Chapter 20 Anthropogenic and Naturally Produced Contaminants in Fish Oil: Role in III Health

Adrian Covaci and Alin C. Dirtu

Key Points

- Fish oil dietary supplements are recommended to increase the intake of polyunsaturated fatty acids (PUFAs), such as eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA), renowned for their beneficial effects to human health.
- Fish oil dietary supplements contain anthropogenic contaminants, such as organochlorine pesticides, polychlorinated biphenyls, polychlorinated dioxins and furans, polybrominated diphenyl ethers and mercury. Recently, a number of organobrominated compounds, such as methoxylated-PBDEs and polybrominated hexahydroxanthenes derivatives, naturally produced by marine organisms (e.g., algae and sponges) have also been identified in commercial fish oil dietary supplements.
- Since fish oil dietary supplements are consumed on a daily basis, concerns are issued about the presence of various contaminants in these capsules with improvements in the preparation and purification of supplements have reduced dramatically the contaminant's concentrations.
- Fish oil dietary supplements might be a suitable alternative to fish consumption for certain groups of the population for which fish consumption advice has been issued such as pregnant women or children.
- There is also a stringent need to regularly monitor the presence of "classical" and "new" contaminants together with naturally occurring compounds, in marine products destined for human consumption.

Keywords Anthropogenic \cdot Organohalogenated contaminants \cdot Naturally produced \cdot Beneficial health effect \cdot Fish oil dietary supplements \cdot Dietary intake

A. Covaci (🖂)

Department of Pharmaceutical Sciences, Toxicological Center, University of Antwerp, Universiteitsplein 1, 2610, Antwerp, Belgium e-mail: adrian.covaci@ua.ac.be

1 Beneficial Effects of Consumption of Fish Oils Rich in *n*-3 Unsaturated Fatty Acids

Both marine fish and fish-derived products, e.g., fish oils, contain essential long-chain polyunsaturated fatty acids (PUFAs), such as 5,8,11,14,17-eicosapentaenoic acid (EPA) and 4,7,10,13,16,19-docosahexaenoic acid (DHA), which are essential in the human diet [1]. They are needed for many metabolic functions including growth, structural maintenance, repairing of nervous tissue, cellular membrane phospholipid structure, or regulation of lipid metabolism [1–3]. Moreover, the intake of high amounts of PUFAs has been suggested to have several beneficial effects to human health, including decreasing the incidence and progression of vascular diseases, as well as reducing the symptoms of multiple sclerosis and/or osteoporosis [2, 3]. In recent years, fish oil dietary supplements (FODS) have been increasingly promoted as an alternative to fish consumption. Indeed, FODS may contain high and balanced custom-made amounts of DHA and EPA [1] and can sometimes be found in combinations with other nutritional supplements, such as vitamins, minerals, or even other PUFAs.

1.1 n – 3 Fatty Acids and Cardiovascular Diseases

The evidence from prospective studies and randomized trials [4–25] suggests that ingestion of n - 3 fatty acids (especially EPA and DHA) through consumption of fish or fish oil might beneficially influence cardiovascular disease. The first studies that showed the importance of fish consumption demonstrated low rates of death from coronary heart disease (CHD) among Greenland Eskimos [26]. Therefore, many effects were reported in relation with EPA and DHA ingestion, namely preventing arrhythmias [27], lowering plasma triacylglycerols [28, 29], decreasing blood pressure [30], decreasing platelet aggregation [31, 32], improving vascular reactivity [33, 34], and decreasing inflammation [35].

Across different studies, compared with little or no intake, modest consumption of n - 3 fatty acids (250–500 mg/d of EPA and DHA) lowers relative risk by more than 25%. Higher intakes do not substantially further lower CHD mortality, suggesting a threshold effect [36]. This threshold effect explains findings among Japanese populations [18, 24] in whom high background fish intake (e.g., median 900 mg/d of EPA and DHA) is associated with very low CHD death rates (87% lower than comparable Western populations) [10, 18], and additional n - 3 PUFA intake predicts little further reduction in CHD death. When comparing different types of fish, lower risk appears more strongly related to intake of oily fish (e.g., salmon, herring, sardines), rather than lean fish (e.g., cod, catfish, halibut) [11, 16]. Fish intake may modestly affect other cardiovascular outcomes, but evidence is not as robust as for CHD death [24, 37–44].

n - 3 PUFAs may influence several cardiovascular risk factors [23, 24, 27, 42–50]. Effects occur within weeks of intake and may result in altered membrane fluidity and receptor responses following incorporation of omega-3 PUFAs into cell membranes [51, 52] and direct binding of omega-3 PUFAs to intracellular receptors regulating gene transcription [53].

The heterogeneity of the effects of fish or fish oil intake on cardiovascular outcomes is likely related to varying dose and time responses of effects on the risk factors [3]. At typical dietary

intakes, anti-arrhythmic effects predominate, reducing risk of sudden death and CHD death within weeks. At higher doses, maximum anti-arrhythmic effects have been achieved, but other physiologic effects may modestly impact other clinical outcomes (possibly requiring years to produce clinical benefits). Yet, the heterogeneity of clinical effects may also be related to differing pathophysiologies of the clinical outcomes. Biological differences in the development of atherosclerosis vs. acute plaque rupture/thrombosis vs. arrhythmia would account for heterogeneous effects of n - 3 PUFAs on plaque progression vs. nonfatal myocardial infarction vs. CHD death.

Fish may replace other foods in the diet, such as meats or dairy products. However, the increase in fish consumption is unlikely to have important health benefits, since the replaced foods are highly variable among individuals and across cultures. n-3 PUFAs most strongly affect CHD death [10, 14, 15] and are unlikely to affect appreciably other causes of mortality.

Effects on total mortality in a population would therefore depend on the proportion of deaths due to CHD, ranging from one quarter of deaths in middle-age populations [54] to one half of deaths in populations with established CHD [10]. This is consistent with a meta-analysis of randomized trials through 2003 that found a nonsignificant 14% reduction in total mortality with n - 3 PUFAs [5, 10, 25, 55, 56]. When additional placebo-controlled, double-blind, randomized trials performed since 2003 were added [41], marine n - 3 PUFAs reduced total mortality by 17% (pooled relative risk, 0.83; 95% CI, 0.68–1.00; P = 0.046).

1.2 Neurologic Development

DHA is preferentially incorporated into the rapidly developing brain during gestation and the first 2 years of infancy, concentrating in gray matter and retinal membranes [57]. Infants can convert shorter chain n - 3 PUFAs to DHA [58], but it is unknown whether such conversion is adequate for the developing brain in the absence of maternal intake of DHA [59].

Effects of maternal DHA consumption on neurodevelopment have been investigated in observational studies and randomized trials, with heterogeneity in assessed outcomes (visual acuity, global cognition, specific neurologic domains) and timing of DHA intake (gestational vs. nursing). In a meta-analysis of 14 trials, DHA supplementation improved visual acuity in a dose-dependent manner [60]. Results for cognitive testing are less consistent, possibly due to differences in neurologic domains evaluated [57, 59, 61]. A quantitative pooled analysis of eight trials estimated that increasing maternal intake of DHA by 100 mg/d leads to an increased child IQ by 0.13 points (95% CI, 0.08–0.18) [62].

Most trials evaluated effects of maternal DHA intake during nursing, rather than pregnancy. In a trial among 341 pregnant women, treatment with cod liver oil from week 18 until 3 months postpartum increased DHA levels in cord blood by 50% and raised mental processing scores, a measure of intelligence, at age 4 [63]. This is consistent with observational studies showing positive associations between maternal DHA levels or fish intake during pregnancy and behavioral attention scores, visual recognition memory, and language comprehension in infancy [64–66]. Thus, while dose responses and specific effects require further investigation, these studies together indicate that maternal intake of DHA is beneficial for early neurodevelopment.

2 Toxic Contaminants from Fish Oil Dietary Supplements

2.1 Anthropogenic Contaminants

Besides PUFAs, it was already shown in many studies that fish may contain a variety of persistent contaminants, such as polychlorinated dibenzo-*p*-dioxins and furans (PCDD/PCDFs), polychlorinated biphenyls (PCBs), or polybrominated diphenyl ethers (PBDEs). This may result in a potential increase of health risks that could counteract the beneficial effects of n - 3 PUFAs [67, 68]. In general, the concentrations of such contaminants are proportional with the position of the fish in the food chain. Therefore, fish situated at the base of the food chain usually carries lower levels of contaminants compared to predatory fish situated high on the fish chain [69]. Another important parameter related to contaminant's concentrations in fish is the content of fat; fatty fishes (e.g., salmon, herring) contain significantly higher concentrations of persistent contaminants when compared to lean fish species [69].

The ingestion of persistent contaminants through fish consumption may lead to a wide range of toxicological and hormonal effects, including endocrine disruption, reproductive, neurobehavioral, and developmental disturbances [3, 70]. Such toxic effects on human health recorded for these contaminants made several environmental and health agencies to have already issued consumption recommendations, which range between 0.5 and 2 meals of fatty fish per month [67]. The general public is given seemingly conflicting reports about the risks and benefits of fish intake, resulting in controversy and confusion over fish and fish-derived products and their role in regard to a healthy diet [71].

Nevertheless, these contaminants persist for long periods in the environment, and thus, while levels are steadily declining, PCBs and dioxins continue to be present in low concentrations in many food items.

Considering these information, it become obvious that FODS may also be a potential source of toxic contaminants, especially when the fish oil produced originates from fish caught in contaminated waters or from farmed fish fed with contaminated feed. Fish oil produced from these sources may contain markedly higher amounts of contaminants than fish originating from less polluted sites [72–74]. Since FODS are recommended to be taken on a daily basis, it is therefore important to closely monitor the levels of contaminants that might be contained by these PUFA-enriched products. Hereby, the presence of above-mentioned contaminants in fish oils may counteract the highly claimed benefits of such capsules.

