Dossus L, Rinaldi S, Overvad K, Gronbaek H, Tjonneland A, Olsen A et al (2007) Diet, serum insulin-like growth factor-I and IGF-binding protein-3 in European women. *Eur J Clin Nutr* 61(1):91–98
52. Hankinson SE, London SJ, Chute CG, Barbieri RL, Jones L, Kaplan LA, Sacks FM, Stampfer MJ (1989) Effect of transport conditions on the stability of biochemical markers in blood. *Clin Chem* 35(12):2313–2316
53. Schernhammer ES, Holly JM, Hunter DJ, Pollak MN, Hankinson SE (2006) Insulin-like growth factor-I, its binding proteins (IGFBP-1 and IGFBP-3), and growth hormone and breast cancer risk in the nurses health study II. *Endocr Relat Cancer* 13(2):583–592
54. Tworoger SS, Sluss P, Hankinson SE (2006) Association between plasma prolactin concentrations and risk of breast cancer among predominately premenopausal women. *Cancer Res* 66(4):2476–2482
55. Eliassen AH, Missmer SA, Tworoger SS, Hankinson SE (2006) Endogenous steroid hormone concentrations and risk of breast cancer: does the association vary by a woman's predicted breast cancer risk? *J Clin Oncol* 24(12):1823–1830
56. Tworoger SS, Missmer SA, Eliassen AH, Spiegelman D, Folkert E, Dowsett M, Barbieri RL, Hankinson SE (2006) The association of plasma DHEA and DHEA sulfate with breast cancer risk in predominantly premenopausal women. *Cancer Epidemiol Biomark Prev* 15(5):967–971
57. Missmer SA, Spiegelman D, Bertone-Johnson ER, Barbieri RL, Pollak MN, Hankinson SE (2006) Reproducibility of plasma steroid hormones, prolactin, and insulin-like growth factor levels among premenopausal women over a 2- to 3-year period. *Cancer Epidemiol Biomark Prev* 15(5):972–978
58. Salvini S, Hunter DJ, Sampson L, Stampfer MJ, Colditz GA, Rosner B, Willett WC (1989) Food-based validation of a dietary questionnaire: the effects of week-to-week variation in food consumption. *Int J Epidemiol* 18(4):858–867
59. Willett W (1998) *Nutritional epidemiology*, 2nd edn. Oxford University Press, New York
60. Rimm EB, Giovannucci EL, Stampfer MJ, Colditz GA, Litin LB, Willett WC (1992) Reproducibility and validity of an expanded self-administered semiquantitative food frequency questionnaire among male health professionals. *Am J Epidemiol* 135(10):1114–1126, discussion 1127–1136
61. Holmes MD, Spiegelman D, Willett WC, Manson JE, Hunter DJ, Barbieri RL, Colditz GA, Hankinson SE (2000) Dietary fat intake and endogenous sex steroid hormone levels in postmenopausal women. *J Clin Oncol* 18(21):3668–3676
62. Haytowitz DB (1995) Information from USDA's nutrient data bank. *J Nutr* 125(7):1952–1955
63. Rosner B (1983) Percentage points for a generalized ESD many-outlier procedure. *Technometrics* 25:165–172
64. Wu AH, Pike MC, Stram DO (1999) Meta-analysis: dietary fat intake, serum estrogen levels, and the risk of breast cancer. *J Natl Cancer Inst* 91(6):529–534
65. Holmes MD, Schisterman EF, Spiegelman D, Hunter DJ, Willett WC (1999) Re: meta-analysis: dietary fat intake, serum estrogen levels, and the risk of breast cancer. *J Natl Cancer Inst* 91(17):1511–1512
66. Rose DP, Goldman M, Connolly JM, Strong LE (1991) High-fiber diet reduces serum estrogen concentrations in premenopausal women. *Am J Clin Nutr* 54(3):520–525