The following paragraphs will be focused on discussing the levels and profiles of each class of contaminants reported as present in FODS and also in relation with their acceptable norms.

2.1.1 Polychlorinated Biphenyls

PCBs are synthetic organochlorine compounds previously used in industrial and commercial processes, but their manufacture and processing was prohibited in 1977 [75]. Based on exposed animal experiments and also some evidence in humans, PCBs may cause adverse human health effects, such as cancer (possibly related to effects on the aryl hydrocarbon receptor), may interfere with transcription factors affecting gene expression, may affect the immune system, and may cause neurological effects [76, 77]. Also prenatal exposure to PCBs has been associated

with childhood neurodevelopmental deficits in several studies [78–80]. Hereby, in order to counteract the negative effects on human health by the presence of PCBs in FODS, the additional health risks would have to exceed possible benefits by more than 100-fold to meaningfully alter the present estimates of risks vs. benefits [3].

From their chemical structure, theoretically PCBs may exist in a maximum number of 209 congeners (generically listed according to the number/position of the chlorine atoms in the molecule), although only about 130 are found in commercial PCB mixtures [75]. From all possible PCBs, 12 congeners do not present any chlorine atoms on *ortho*-positions (non-*ortho* PCBs) or have only one chlorine in *ortho*-position (mono-*ortho* PCBs). These congeners present a coplanar chemical structure, having a geometrical configuration like 2,3,7,8-tetrachlorodibenzo-*p*-dioxin (2,3,7,8-TCDD), being therefore called *dioxin-like PCBs* (DL-PCBs). The coplanar PCBs have demonstrated a close toxicological similarity to dioxins and are thought to operate by the same general mechanism [81].

However, depending on the mixture containing PCBs used, the number of possible congeners which may be measured in fish and fish oil samples differs. In general, the profile for these contaminants consists from a smaller number of congeners, mainly tri- (CB-28), tetra- (CB-52), penta- (CB-101, 118), hexa- (CB-138, 149, 153), and hepta- (CB-170, 180) chlorinated components [75]. From above-mentioned PCB congeners, the least important are CB-28 and CB-52, which usually are measured in such samples close to the limit of quantitation of different methods applied, while the most important are CB-153, CB-118, and CB-138. Considering only the number of chlorine atoms in molecule, the profile of PCBs measured in general in FODS consists from penta- and hexa-CBs which are usually present at highest concentrations, followed immediately by hepta-CBs and at the lowest levels being usually measured tetra-CB congeners. Regarding the concentration in which PCBs are usually measured in such samples, this parameter differs according to a multitude of factors, namely year/region of sample collection, type of purification process applied on the fish oil considered, type of fish, or fish organ used for oil manufacture. A non-exhaustive summary of the available literature related to the content of PCBs, including DL-PCBs, from fish oils is presented in Table 20.1.

Most of the samples included in presented studies from Table 20.1 shows that reported Σ PCB concentrations for are below of the Belgian regulatory limit (75 ng PCBs/g), with very low levels in some samples [73, 85, 86]. There are also cases where the concentrations seem to exceed the regulatory limits for PCBs, namely for unrefined oil [87] or for samples collected in the late 1990s [82], when most probably the control of such hazardous substances was not as rigorous as it is nowadays or the refining processes were not very developed. The type of fish or fish tissue used to produce fish oils can easily influence the concentrations of contaminants in the obtained products. Therefore, oils obtained from cod liver appear to be significantly more contaminated compared with the one obtained from whole fish [73].

2.1.2 Polychlorinated Dibenzo-p-dioxins and Polychlorinated Dibenzofurans

The chemical compounds included in the class of polychlorinated dibenzo-*p*-dioxins (PCDDs) (75 different components) and also in the class of polychlorinated dibenzofurans (PCDFs) (135 different components) are commonly referring to the term "*dioxins*." They are organochlorine by-products of waste incineration, paper bleaching, pesticide production, and production

	Median (range) concent	rations (ng/g)			
	ΣPCBs (ng/g)	ΣOCPs (ng/g)	ΣDL-PCBs (pg WHO-TEQ/g)	ΣPCDD/Fs (pg WHO-TEQ/g)	
Legal limit (ng/g)	75	1,000	10	2	
Sample (origin of sample), N					Year (Reference)
FODS (mainly cod liver oil) (Australia), $N = 5$	9 (5–341) ^a	9 (4–227) ^{a,b}	1	I	1994–1995 [82]
FODS (fish oil) (Belgium), $N = 4$	124 (20–744) ^a	19 (5–851) ^{a,b}	I	I	1994–1995 [82]
FODS (mainly cod liver oil) (the United Kingdom), N = 6	655 (5–975) ^a	84 (5–128) ^{a,b}	I	I	1994–1995 [82]
Fish oil (Japan), $N = 41$	I	I	8.2 (0.91–19.3)	2.0 (0.2-4.9)	2000-2005 [83]
FODS (cod liver oil), $N = 7$	108 (87-202)	138 (100–224)	I	I	2001-2002 [73]
FODS (fish oil), $N = 6$	34 (0-49)	31 (5-47)	I	I	2001–2002 [73]
FODS (unspecified) (the USA), $N = 20$	50.4 (10.3–94.3)	I	I	I	2003 [84]
FODS (cod oil) (the USA), $N = 4$	153.3 (47.4–276.2)	I	I	I	2003 [84]
FODS (cod liver) (Italy), $N = 15$	86 (25–201)	71 (25–133) ^b	I	I	2004 [74]
FODS (Belgium), $N = 27$	12 (<0.3–57)	3.9 (<0.3–150)	I	I	2004-2006 [85]
FODS (The Netherlands), $N = 17$	14 (<0.3–60)	2.9 (<0.3–230)	I	I	2004–2006 [85]
FODS (the United Kingdom), $N = 12$	7.6 (<0.3–22)	4.4 (<0.3–62)	I	I	2004–2006 [85]
FODS (other countries) ^c , $N = 13$	6.0 (<0.3–95)	3.8 (<0.3–21)	I	I	2004–2006 [85]
FODS (mixed—no salmon), $N = 8$	24.2 (0.711–37.9)	10.6 (0.189–15.2) ^b	I	I	2005–2007 [86]

326

(continued)	
able 20.1	

	Median (range) concen	trations (ng/g)			
	ΣPCBs (ng/g)	ΣOCPs (ng/g)	ΣDL-PCBs (pg WHO-TEQ/g)	ΣPCDD/Fs (pg WHO-TEQ/g)	
Legal limit (ng/g)	75	1,000	10	2	
Sample (origin of sample), N					Year (Reference)
FODS (mixed— including salmon), $N = 6$	25.1 (19.3–26.5)	11.1 (9.31–24.9) ^b	I	I	2005–2007 [86]
FODS (salmon), $N = 7$	95.3 (36.1–170)	59.2 (4.76–250) ^b	I	I	2005-2007 [86]
Shark liver oil (Japan), $N = 3^d$	320 (290–340)	I	I	I	2006 [87]
Shark liver oil (New Zealand), $N = 3^d$	22 (19–43)	I	I	I	2006 [87]
Shark liver oil, $N = 6$	18.5 (16-31)	I	I	I	2006 [87]
FODS (mainly Pacific fish, for Switzerland), $N = 6$	13 (0.23–17)	Ι	1.15 (0.038–1.3)	0.65 (0.32–0.83)	2006 [88]
FODS (mainly cod liver oil) (the United Kingdom), N = 30	I	I	9.4 (1.1–41.5)	0.9 (0.2–8.4)	2006 [89]
Fish oil (cod liver oil) $(S_{moin}) = N - 1$	I	13.2	I	I	2007 [90]
(Spain), $N = 1$ Fish oil (salmon) (Spain), N = 2	I	38.4 (25.6–51.3)	I	I	2007 [90]
Fish oil (Sweden), $N = 5$	I	0.74 (0.16–33) ^e	0.17 (0.02–1.1)	0.57 (0.09 - 0.86)	2008 [91]
Seal Oil (Sweden), $N = 1$	I	210 ^e	4.9	2.3	2008 [91]
^a Results were recalculated in ng ^b DOCPs are reported as DDDT	g/g using an average densit. Γs	/ of oil of 0.924 g/mL			
^c Denmark, South Africa, USA,	France, and Sweden				
^d No heating step in the refining $^{e}\Sigma OCPs$ are reported as ΣHCF	; process Hs + ΣDDTs				

of polyvinyl chloride plastics [92]. Their toxicity was well established through various studies [93, 94] and therefore controlling emissions of these chemicals was of a high concern. Because of monitoring programs implemented by most of the countries, total environmental releases of dioxins from all quantifiable sources decreased by 90% between 1987 and 2000 [92]. The results presented in Table 20.1 shows that in case of PCDD/Fs, median concentrations measured in different FODS collected from various locations are well below the maximum tolerable limit. Only one study reported higher concentrations of PCDD/Fs, but the value is assigned to an oil sample obtained from seal [91].

2.1.3 Organochlorine Pesticides

The most important pesticides from this class which are usually measured in fish or fish oil samples are hexachlorocyclohexanes (HCHs) (usually present in their four isomers: α -, β -, γ -, δ -HCH), DDT and metabolites (DDTs), and chlordanes. Commercial DDT is actually a mixture of several closely related compounds where the major component (up to 77%) is the p,p' isomer. The o,p' isomer is also present in significant amounts (15%), the rest of the mixture being constituted from p,p'-dichlorodiphenyldichloroethylene (p,p'-DDE) and p,p'-dichlorodiphenyldichloroethane (p,p'-DDD). p,p'-DDE and p,p'-DDD are also the major metabolites and breakdown products of DDT in the environment [95]. The term "*total DDT*" or $\Sigma DDTs$ is often used to refer to the sum of all DDT-related compounds (p,p'-DDT, o,p'-DDT, p,p'-DDE, and p,p'-DDD) in a sample.

From the pesticide formulations mentioned above, DDTs are by far the most abundant OCPs measured in fish and/or FODS. In fact, together with PCBs, DDTs constitute the main chlorinated contaminants found in general in marine samples. Therefore, the results from Table 20.1 related to the content of OCPs from FODS reported in the literature will often refer to the concentrations of Σ DDTs. Because of its ban in most of the countries, usually the main contributor to the Σ DDTs is the principal metabolite of *p*,*p*'-DDT, which is *p*,*p*'-DDE. However, none of the presented studies from Table 20.1 show concentration values for Σ DDTs higher than the EU regulatory limit (1,000 ng/g). Again, concentrations of Σ DDTs depend on the type of fish or organ used to produce FODS. As a consequence, oil supplements obtained from cod liver showed also for DDTs the highest median concentrations compared with other products [73, 82, 85].

2.1.4 Brominated Flame Retardants

Although several BFRs, such as polybrominated diphenyl ethers (PBDEs) and hexabromocyclododecane (HBCD), are found in quantifiable levels in wildlife and humans and have been extensively investigated in the last decade, we are still lacking information on the health effects caused by these compounds [70]. In humans, they are absorbed from the gastrointestinal tract and accumulate in fatty tissues [70]. It seems that they present acute toxicity at average doses, but their health effects from chronic exposure are of more concern, especially when they are related to the exposure of developing infants and wildlife. However, based on the available data, it is known that BFRs are associated with several health effects in animal studies, including neurobehavioral toxicity, thyroid hormone disruption, and possibly cancer, only for some PBDE congeners [70]. Even if limited information is available in the literature, there is some evidence that BFRs can cause developmental effects, endocrine disruption, immunotoxicity, reproductive, and long-term effects, including second-generation effects, reviewed by Birnbaum and Staskal [70] and Darnerud [96]. For PBDEs, there is some evidence available for estrogenic activity [96, 97], but more studies have to be undertaken to determine if low-dose exposures have estrogenic activity in humans or other species. However, their presence in fish in general and in FODS in particular is important and their monitoring should be carefully addressed for these new persistent contaminants.

Polybrominated Diphenyl Ethers

PBDEs are flame-retardant additives which are used in a wide array of household products in concentrations up to 30% by weight, typically between 2 and 6% (per weight) [98]. They are structurally related to PCBs and are produced commercially as mixtures. However, PBDE mixtures contain fewer congeners than the commercial PCB mixtures. The three commercial mixtures of PBDEs are penta-BDE, octa-BDE, and deca-BDE according to the number of bromine atoms in the dominating congeners of the mixtures. Since August 2004, the use of penta- and octa-BDE technical mixtures has been banned in the EU, followed in July 2008 by the ban on the use of deca-BDE mixture [98]. In USA, only California has banned the use of penta- and octa-BDE mixtures by the end of 2008, while other US states are currently in the phase-out legislation for PBDEs [98].

Even if PBDEs have been reported (sometimes in high levels) in marine environments [99], there are very few studies which monitored their presence in FODS. Generally, the most detected PBDE congeners in FODS are BDE 47, 99, and 100 (which usually contribute with >75% to the total PBDEs), while higher brominated congeners, such as BDE 153, 154, and 183, are in most of FODS below quantification limit [100], though reported levels of PBDEs are considerably lower than those of organochlorine contaminants, such as PCBs or OCPs [85]. This is probably due to improved selection of fish used for the FODS preparation and/or to the final purification methods used by different producers. Similar to PCBs and OCPs, the most contaminated samples were reported as FODS obtained from cod liver (Table 20.2) [73, 87, 101]. Interestingly, several FODS with an elevated PBDE content had also higher DHA content [100]. It is not clear whether this is due to the fish sources used for the preparation of FODS with high DHA content (e.g., tuna) or to the purification processes specific for DHA-enriched FODS.

Hexabromocyclododecane

HBCD is the third most widely used BFR in the world and on the second place in the EU [98]. It is mostly used in extruded and expanded polystyrene foams but also is used as insulation material in construction industry. HBCD is highly efficient so that very low levels are required to reach the desired flame retardancy. Other uses of HBCD are upholstered furniture, automobile interior textiles, car cushions and insulation blocks in trucks, packaging material, video cassette recorder housing, and electric and electronic equipment [98].

	Median (range)	Median (range) concentrations (ng/g)			
Sample (origin of sample)	ΣPBDEs	ΣHBCDs	Hg-MeHg	Year (Reference)	
FODS (cod liver oil), $N = 7$	20 (15-35)	-	-	2001–2002 [73]	
FODS (fish oil), $N = 6$	1.7 (0.8–2.6)	-	-	2001–2002 [73]	
FODS (Belgium), $N = 27$	0.4 (<0.1-45)	_	-	2004–2006 [100]	
FODS (The Netherlands), $N = 17$	0.5 (0.1–17)	-	-	2004–2006 [100]	
FODS (the United Kingdom), $N = 12$	0.2 (0.1-2.8)	_	-	2004–2006 [100]	
FODS (other countries) ^a , $N = 13$	0.4 (0.1–7.5)	_	-	2004–2006 [100]	
FODS (mainly Pacific fish, for Switzerland), $N = 6$	0.7 (0.069– 3.8)	_	_	2006 [88]	
Cod liver oil (North Sea), $N = 3$	28 (16–32)	5.4 (4–6.2)	-	2004–2008 [87, 101]	
Shark liver oil (Japan), $N = 3^{b}$	52 (49–53)	44 (44–45)	-	2004–2008 [87, 101]	
Shark liver oil (New Zealand), $N = 3^{b}$	0.6 (0.2–0.7)	<0.2	-	2004–2008 [87, 101]	
Shark liver oil^c , $N = 6$	0.2 (0.1–15)	<0.2 (<0.2–7.3)	-	2004–2008 [87, 101]	
Seal oil (Canada), $N = 2$	0.85 (0.8–0.9)	0.45 (0.4–0.5)	_	2004–2008 [87, 101]	
Catfish-eggs oil (Venezuela), $N = 22$	_	_	2.16 (1.8-2.97) ^d	1998 [108]	
FODS (fish oil), $N = 3$	-	_	38.8 (9.9–123)	2005 [109]	

Table 20.2 Median and concentration range (ng/g oil) of polybrominated diphenyl ethers (PBDEs), hexabromocyclododecane (HBCD), and total mercury (Hg + MeHg) in fish oil dietary supplements

^aDenmark, South Africa, USA, France, and Sweden

^bNo heating step in the refining process

^cOrigin (country of production) unspecified

^dResults were recalculated in ng/g using a density value of 0.9 g/mL

The concentrations of HCHD in fish are usually strongly correlated with the contamination level of the area from which the samples are collected or with the proximity of industrial activity areas since there are several studies reporting high levels of HBCD in such samples [102]. Because of the increasing temporal trends of HBCD in various environmental compartments, especially in aquatic environmental samples [99], HBCD content of fish oils used to prepare FODS should also be monitored. The results presented in Table 20.2 show that the reported levels of HBCD in FODS are slightly lower compared to PBDEs measured in the same samples [101].

2.1.5 Mercury and Methyl Mercury

The forms in which mercury is present in the environment are various, namely elemental (metallic) mercury (Hg⁰), inorganically bound mercury (Hg²⁺), and organically bound mercury, for example, monomethyl mercury (MeHg) or dimethyl mercury (Me₂Hg). When assessing the risk for the human health, one has to consider that organically bound mercury species are much more toxic than elemental or inorganic species [103]. MeHg is formed in aquatic systems from inorganic mercury through a methylation process by the action of anaerobic organisms. Because MeHg is formed in aquatic systems and because it is not readily eliminated from organisms, it is biomagnified in aquatic food chains from bacteria, to plankton, through macroinvertebrates, to herbivorous fish, and further to piscivorous fish. Therefore, the concentration of MeHg in the top-level aquatic predators can reach a level a million times higher than the level in the water [103]. There are many factors which may influence Hg concentrations in any given fish, namely fish species, the age and size of the fish, and the type of water body in which the sample was collected [103].

Once entered in human body, MeHg is readily and completely absorbed by the gastrointestinal tract and afterward it reacts through a complexation process mostly with amino acids. MeHg is a risk factor for cardiovascular disease through a variety of mechanisms potentially involving prooxidant effects via the generation of radical species and the inactivation of cellular antioxidant systems such as glutathione peroxidase and catalase [104]. Mechanistic studies indicate that MeHg can exert toxic effects on the vascular endothelium by depletion of sulfhydryls, increased oxidative stress, and activation of phospholipases [105, 106]. Nevertheless, estimation of the benefits for n - 3 FA intake vs. MeHg risks should be carefully addressed for different fish species, according to their content in toxic/nontoxic compounds [107].

Therefore, even if the toxicity of Hg and its derivatives was shown in various studies involving aquatic environment, their monitoring in FODS was not as regular as it might be expected and there are very few reports of Hg content for such supplements (Table 20.2).

2.2 Naturally Produced Halogenated Compounds

Several recent studies have described the presence in marine environment of naturally produced halogenated compounds, such as methoxylated polybrominated diphenyl ethers (MeO-PBDEs) [110–112], polybrominated hexahydroxanthene derivatives (PBHDs) [113, 114], or halogenated dimethyl bipyrroles (HDBPs) [115, 116]. Their presence was already confirmed in some fish species used for the preparation of FODS [117, 118]. All mentioned classes of naturally produced compounds have been sometimes measured in concentrations higher than contaminants usually targeted in monitoring schemes, but not much is known about their occurrence, their dietary intake from fish and fish-derived products, or about the potential toxicological effects of these compounds.

2.2.1 Polybrominated Methoxylated Diphenyl Ethers

MeO-PBDEs are produced by algae, bacteria, or sponges (cyanobacteria and red algae— *Ceramium tenuicorne*) [112] and have previously been found in various marine organisms, including fish and marine mammals [110, 111]. The presence of elevated concentrations of these compounds found in the higher levels of the marine food chain demonstrates their bioaccumulative properties. However, little is known of their potential toxicological effects.