67. Gann PH, Chatterton RT, Gapstur SM, Liu K, Garside D, Giovanazzi S, Thedford K, Van Horn L (2003) The effects of a low-fat/high-fiber diet on sex hormone levels and menstrual cycling in premenopausal women: a 12-month randomized trial (the diet and hormone study). *Cancer* 98(9):1870–1879
68. Brown BD, Thomas W, Hutchins A, Martini MC, Slavin JL (2002) Types of dietary fat and soy minimally affect hormones and biomarkers associated with breast cancer risk in premenopausal women. *Nutr Cancer* 43(1):22–30
69. Nagata C, Takatsuka N, Kawakami N, Shimizu H (2000) Total and monounsaturated fat intake and serum estrogen concentrations in premenopausal Japanese women. *Nutr Cancer* 38(1):37–39
70. Dorgan JF, Reichman ME, Judd JT, Brown C, Longcope C, Schatzkin A, Forman M et al (1996) Relation of energy, fat, and fiber intakes to plasma concentrations of estrogens and androgens in premenopausal women. *Am J Clin Nutr* 64(1):25–31
71. Aubertin-Leheudre M, Gorbach S, Woods M, Dwyer JT, Goldin B, Adlercreutz H (2008) Fat/fiber intakes and sex hormones in healthy premenopausal women in USA. *J Steroid Biochem Mol Biol* 112(1–3):32–39
72. Schaefer EJ, Lamon-Fava S, Spiegelman D, Dwyer JT, Lichtenstein AH, McNamara JR, Goldin BR et al (1995) Changes in plasma lipoprotein concentrations and composition in response to a low-fat, high-fiber diet are associated with changes in serum estrogen concentrations in premenopausal women. *Metabolism* 44(6):749–756
73. Maskarinec G, Morimoto Y, Takata Y, Murphy SP, Stanczyk FZ (2006) Alcohol and dietary fibre intakes affect circulating sex hormones among premenopausal women. *Public Health Nutr* 9(7):875–881
74. Heald AH, Golding C, Sharma R, Siddals K, Kirk S, Lawton C, Anderson S, Martin Gibson J, Cade JE (2004) A substitution model of dietary manipulation is an effective means of optimising lipid profile, reducing C-reactive protein and increasing insulin-like growth factor-1. *Br J Nutr* 92(5):809–818

75. Al-Delaimy WK, Natarajan L, Rock CL, Sun S, Flatt SW, Pierce JP (2006) Insulin-like growth factor I, insulin-like growth factor I binding protein 1, insulin, glucose, and leptin serum levels are not influenced by a reduced-fat, high-fiber diet intervention. *Cancer Epidemiol Biomark Prev* 15(6):1238–1239
76. Gann PH, Kazer R, Chatterton R, Gapstur S, Thedford K, Helenowski I, Giovanazzi S, Van Horn L (2005) Sequential, randomized trial of a low-fat, high-fiber diet and soy supplementation: effects on circulating IGF-I and its binding proteins in premenopausal women. *Int J Cancer* 116(2):297–303
77. Kaaks R, Bellati C, Venturelli E, Rinaldi S, Secreto G, Biessy C, Pala V, Sieri S, Berrino F (2003) Effects of dietary intervention on IGF-I and IGF-binding proteins, and related alterations in sex steroid metabolism: the diet and androgens (DIANA) randomised trial. *Eur J Clin Nutr* 57(9):1079–1088
78. Maskarinec G, Takata Y, Kaaks R (2005) The relation between nutritional factors and insulin-like growth factor-I in premenopausal women of different ethnicity. *Eur J Nutr* 44(2):105–113
79. Hu FB, Stampfer MJ, Manson JE et al (1997) Dietary fat intake and the risk of coronary heart disease in women. *N Engl J Med* 337(21):1491–1499
80. Wolk A, Manson JE, Stampfer MJ, Colditz GA, Hu FB, Speizer FE, Hennekens CH, Willett WC (1999) Long-term intake of dietary fiber and decreased risk of coronary heart disease among women. *JAMA* 281(21):1998–2004