Covaci et al. [100] analyzed several FODS collected from various locations (Table 20.3), and MeO-PBDEs were found at elevated levels in most of the samples. Moreover, there was no significant correlation between PBDEs and MeO-PBDEs levels (namely between BDE 47 and 6-MeO-BDE 47, one of the most abundant compounds measured in fish samples usually) showing

that these compounds could originate from other marine sources. However, some significant correlation between individual MeO-PBDEs (6-MeO-BDE 47 and 2-MeO-BDE 68) shows that it is highly plausible that these compounds have both accumulated from the similar (natural) sources. The main difference between the results within the samples was based on the origin of fish included in FOD manufacture: fish from Pacific and Atlantic Oceans showed higher levels on MeO-PBDEs, similar results being reported for marine mammals from the Southern hemisphere [110].

Table 20.3 Median and concentration range (ng/g oil) of naturally produced brominated compounds: methoxylated polybrominated diphenyl ethers (MeO-PBDEs) and polybrominated hexahydroxanthenes (PBHDs) in fish oil dietary supplements

	Median (range) con	ncentrations (ng/g)	
Sample (origin of sample)	ΣMeO-PBDEs	ΣPHBDs	Year (Reference)
FODS (Belgium), $N = 27$	4.6 (<0.2-1670)	3.8 (<1.3–98)	2004–2006 [100]
FODS (The Netherlands), $N = 17$	9.2 (<0.2–180)	22.9 (<1.3-158)	2004-2006 [100]
FODS (the United Kingdom), $N = 12$	14 (<0.2–315)	2.8 (<1.3-163)	2004–2006 [100]
FODS (other countries) ^a , $N = 13$	4.1 (<0.2–230)	14.2 (<1.3-203)	2004–2006 [100]

^aDenmark, South Africa, USA, France, and Sweden

2.2.2 Polybrominated Hexahydroxanthene Derivatives

PBHDs were recently identified by Hiebl et al. [113] as two congeners in fish and shellfish. Indeed, sponges of the *Cacospongia* genus, reported to occur in Australia, but also in the Mediterranean Sea, have been suggested as potential natural producers of PBHDs [113, 114]. In the fish oil survey by Covaci et al. [100], concentrations of PBHDs were similar to concentrations of MeO-PBDEs (Table 20.3), and in the most samples, higher than of PBDEs. Within the two mentioned classes of naturally produced compounds, the levels of Σ PBHDs did not correlate with Σ MeO-PBDEs ($R_s = 0.24$, p > 0.01), suggesting separate (natural) sources of their presence in fish/fish oil samples [100]. Similar concentrations of PBHDs were already determined in farmed fish from the Mediterranean Sea [113], in farmed mussels from New Zealand [113, 114], but also in bird eggs from Norway [119], indicating the transfer of these compounds throughout the marine food web.

2.2.3 Halogenated Dimethyl Bipyrroles

Specific sources of these compounds have not been yet identified, but radiocarbon analysis strongly suggests that halogenated dimethyl bipyrroles (HDBPs) are synthesized using a relatively recent source of carbon and thus likely have a biogenic origin [120, 121]. It was also assessed that they show persistency, have bioaccumulation potential, and are already globally distributed in marine environments. Due to similarities in physical properties and persistence, their environmental behavior has somewhat paralleled the behavior of persistent anthropogenic organohalogens, such as the higher chlorinated PCB congeners [115]. Although the geographical sources of HDBPs are poorly understood, these compounds seem to be more abundant in marine samples from the North Pacific Ocean rather than the Atlantic Ocean [122, 123]. The presence of HDBPs in FODS was not reported until now, but because of their levels in fish samples, it

might be suggested that this is a matter of time. Moreover, it is important to report their levels in fish samples when discussion about comparison of risks/benefits of fish/FODS consumption is addressed, even if their potential toxicological effects are not yet elucidated.

3 Intake of Contaminants Through Consumption of FODS

Since FODS are intended to be consumed on a daily basis, several studies have also calculated the daily intake of organohalogenated compounds from FODS.

In a study by Covaci et al. [100], PUFA-rich FODS (n = 69) from 37 producers were collected in 2006 from products available for sale on the Belgian market, but also from The Netherlands, Ireland, the United Kingdom, and South Africa. Information on recommended dosage as provided on the product labels together with the EPA and DHA composition was used to calculate the daily intake in pollutants. In order to estimate the contaminant intake, daily recommended consumption of the supplements, as provided on the product labels, was multiplied by the corresponding concentrations. Intakes (ng/day) were calculated using lower bound (LB) and upper bound (UB) methods, where non-detects were replaced with a value equal to zero or LOQ, respectively.

The investigated FODS contained 200–800 mg/g EPA and DHA and the recommended dosing for human consumption ranged between 1 and 3 g/day [3]. Due to the low contamination levels in the FODS analyzed in this particular survey [85, 100], FODS do not appear to increase substantially the dietary intake of PCBs: median daily intake was 29 and 42 times lower than the intake from fish consumption alone or from total diet, respectively (Fig. 20.1).

Similarly, the median daily intake of PBDEs was 8 and 16 times lower than the intake from fish consumption alone or from total diet, respectively (Fig. 20.1). Although fish consumption is an important contributor to the total dietary intake of PCBs or PBDEs (Fig. 20.1), the low intake of these contaminants from FODS (<10% of that from fish) suggests that purification processes were present during the preparation of the vast majority of the investigated FODS. Since the PBDE intake from FODS covers yet a wide range of values (Fig. 20.1), some FODS brands are either produced from contaminated fish or are insufficiently purified.

The low intake of PCBs and PBDEs through FODS therefore makes these supplements a suitable alternative for populations with low consumption of PUFA-rich food or for which fish consumption recommendations have been issued (e.g., pregnant women). FODS also prove to be a powerful source of EPA and DHA compared with PUFA-containing vegetables oils, such as soybean and rapeseed oil, which have approximately 10 times less EPA and do not appear to provide an effective metabolic source of DHA for the average consumer [73]. This renders FODS an efficient, relatively clean, and low caloric source of PUFAs.

In the same survey by Covaci et al. [100], the presence of naturally produced halogenated compounds has been reported in FODS. Using a similar approach, it was calculated that the median daily intakes of MeO-PBDEs and PBHDs from FODS were, respectively, 3 and 6 times higher than the median intake of PBDEs (3 ng/day), respectively (Fig. 20.1). For some brands, the daily intake of MeO-PBDEs and PBHDs from FODS was even higher than the total dietary intake of PBDEs. This emphasizes that the presence of these scarcely investigated compounds should not be overlooked. Likewise, MeO-PBDEs and PBHDs showed a large variation of intake estimates, encompassing the range of PBDE values.



Fig. 20.1 Dietary intake (ng/day) of PCBs and PBDEs from fish oil dietary supplements, fish consumption, and total diet. The box plots show the median, 25th and 75th percentiles, the lines give the 10th and 90th percentiles, while the dots represent the outliers. Values in the *box* plots represent the medians for each category. Literature data used for the daily intake of PBDEs and PCBs from fish consumption and total diet are taken from references [124–144]

An estimation of the dietary intake of brominated FODS compounds for children has also been determined [100]. For each group of compounds, the intake was lower for children than for adults due to a combination of lower amounts needed to be daily ingested and lower concentrations of brominated compounds in the FODS used for children.

The dietary intake from fish and seafood has also been reported for other natural compounds, such as DBPs. The daily intake estimate for Σ DBPs was <3.5 ng/day [115], a lower result compared to the naturally produced compounds investigated by Covaci et al. [100].

4 Decontamination of Commercial Fish Oil Supplements

Previously, we showed that a large variety of both anthropogenic contaminants and naturally produced compounds are present in FODS at sometime considerable amounts. Most of the abovementioned compounds are lipophilic and therefore they are mainly retained in the fatty tissues of fish used to produce the FODS. It is thus expected that such fish oils may incorporate a large variety of contaminants. If these fish oils are used as primary products directly obtained without any purification, they might represent a real risk to human health.

Fish oils are usually refined by neutralization, absorption, and distillation processes. The main refining techniques developed and published until now were focused on removing of mainly dioxins and DL-PCBs. The refining process is usually not an easy technique to be applied, many parameters being necessary to be rigorously optimized before its proper applicability [145].

The process optimization should consist of selecting purification parameters that would allow for maximum reduction of the toxic contaminants present in fish oil while retaining the favorable high fatty acid content. However, in many cases, the levels of contaminants following these refining processes are not adequately low for human consumption; furthermore, it is known that the removal of DL-PCBs from fish oil, particularly mono-*ortho* PCBs, is difficult [82, 146, 147].

The removal of contaminants from fish oil was applied using supercritical CO₂ extraction (SCE) and different adsorbents [148]. It was shown that when using SCE alone the removal efficiency was higher in case of DL-PCBs (70–90% efficiency), but lower for dioxins (some of PCDD/F congeners were removed only at an efficiency of 15%). In contrast, when using different adsorbent treatments, activated carbon showed high removal efficiencies (>90%) for PCDDs and PCDFs, but low (<30%) for DL-PCBs. A combination of both of these methods was suggested to be more effective in order to reduce the total TEQ value for dioxin-like contaminants [148].

These results were confirmed in 2007 when activated carbon adsorption on the reduction of POPs in fish oil was studied based on response surface methodology [149]. PCDD/Fs showed a rapid adsorption behavior and the TEQ levels could be reduced by 99%. Adsorption of DL-PCBs was less effective and depended on *ortho*-substitution, i.e., non-*ortho* PCBs were adsorbed more effectively than mono-*ortho* PCBs with a maximum of 87 and 21% reduction, respectively, corresponding to the DL-PCB-TEQ reduction of 73%.

In order to combine the best conditions for a proper purification of fish oils, refining techniques based on countercurrent supercritical CO_2 extraction (CC-SCE) and activated carbon treatment were developed [150]. This resulted in a 93% reduction in the sum of PCDD/Fs and DL-PCBs levels and by 85% reduction in the TEQ values. It was shown that CC-SCE is effective for the removal of DL-PCBs, whereas activated carbon treatment is effective for the removal of PCDD/Fs.

5 Conclusions

Diets rich in n - 3 fatty acids present in fish offer a number of health benefits, from fighting heart disease to boosting immunity and neurological improvement. However, many noxious persistent contaminants, such as PCBs, OCPs, PCDD/PCDFs, and PBDEs, accumulate in fatty tissue of fish and, as a consequence, they may end up in commercial fish oils produced from these fishes. In general, the levels of contaminants measured in FODS are below the legal consumption norms. In some cases, very low contaminant levels (close to the detection limit) were measured indicating improved sourcing and advanced refining processes. Consequently, the daily intake of persistent contaminants through FODS represents only a small percentage (between 3 and 12%) of the intake resulting from consumption of fish. Another advantage is the "à la carte" preparation of FODS containing high and balanced custom-made amounts of DHA and EPA, sometimes in combination with other nutritional supplements, such as vitamins, minerals, or even other PUFAs.

However, some FODS products contain high levels of contaminants and this aspect needs to be better investigated. There are a number of general factors which can be linked to the purity of the fish oil capsules.

- 1. A high degree of purity has been seen in FODS prepared from the oil of fish caught in clean waters (open-ocean) compared to more polluted fish from coastal areas or closed seas.
- 2. Oil derived from smaller species, such as anchovy, typically contains fewer contaminants than oil from larger fish (cod or salmon). This is most probably due to the shorter life of smaller fish, which leads to a lower accumulation of contaminants.
- 3. In general, oil produced from fish body tends to have lower contaminant concentrations than from (cod) liver or from oil of marine mammals (e.g., seal).
- 4. Some companies go through a number of advanced chemical-stripping or refining processes to remove contaminants from fish oil and consequently, products from these suppliers tend to be clean. Yet, most companies selling fish oil supplements do not advertise about their processing or refining procedures, nor they do identify the source of their fish, much less what species they had tapped for its oil.
- 5. In general, companies that had labeled their supplements as being certified free of halogenated contaminants or methyl mercury should be given credit for their efforts to improve their products and for transparency toward the consumers.
- 6. There is a need for inclusion of a larger number of anthropogenic contaminants, e.g., PBDEs, but also naturally produced halogenated compounds, in monitoring schemes of marine products destined for human consumption. There is also a need for appropriate monitoring and legislation for FODS.

Acknowledgments Dr. Adrian Covaci was financially supported by the Research Council of the University of Antwerp and by a postdoctoral fellowship from the Research Scientific Foundation—Flanders (FWO). Dr. Alin Dirtu acknowledges financial support from the University of Antwerp.

References

- 1. Sidhu KS. Health benefits and potential risks related to consumption of fish or fish oil. Regul Toxicol Pharm 2003; 38: 336–344.
- 2. Simopoulos A. Essential fatty acids in health and chronic disease. Am J Clin Nutr 1999; 70: 560S-569S.
- Mozaffarrian D, Rimm EB. Fish intake, contaminants, and human health. Evaluating the risks and the benefits. JAMA 2006; 296: 1885–1899.
- Kromhout D, Bosschieter EB, de Lezenne Coulander C. The inverse relation between fish consumption and 20-year mortality from coronary heart disease. New Engl J Med 1985; 312: 1205–1209.
- Burr ML, Fehily AM, Gilbert JF, Elwood PC, Fehily AM, Rogers S, Sweetnam PM, Deadman NM. Effects of changes in fat, fish, and fibre intakes on death and myocardial reinfarction: diet and reinfarction trial (DART). Lancet 1989; 2: 757–761.
- 6. Dolecek TA, Granditis G. Dietary polyunsaturated fatty acids and mortality in the Multiple Risk Factor Intervention Trial (MRFIT. World Rev Nutr Diet 1991; 66: 205–216.
- Kromhout D, Feskens EJ, Bowles CH. The protective effect of a small amount of fish on coronary heart disease mortality in an elderly population. Int J Epidemiol 1995; 24: 340–345.
- Daviglus ML, Stamler J, Orencia AJ, Dyer AR, Liu K, Greenland P, Walsh MK, Morris D, Shekelle RB. Fish consumption and the 30-year risk of fatal myocardial infarction. New Engl J Med 1997; 336: 1046–1053.
- 9. Albert CM, Hennekens CH, O'Donnell CJ, Ajani UA, Carey VJ, Willett WC, Ruskin JN, Manson JE. Fish consumption and risk of sudden cardiac death. JAMA 1998; 279: 23–28.
- Gruppo Italiano per lo Studio della Sopravvivenza nell'Infarto miocardico. Dietary supplementation with n-3 polyunsaturated fatty acids and vitamin E after myocardial infarction: results of the GISSI Prevenzione trial. Lancet 1999; 354: 447–455.
- 11. Oomen CM, Feskens EJ, Rasanen L, Fidanza F, Nissinen AM, Menotti A, Kok FJ, Kromhout D. Fish consumption and coronary heart disease mortality in Finland, Italy, and The Netherlands. Am J Epidemiol 2000; 151: 999–1006.

- Hooper L, Thompson RL, Harrison RA, Summerbell CD, Ness AR, Moore HJ, Worthington HV, Durrington PN, Higgins JPT, Capps NE, Riemersma RA, Ebrahim SBJ, Smith GD. Risks and benefits of omega-3 fats for mortality, cardiovascular disease, and cancer: systematic review. BMJ 2006; 332: 752–755.
- Hu FB, Bronner L, Willett WC, Stampfer MJ, Rexrode KM, Albert CM, Hunter D, Manson JE. Fish and omega-3 fatty acid intake and risk of coronary heart disease in women. JAMA 2002; 287: 1815–1821.
- 14. Albert CM, Campos H, Stampfer MJ, Ridker PM, Manson JE, Willett WC, Ma J. Blood levels of long-chain n-3 fatty acids and the risk of sudden death. New Engl J Med 2002; 346: 1113–1118.
- Lemaitre RN, King IB, Mozaffarian D, Kuller LH, Tracy RP, Siscovick DS. N-3 Polyunsaturated fatty acids, fatal ischemic heart disease, and nonfatal myocardial infarction in older adults: the Cardiovascular Health Study. Am J Clin Nutr 2003; 77: 319–325.
- Mozaffarian D, Lemaitre RN, Kuller LH, Burke GL, Tracy RP, Siscovick DS. Cardiac benefits of fish consumption may depend on the type of fish meal consumed: the Cardiovascular Health Study. Circulation 2003; 107: 1372–1377.
- Mozaffarian D, Ascherio A, Hu FB, Stampfer MJ, Willett WC, Siscovick DS, Rimm EB. Interplay between different polyunsaturated fatty acids and risk of coronary heart disease in men. Circulation 2005; 111: 157–164.
- Yokoyama M, Origasu H, Matsuzaki M et al. Effects of eicosapentaenoic acid (EPA) on major cardiovascular events in hypercholesterolemic patients: the Japan EPA Lipid Intervention Study (JELIS). Presented at: American Heart Association Scientific Sessions; Dallas, Tex, November 17, 2005.
- Fraser GE, Sabate J, Beeson WL, Strahan TM. A possible protective effect of nut consumption on risk of coronary heart disease: the Adventist Health Study. Arch Intern Med 1992; 152: 1416–1424.
- Mann JI, Appleby PN, Key TJ, Thorogood M. Dietary determinants of ischaemic heart disease in health conscious individuals. Heart 1997; 78: 450–455.
- Osler M, Andreasen AH, Hoidrup S. No inverse association between fish consumption and risk of death from all-causes, and incidence of coronary heart disease in middle-aged, Danish adults. J Clin Epidemiol 2003; 56: 274–279.
- Folsom AR, Demissie Z. Fish intake, marine omega-3 fatty acids, and mortality in a cohort of postmenopausal women. Am J Epidemiol 2004; 160: 1005–1010.
- Nakamura Y, Ueshima H, Okamura T, Kadowaki T, Hayakawa T, Kita Y, Tamaki S, Okayama A. Association between fish consumption and all-cause and cause-specific mortality in Japan: NIPPON DATA80, 1980–1999. Am J Med 2005; 118: 239–245.
- 24. Iso H, Kobayashi M, Ishihara J, Sasaki S, Okada K, Kita Y, Kokubo Y, Tsugane S. Intake of fish and n-3 fatty acids and risk of coronary heart disease among Japanese: the Japan Public Health Center-Based (JPHC) Study Cohort I. Circulation 2006; 113: 195–202.
- Burr ML, Ashfield-Watt PA, Dunstan FD, Fehily AM, Breay P, Ashton T, Zotos PC, Haboubi NAA, Elwood PC. Lack of benefit of dietary advice to men with angina: results of a controlled trial. Eur J Clin Nutr 2003; 57: 193–200.
- Bang HO, Dyerberg J. Lipid metabolism and ischemic heart disease in Greenland Eskimos. In: Draper H, (ed.), Advances in Nutrition Research. New York, NY: Plenum Press, 1–22, 1980.
- Leaf A, Kang JX, Xiao YF, Billman GE. Clinical prevention of sudden cardiac death by n-3 polyunsaturated fatty acids and mechanism of prevention of arrhythmias by n-3 fish oils. Circulation 2003; 107: 2646–2652.
- Harris WS. N-3 Fatty acids and serum lipoproteins: human studies. Am J Clin Nutr 1997; 65(suppl): 1645S–1654S.
- 29. Sacks FM, Katan M. Randomized clinical trials on the effects of dietary fat and carbohydrate on plasma lipoproteins and cardiovascular disease. Am J Med 2002; 113(suppl): 13S–24S.
- Geleijnse JM, Giltay EJ, Grobbee DE, Donders AR, Kok FJ. Blood pressure response to fish oil supplementation: meta-regression analysis of randomized trials. J Hypertens 2002; 20: 1493–1499.
- 31. Knapp HR. Dietary fatty acids in human thrombosis and hemostasis. Am J Clin Nutr 1997; 65(suppl): 1687S–1698S.
- 32. Hornstra G. Influence of dietary fat type on arterial thrombosis tendency. J Nutr Health Aging 2001; 5: 160–166.
- Harris WS, Rambjor GS, Windsor SL, Diederich D. N-3 Fatty acids and urinary excretion of nitric oxide metabolites in humans. Am J Clin Nutr 1997; 65: 459–464.
- Goodfellow J, Bellamy MF, Ramsey MW, Jones CJ, Lewis MJ. Dietary supplementation with marine omega-3 fatty acids improve systemic large artery endothelial function in subjects with hypercholesterolemia. J Am Coll Cardiol 2000; 35: 265–270.

- 35. Calder PC. Polyunsaturated fatty acids, inflammation, and immunity. Lipids 2001; 36: 1007–1024.
- Siscovick DS, Lemaitre RN, Mozaffarian D. The fish story: a diet-heart hypothesis with clinical implications: n-3 polyunsaturated fatty acids, myocardial vulnerability, and sudden death. Circulation 2003; 107: 2632–2634.
- Erkkila AT, Lichtenstein AH, Mozaffarian D, Herrington DM. Fish intake is associated with a reduced progression of coronary artery atherosclerosis in postmenopausal women with coronary artery disease. Am J Clin Nutr 2004; 80: 626–632.
- 38. He K, Song Y, Daviglus ML, Liu K, Van Horn L, Dyer AR, Goldbourt U, Greenland P. Fish consumption and incidence of stroke: a meta-analysis of cohort studies. Stroke 2004; 35: 1538–1542.
- Angerer P, Kothny W, Stork S, von Schacky C. Effect of dietary supplementation with omega-3 fatty acids on progression of atherosclerosis in carotid arteries. Cardiovasc Res 2002; 54: 183–190.
- Balk EM, Lichtenstein AH, Chung M, Kupelnick B, Chew P, Lau J. Effects of omega-3 fatty acids on coronary restenosis, intima-media thickness, and exercise tolerance: a systematic review. Atherosclerosis 2006; 184: 237–246.
- Leaf A, Albert CM, Josephson M, Steinhaus D, Kluger J, Kang JX, Cox B, Zhang H, Schoenfeld D. Prevention of fatal arrhythmias in high-risk subjects by fish oil n-3 fatty acid intake. Circulation 2005; 112: 2762–2768.
- 42. Mozaffarian D, Psaty BM, Rimm EB, Lemaitre RN, Burke GL, Lyles MF, Lefkowitz D, Siscovick S. Fish intake and risk of incident atrial fibrillation. Circulation 2004; 110: 368–373.
- Frost L, Vestergaard P. N-3 Fatty acids consumed from fish and risk of atrial fibrillation or flutter: the Danish Diet, Cancer, and Health Study. Am J Clin Nutr 2005; 81: 50–54.
- Mozaffarian D, Bryson CL, Lemaitre RN, Burke GL, Siscovick DS. Fish intake and risk of incident heart failure. J Am Coll Cardiol 2005; 45: 2015–2021.
- 45. Mori TA, Beilin LJ. Omega-3 fatty acids and inflammation. Curr Atheroscler Rep 2004; 6: 461-467.
- 46. Chin JP, Gust AP, Nestel PJ, Dart AM. Marine oils dose-dependently inhibit vasoconstriction of forearm resistance vessels in humans. Hypertension 1993; 21: 22–28.
- 47. Geleijnse JM, Giltay EJ, Grobbee DE, Donders AR, Kok FJ. Blood pressure response to fish oil supplementation: meta-regression analysis of randomized trials. J Hypertens 2002; 20: 1493–1499.
- Mozaffarian D, Geelen A, Brouwer IA, Geleijnse JM, Zock PL, Katan MB. Effect of fish oil on heart rate in humans: a meta-analysis of randomized controlled trials. Circulation 2005; 112: 1945–1952.
- 49. Nestel PJ. Fish oil and cardiovascular disease: lipids and arterial function. Am J Clin Nutr 2000; 71: 228S–2231S.
- Kristensen SD, Iversen AM, Schmidt EB. n-3 polyunsaturated fatty acids and coronary thrombosis. Lipids 2001; 36(suppl): S79–S82.
- Clandinin MT, Cheema S, Field CJ, Garg ML, Venkatraman J, Clandinin TR. Dietary fat: exogenous determination of membrane structure and cell function. FASEB J 1991; 5: 2761–2769.
- 52. Feller SE, Gawrisch K. Properties of docosahexaenoic-acid-containing lipids and their influence on the function of rhodopsin. Curr Opin Struct Biol 2005; 15: 416–422.
- Vanden Heuvel JP. Diet, fatty acids, and regulation of genes important for heart disease. Curr Atheroscler Rep 2004; 6: 432–440.
- Anderson RN, Smith LB. Division of Vital Statistics, Centers for Disease Control and Prevention. National Vital Statistics Reports: deaths: leading causes for 2002. http://www.cdc.gov/nchs/ data/nvsr/nvsr53/nvsr53_17.pdf. Accessed June 14, 2009.
- 55. Brox J, Olaussen K, Osterud B, Elvevoll EO, Bjornstad E, Brenn T, Brattebo G, Iversen H. A long-term seal- and cod-liver-oil supplementation in hypercholesterolemic subjects. Lipids 2001; 36: 7–13.
- Leaf A, Jorgensen MB, Jacobs AK, Cote G, Schoenfeld DA, Scheer J, Weiner BH, Slack JD, Kellett MA, Raizner AE, Weber PC, Mahrer PR, Rossouw JE. Do fish oils prevent restenosis after coronary angioplasty?. Circulation 1994; 90: 2248–2257.
- 57. Lewin GA, Schachter HM, Yuen D, Merchant P, Mamaladze V, Tsertsvadze A. Agency for Healthcare Research and Quality (AHRQ). Effects of omega-3 fatty acids on child and maternal health. Evid Rep Technol Assess (Summ) 2005; 118: 1–11.
- 58. Uauy R, Mena P, Wegher B, Nieto S, Salem N Jr. Long chain polyunsaturated fatty acid formation in neonates: effect of gestational age and intrauterine growth. Pediatr Res 2004; 47: 127–135.
- McCann JC, Ames BN. Is docosahexaenoic acid, an n-3 long-chain polyunsaturated fatty acid, required for development of normal brain function? An overview of evidence from cognitive and behavioral tests in humans and animals. Am J Clin Nutr 2005; 82: 281–295.

- 60. Uauy R, Hoffman DR, Mena P, Llanos A, Birch EE. Term infant studies of DHA and ARA supplementation on neurodevelopment: results of randomized controlled trials. J Pediatr 2003; 143: S17–S25.
- 61. Simmer K. Long-chain polyunsaturated fatty acid supplementation in infants born at term. Cochrane Database Syst Rev 2001; 4: CD000376.
- 62. Cohen JT, Bellinger DC, Connor WE, Shaywitz BA. A quantitative analysis of prenatal intake of n-3 polyunsaturated fatty acids and cognitive development. Am J Prev Med 2005; 29: 366–374.
- Helland IB, Smith L, Saarem K, Saugstad OD, Drevon CA. Maternal supplementation with very long-chain n-3 fatty acids during pregnancy and lactation augments children's IQ at 4 years of age. Pediatrics 2003; 111: e39–e44.
- Oken E, Wright RO, Kleinman KP, Bellinger D, Amarasiriwardena CJ, Hu H, Rich-Edwards JW, Gillman MW. Maternal fish consumption, hair mercury, and infant cognition in a US Cohort. Environ Health Persp 2005; 113: 1376–1380.
- Colombo J, Kannass KN, Shaddy DJ, Kundurthi S, Maikranz JM, Anderson CJ, Blaga OM, Carlson SE. Maternal DHA and the development of attention in infancy and toddlerhood. Child Dev 2004; 75: 1254–1267.
- Daniels JL, Longnecker MP, Rowland AS, Golding J. Fish intake during pregnancy and early cognitive development of offspring. Epidemiology 2004; 15: 394–402.
- Hites RA, Foran JA, Carpenter DO, Hamilton MC, Knuth BA, Schwager SJ. Global assessment of organic contaminants in farmed salmon. Science 2004; 303: 226–229.
- Hites RA, Foran JA, Schwager SJ, Knuth BA, Hamilton MC, Carpenter DO. Global assessment of polybrominated diphenyl ethers in farmed and wild salmon. Environ Sci Technol 2004; 38: 4945–4949.
- 69. van Leeuwen SPJ, van Velzen MJM, Swart CP, van der Veen I, Traag WA, de Boer J. Halogenated contaminants in farmed salmon, trout, tilapia, pangasius, and shrimp. Environ Sci Technol 2009; 43: 4009–4015.
- Birnbaum LS, Staskal DF. Brominated flame retardants: Cause for concern?. Environ Health Persp 2004; 112: 9–17.
- 71. Verbeke W, Sioen I, Pieniak Z, Van Camp J, De Henauw S. Consumer perception versus scientific evidence about health benefits and safety risks from fish consumption. Pub Health Nutr 2005; 8: 422–429.
- Jacobs M, Covaci A, Schepens P. Investigation of selected persistent organic pollutants in farmed Atlantic salmon (*Salmo salar*), salmon aquaculture feed, and fish oil components of the feed. Environ Sci Technol 2002; 36: 2797–2805.
- 73. Jacobs MN, Covaci A, Gheorghe A, Schepens P. Time trend investigation of PCBs, OCPs and PBDEs in n-3 polyunsaturated fatty acid rich dietary fish oil and vegetable oil supplements, nutritional relevance for human essential n-3 fatty acid requirements. J Agric Food Chem 2004; 52: 1780–1788.
- Storelli MM, Storelli A, Marcotrigiano GO. Polychlorinated biphenyls, hexachlorobenzene, hexachlorocyclohexane isomers, and pesticide organochlorine residues in cod-liver oil dietary supplements. J Food Protect 2004; 67: 1787–1791.
- 75. US Environmental Protection Agency. Polychlorinated biphenyls (PCBs). http://www.epa.gov/pcbs. Accessed June, 2009.
- World Health Organization (WHO). Assessment of the health risk of dioxins: re-evaluation of the Tolerable Daily Intake (TDI). WHO Consultation; May 25–29, Geneva, Switzerland. 1998.
- National Center for Environmental Assessment, US Environmental Protection Agency. PCBs: cancer doseresponse assessment and application to environmental mixtures. Washington, DC: US Environmental Protection Agency; 1996.
- Stewart PW, Reihman J, Lonky EI, Darvill TJ, Pagano J. Cognitive development in preschool children prenatally exposed to PCBs and MeHg. Neurotoxicol Teratol 2003; 25: 11–22.
- Schantz SL, Widholm JJ, Rice DC. Effects of PCB exposure on neuropsychological function in children. Environ Health Persp 2003; 111: 357–576.
- Nakajima S, Saijo Y, Kato S, Sasaki S, Uno K, Kanagami N, Hirakawa H, Hori T, Tobiishi K, Todaka T, Nakamura Y, Yanagiya S, Sengoku Y, Iida T, Sata F, Kishi R. Effects of prenatal exposure to polychlorinated biphenyls and dioxins on mental and motor development in Japanese children at 6 months of age. Environ Health Persp 2006; 114: 773–778.
- 81. Van den Berg M, Birnbaum L, Bosveld ATC, Brunstrom B, Cook P, Feeley M, Giesy JP, Hanberg A, Hasegawa R, Kennedy SW, Kubiak T, Larsen JC, van Leeuwen FXR, Liem AKD, Nolt C, Peterson RE, Poellinger L, Safe S, Schrenk D, Tillitt D, Tysklind M, Younes M, Waern F, Zacharewski T. Toxic equivalency factors (TEFs) for PCBs, PCDDs, PCDFs for humans and wildlife. Environ Health Persp 1998; 106: 775–792.

- Jacobs MN, Santillo D, Johnston PA, Wyatt CL, French MC. Organochlorine residues in fish oil dietary supplements: comparison with industrial grade oils. Chemosphere 1998; 37: 1709–1721.
- Hasegawa J, Guruge KS, Seike N, Shirai Y, Yamata T, Nakamura M, Handa H, Yamanaka N, Miyazaki S. Determination of PCDD/Fs and dioxin-like PCBs in fish oils for feed ingredients by congener-specific chemical analysis and CALUX bioassay. Chemosphere 2007; 69: 1188–1194.
- Shim SM, Santerre CR, Burgess JR, Deardoff DC. Omega-3 fatty acids and total polychlorinated biphenyls in 26 dietary supplements. J Food Sci 2003; 68: 2436–2440.
- Covaci A, Voorspoels S, Wyckmans J, Gelbin A, Neels H. Anthropogenic and natural organohalogenated compounds in fish oil dietary supplements from various countries. Organohalogen Compd 2006; 68: 5–8.
- Rawn DFK, Breakell K, Verigin V, Nicolidakis H, Sit D, Feeley M. Persistent organic pollutants in fish oil supplements on the Canadian market: Polychlorinated biphenyls and organochlorine insecticides. J Food Sci 2008; 74: T14–T19.
- Akutsu K, Tanaka Y, Hayakawa K. Occurrence of polybrominated diphenyl ethers and polychlorinated biphenyls in shark liver oil supplements. Food Addit Contam 2006; 23: 1323–1329.
- Zennegg M, Schmid P. PCDD/F, PCB, dioxin-like PCB and PBDE in fish oil used as dietary supplement in Switzerland. Organohalogen Compd 2006; 68: 1967–1971.
- Fernandes AR, Rose M, White S, Mortimer DN, Gem M. Dioxins and polychlorinated biphenyls (PCBs) in fish oil dietary supplements: Occurrence and human exposure in the UK. Food Addit Contam 2006; 23: 939–947.
- Nevado JJB, Martín-Doimeadiós RCR, Bernardo FJG, Fariñas NR. Development and validation of an analytical methodology for the determination of p,p'-DDT, p,p'-DDE and p,p'-DDD in fish oil pills. Microchem J 2007; 86: 183–188.
- Rehnmark M, Rehnmark S, Henkelmann B, Kotalik J, Bernhöft S, Pandelova M, Schramm KW. Omega-3 health products; the health beneficial effects of certain oils may be compromised by contaminating chemical pollutants. Organohalogen Compd 2008; 70: 1966–1969.
- 92. US EPA (Environmental Protection Agency). An inventory of sources and environmental releases of dioxin-like compounds in the United States for the years 1987, 1995, and 2000. National Center for Environmental Assessment, Washington, DC; EPA/600/P-03/002F. (http://cfpub.epa.gov/ncea/ cfm/recordisplay.cfm?deid=159286), 2006.
- Devito MJ, Birnbaum LS, Farland WH, Gasiewicz TA. Comparisons of estimated human body burdens of dioxin-like chemicals and TCDD body burdens in experimentally exposed animals. Environ Health Persp 1995; 103: 820–831.
- Ishimura R, Kawakmi T, Ohsako S, Tohyama C. Dioxin-induced toxicity on vascular remodeling of the placenta. Biochem Pharmacol 2009; 77: SI660–SI669.
- 95. Environmental Health Criteria 9: DDT and its derivatives, World Health Organization, 1979.
- 96. Darnerud PO. Toxic effects of brominated flame retardants in man and in wildlife. Environ Int 2003; 29: 841–853.
- 97. Legler J, Brouwer A. Are brominated flame retardants endocrine disruptors?. Environ Int 2003; 29: 879–885.
- Bromine Science and Environmental Forum (BSEF). Website: http://www.bsef.com. Last accessed June 14, 2009.
- Law RJ, Allchin CR, de Boer J, Covaci A, Herzke D, Lepom P, Morris S, Tronczynski J, de Wit CA. Levels and trends of brominated flame retardants in the European environment. Chemosphere 2006; 64: 187–208.
- Covaci A, Voorspoels S, Vetter W, Gelbin A, Jorens PG, Blust R, Neels H. Anthropogenic and naturallyproduced brominated compounds in fish oil dietary supplements. Environ Sci Technol 2007; 41: 5237–5244.
- Kakimoto K, Akutsu K, Konishi Y, Tanaka Y. Evaluation of hexabromocyclododecane in fish and marine mammal oil supplements. Food Chem 2008: 107: 1724–1727.
- Roosens L, Dirtu A, Goemans G, Belpaire C, Gheorghe A, Neels H, Blust R, Covaci A. Brominated flame retardants and organochlorine contaminants in fish samples along Scheldt River, Belgium. Environ Int 2008; 34: 976–983.
- 103. Wiener JG, Krabbenhoft DP, Heinz GH, Scheuhammer AM. Ecotoxicology of mercury. Chapter 16 In: Hoffman, DJ, Rattner, BA, Burton, GA Jr., Cairns, J Jr., eds., Handbook of Ecotoxicology. 2nd edn. Boca Raton, Florida: CRC Press, 409–463, 2003.
- 104. Guallar E, Sanz-Gallardo MI, van't Veer P, Bode P, Aro A, Gomez-Aracena J, Kark JD, Riemersma RA, Martin-Moreno JM, Kok FJ. Heavy metals and myocardial infarction study group. Mercury, fish oils, and the risk of myocardial infarction. New Engl J Med 2002; 347: 1747–1754.
- Hagele TJ, Mazerik JN, Gregory A, Kaufman B, Magalang U, Kuppusamy ML. Mercury activates vascular endothelial cell phospholipase D through thiols and oxidative stress. Int J Toxicol 2007; 26: 57–69.

- Mazerik JN, Hagele T, Sherwani S, Ciapala V, Butler S, Kuppusamy ML, Hunter M, Kuppusamy P, Marsh CB, Parinandi NL. Phospholipase A2 activation regulates cytotoxicity of methyl-mercury in vascular endothelial cells. Int J Toxicol 2007; 26: 553–569.
- Ginsberg GL, Toal BF. Quantitative approach for incorporating methyl-mercury risks and omega-3 fatty acid benefits in developing species-specific fish consumption advice. Environ Health Persp 2009; 117: 267–275.
- 108. Burguera JL, Quintan IA, Salager JL, Burguera M, Rondón C, Carrero P, Anton de Salager R, Petit de Peña Y. The use of emulsions for the determination of methylmercury and inorganic mercury in fish-eggs oil by cold vapor generation in a flow injection system with atomic absorption spectrometric detection. Analyst 1999; 124: 593–599.
- Levine KE, Levine MA, Weber FX, Hu Y, Perlmutter J, Grohse PM. Determination of mercury in an assortment of dietary supplements using an inexpensive combustion atomic absorption spectrometry technique. J Autom Method Manag 2005; 4: 211–216.
- Melcher J, Olbrich D, Marsh G, Nikiforov V, Gaus C, Gaul S, Vetter W. Tetra- and tribromophenoxyanisoles in marine samples from Oceania. Environ Sci Technol 2005; 39: 7784–7789.
- Teuten EL, Xu L, Reddy CM. Two abundant bioaccumulated halogenated compounds are natural products. Science 2005; 307: 917–920.
- 112. Malmvarn A, Marsh G, Kautsky L, Athanasiadou M, Bergman Å, Asplund L. Hydroxylated and methoxylated brominated diphenyl ethers in the red algae (*Ceramium tenuicorne*) and blue mussels from the Baltic Sea. Environ Sci Technol 2005; 39: 2990–2997.
- 113. Hiebl J, Melcher J, Gundersen H, Schlabach M, Vetter W. Identification and quantification of polybrominated hexahydroxanthene derivatives and other halogenated natural products in commercial fish and other marine samples. J Agric Food Chem 2006; 54: 2652–2657.
- 114. Melcher J, Janussen D, Garson MJ, Hiebl J, Vetter W. Polybrominated hexahydroxanthene derivatives (PBHDs) and other halogenated natural products from the Mediterranean sponge (*Scalarispongia scalaris*) in marine biota. Arch Environ Contam Toxicol 2007; 52: 512–518.
- 115. Titlemier SA. Dietary exposure to a group of naturally-produced organohalogens (halogenated dimethyl bipyrroles) via consumption of fish and seafood. J Agric Food Chem 2004; 52: 2010–2015.
- Haraguchi K, Hisamichi Y, Endo T. Bioaccumulation of naturally-occurring mixed halogenated dimethylbipyrroles in whale and dolphin products on the Japanese market. Arch Environ Contam Toxicol 2006; 51: 135–141.
- Sinkkonen S, Rantalainen AL, Paasivirta J, Lahtipera M. Polybrominated methoxy diphenyl ethers (Meo-PBDEs) in fish and guillemot of Baltic, Atlantic and Arctic environments. Chemosphere 2004; 56: 767–775.
- Marsh G, Athanasiadou M, Bergman A, Asplund L. Identification of hydroxylated and methoxylated polybrominated diphenyl ethers in Baltic Sea salmon (*Salmo salar*) blood. Environ Sci Technol 2004; 38: 10–18.
- 119. Vetter W, von der Recke R, Herzke D, Nygard T. Natural and man-made organobromine compounds in marine biota from Central Norway. Environ Int 2006; 33: 17–26.
- Reddy CM, Xu L, O'Neill GW, Nelson RK, Eglinton TI, Faulkner DJ, Fenical W, Norstrom RJ, Ross P, Tittlemier SA. Radiocarbon evidence for a naturally-produced, bioaccumulating halogenated organic compound. Environ Sci Technol 2004; 38: 1992–1997.
- Reddy CM, Xu L, Eglinton TI, Boon JP, Faulkner DJ. Radiocarbon content of synthetic and natural semivolatile halogenated organic compounds. Environ Pollut 2002; 120: 163–168.
- 122. Tittlemier SA, Borrell A, Duffe J, Duignan PJ, Hall A, Hoekstra P, Kovacs K, Krahn MM, Lebeuf M, Lydersen C, Fair P, Muir D, O'Hara TM, Olsson M, Pranschke JL, Ross P, Stern GA, Tanabe S, Norstrom RJ. Global distribution of halogenated dimethyl bipyrroles in marine mammal blubber. Arch Environ Contam Toxicol 2002; 43: 244–255.
- Tittlemier SA, Simon M, Jarman WM, Elliott JE, Norstrom RJ. Identification of a novel C₁₀H₆N₂Br₄Cl₂ heterocyclic compound in seabird eggs. A bioaccumulating marine natural product?. Environ Sci Technol 1999; 33: 26–33.
- 124. Bakker MI, De Winter-Sorkina R, De Mul A, Boon PE, Van Donkersgoed G, Van Klaveren JD, Baumann BA, Hijman WC, Van Leeuwen SPJ, de Boer J, Zeilmaker MJ. Dietary intake of polybrominated diphenyl ethers in The Netherlands. Organohalogen Compd 2006; 68: 387–390.
- 125. Bocio A, Llobet JM, Domingo JL, Corbella J, Teixido A, Casas C. Polybrominated diphenyl ethers (PBDEs) in foodstuffs: Human exposure through the diet. J Agric Food Chem 2003; 51: 3191–3195.
- 126. Darnerud PO, Atuma S, Aune M, Bjerselius R, Glynn A, Petersson Grawé K, Becker W. Dietary intake estimations of organohalogen contaminants (dioxins, PCB, PBDE and chlorinated pesticides, e.g. DDT) based on Swedish market basket data. Food Chem Toxicol 2006; 44: 1597–1606.

- 127. Darnerud PO, Eriksen GS, Johannesson T, Larsen PB, Viluksela M. Polybrominated diphenyl ethers: Occurrence, dietary exposure, and toxicology. Environ Health Persp 2001; 109(Suppl 1): 49–68.
- Domingo JL. Human exposure to polybrominated diphenyl ethers through the diet. J Chromatogr A 2004; 1054: 321–326.
- Harrad S, Wijesekera R, Hunter S, Halliwell C, Baker R. Preliminary assessment of UK human dietary and inhalation exposure to polybrominated diphenyl ethers. Environ Sci Technol 2004; 38: 2345–2350.
- 130. Kiviranta H, Ovaskainen ML, Vartiainen T. Market basket study on dietary intake of PCDD/Fs, PCBs, and PBDEs in Finland. Environ Int 2004; 30: 923–932.
- Lind Y, Aune M, Atuma S, Becker W, Bjerselius R, Glynn A, Darnerud PO. Food intake of the polybrominated flame retardants PBDEs and HBCD in Sweden. Organohalogen Compd 2002; 58: 181–184.
- 132. Nakagawa R, Ashizuka Y, Hori T, Tobiishi K, Yasutake D, Sasaki K. Determination of brominated flame retardants in fish and market basket food samples of Japan. Organohalogen Compd 2005; 67: 498–501.
- Ryan JJ, Patry B. Body burdens and food exposure in Canada for polybrominated diphenyl ethers (PBDEs). Organohalogen Compd 2001; 51: 226–229.
- 134. Schecter A, Päpke O, Harris TR, Tung KC, Musumba A, Olson J, Birnbaum L. Polybrominated diphenyl ether (PBDE) levels in an expanded market basket survey of US food and estimated PBDE dietary intake by age and sex. Environ Health Persp 2006; 114: 1515–1520.
- 135. Voorspoels S, Covaci A, Neels H. Dietary PBDE intake: A market-basket study in Belgium. Environ Int 2007; 33: 93–97.
- 136. Zennegg M, Kohler M, Gerecke AC, Schmid P. Polybrominated diphenyl ethers in whitefish from Swiss lakes and farmed rainbow trout. Chemosphere 2003; 51: 545–553.
- 137. Akutsu K, Takatori S, Nakazawa H, Hayakawa K, Izumi S, Makino T. Dietary intake estimations of polybrominated diphenyl ethers (PBDEs) based on a total diet study in Osaka, Japan. Food Addit Contam Part B Surveillance 2008; 1: 58–68.
- 138. Harrad S, Wang Y, Sandaradura S, Leeds A. Human dietary intake and excretion of dioxin-like compounds. J Environ Monit 2003; 5: 224–229.
- 139. Turrio-Baldassari L, di Domenico A, Fulgenzi A, Iacovella N, La Rocca C. Dietary intake of PCBs in the Italian population. Organohalogen Compd 1998; 38: 195–198.
- 140. Koizumi A, Yoshinaga T, Harada K, Inoue K, Morikawa A, Muroi J, Inoue S, Eslami B, Fujii S, Fujimine Y, Hachiya N, Koda S, Kusaka Y, Murata K, Nakatsuka H, Omae K, Saito N, Shimbo S, Takenaka K, Takeshita T, Todoriki H, Wada Y, Watanabe T, Ikeda M. Assessment of human exposure to polychlorinated biphenyls and polybrominated diphenyl ethers in Japan using archived samples from the early 1980s and mid-1990s. Environ Res 2005; 99: 31–41.
- 141. Zuccato E, Calvarese S, Mariani G, Mangiapan S, Grasso P, Guzzi A, Fanelli R. Levels, sources and toxicity of polychlorinated biphenyls in the Italian diet. Chemosphere 1999; 38: 2753–2760.
- 142. Wilhelm M, Schrey P, Wittsiepe J, Heinzow B. Dietary intake of persistent organic pollutants (POPs) by German children using duplicate portion sampling. Intern J Hyg Environ Health 2002; 204: 359–371.
- Llobet JM, Bocio A, Domingo JL, Teixido A, Casas C, Muller L. Levels of polychlorinated biphenyls in foods from Catalonia, Spain: Estimated dietary intake. J Food Protect 2003; 66: 479–484.
- Kiviranta H, Ovaskainenn MAL, Vartiainen T. Market basket study on dietary intake of PCDD/Fs, PCBs, and PBDEs in Finland. Environ Int 2004; 30: 923–932.
- 145. Usydus Z, Szlinder-Richert J, Polak-Jusuzak L, Malesa-Ciecwierz M, Dobrzanski Z. Study on the raw fish oil purification from PCDD/F and dl-PCB-industrial tests. Chemosphere 2009; 74: 1495–1501.
- Hilbert G, Lillemark L, Balchen S, Højskov CS. Reduction of organochlorine contaminants from fish oil during refining. Chemosphere 1998; 37: 1241–1252.
- 147. Maes J, De Meulenaer B, Van Heerswynghels P, De Greyt W, Eppe G, De Pauw E, Huyghebaert A. Removal of dioxins and PCB from fish oil by activated carbon and its influence on the nutritional quality of the oil. J Am Oil Chem Soc 2005; 82: 593–597.
- 148. Kawashima A, Iwakiri R, Honda K. Experimental study on the removal of dioxins and coplanar polychlorinated biphenyls (PCBs) from fish oil. J Agric Food Chem 2006; 54: 10294–10299.
- Oterhals Å, Solvang M, Nortvedt R, Berntssen MHG. Optimization of activated carbon-based decontamination of fish oil by response surface methodology. Eur J Lipid Sci Technol 2007; 109: 691–705.
- 150. Kawashima A, Watanabe S, Iwakiri R,, Honda K. Removal of dioxins and dioxin-like PCBs from fish oil by countercurrent supercritical CO₂ extraction and activated carbon treatment. Chemosphere 2009; 75: 788–794.